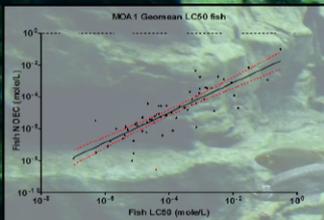


## JRC TECHNICAL REPORTS

# Scientific options for avoiding chronic fish testing on the basis of existing data and extrapolation approaches

Aude Kienzler, Marlies Halder, Andrew Worth

2016



This publication is a Technical report by the Joint Research Centre, the European Commission's in-house science service. It aims to provide evidence-based scientific support to the European policy-making process. The scientific output expressed does not imply a policy position of the European Commission. Neither the European Commission nor any person acting on behalf of the Commission is responsible for the use which might be made of this publication.

**Contact information**

European Union Reference Laboratory for Alternatives to Animal Testing  
Address: Joint Research Centre, Via Enrico Fermi 2749, 21027 Ispra (VA), Italy  
E-mail: [JRC-ECVAM-CONTACT@ec.europa.eu](mailto:JRC-ECVAM-CONTACT@ec.europa.eu)

**JRC Science Hub**

<https://ec.europa.eu/jrc>

JRC101327

EUR 27907

ISBN 978-92-79-58167-0 (PDF)

ISSN 1831-9424 (online)

doi:10.2788/546221 (online)

© European Union, 2016

Reproduction is authorised provided the source is acknowledged.

All images © European Union 2016 cover photo of fish ©edumansur and molecule © 123dartist.

How to cite: Kienzler Aude, Halder Marlies, Worth Andrew; Scientific options for avoiding chronic fish testing on the basis of existing data and extrapolation approaches; EUR 27907; doi:10.2788/546221

## Table of contents

Abstract .....	3
1. Introduction .....	4
2. Regulatory and scientific background .....	6
2.1. REACH requirements and guidance on aquatic toxicity testing .....	6
2.2. Scientific background .....	7
2.2.1. Importance of Mode of Action and mechanistic understanding in environmental risk assessment .....	7
2.2.2. Mode of Action, QSARs and grouping of chemicals .....	7
3. Methodology .....	9
3.1. Extraction of data .....	9
3.1.1. Databases .....	9
3.1.2. Species .....	9
3.1.3. Endpoint and duration of exposure .....	9
3.1.4. Quality check .....	9
3.2. Treatment of data prior to analysis .....	9
3.2.1. Lowest value and geometric mean approaches .....	9
3.2.2. Assignment of Modes of Action .....	10
3.3. Analysis .....	10
4. Results and discussion .....	11
4.1. Interspecies extrapolation (quantitative activity-activity relationships) .....	11
4.1.1. Daphnid to chronic fish relationships – irrespective of the MOA .....	11
4.1.1.1. Relationships .....	11
4.1.1.2. Discussion of outliers .....	13
4.1.1.3. Interspecies sensitivity ratio .....	14
4.1.2. Daphnid to chronic fish relationships by MOA .....	18
4.1.2.1. Relationships .....	18
4.1.2.2. Outliers .....	20
4.1.2.3. Sensitivity ratios .....	21
4.2. Acute to chronic fish data .....	21
4.2.1. Acute to chronic fish relationships – irrespective of the MOA .....	21
4.2.1.1. Relationships .....	21
4.2.1.2. Discussion of outliers .....	23
4.2.1.3. Acute to chronic ratio .....	23
4.2.2. Acute to chronic fish relationship by MOA .....	28
4.2.2.1. Relationships .....	28
4.2.2.2. Discussion of outliers .....	28
4.2.2.3. Acute to chronic ratio and MOA .....	30

5. Conclusions and recommendations .....	31
References .....	33
List of abbreviations and definitions.....	38
List of figures.....	39
List of tables.....	40
Appendix I. Examples of EU regulatory information requirements for <i>Daphnia</i> and/or fish toxicity.....	41

## Abstract

The assessment of aquatic toxicity is an important component of the environmental hazard and risk assessment of all types of chemicals, and is therefore included in several pieces of EU chemicals legislation. Aquatic toxicity refers to the effects of chemicals on organisms living in the water and is usually determined by testing on organisms representing three trophic levels, i.e. plants (or algae), invertebrates (crustaceans such as *Daphnia* spp.) and vertebrates (fish). Whereas acute aquatic toxicity testing is a basic requirement in most pieces of EU chemicals legislation, chronic aquatic toxicity testing may be required when the outcome of the acute testing indicates a risk, or in the case that long-term exposure is expected. EU chemicals legislation encourages the use of all available information for hazard and risk assessment before new tests on vertebrates are proposed or conducted. In this context, scientific options for avoiding chronic fish testing on the basis of existing data and extrapolation approaches have been explored.

For the purposes of this work, data on acute and chronic aquatic toxicity (*Daphnia* and fish) from several databases (US EPA Ecotox database, Aquatic ECETOC, Aquatic OASIS, Aquatic Japan MoE databases and ECHA database as implemented in the OECD QSAR Toolbox Version 2.3) were collated and analysed. Simple linear relationships and interspecies sensitivity ratios were calculated using either acute *Daphnia* data (48h LC<sub>50</sub>) or chronic *Daphnia* data (14 days NOEC) and chronic fish data (>21 days NOEC). Acute to chronic relationships and acute to chronic ratios (ACR) were also calculated based on acute fish data (96h LC<sub>50</sub>) and chronic fish data. These analyses were carried out on the whole set of chemicals and on subgroups of chemicals classified according to the Verhaar mode of action (MOA) scheme, which attribute general mode of acute aquatic toxic action based on the chemical structure of the molecule. Outliers were identified applying the Robust regression and Outlier removal (ROUT) method.

Our results show that the best fitted relationships for the prediction of chronic fish toxicity are obtained based on acute fish data ( $r^2=0.87$ ) and acute *Daphnia* data ( $r^2=0.64$ ) when dealing with the whole set of chemicals regardless of the MOA. The quality of the relationships was increased by using the geometric mean (calculated across all the values extracted for a given chemical and a given endpoint) instead of the lowest value for a given endpoint.

When considering the MOA, MOA 3 and MOA 1 chemicals give the strongest acute *Daphnia* to chronic fish relationship and chronic *Daphnia* to chronic fish relationship; however the relationships obtained with acute *Daphnia* data are better ( $r^2= 0.83$  and  $0.69$  for MOA 3 and MOA 1 respectively) than the one obtained with chronic *Daphnia* data ( $r^2= 0.66$  and  $0.65$  for MOA 1 and 3 respectively). When considering acute fish data, all the MOA classes give strong relationships ( $r^2=0.88$  for MOA 3 and MOA 5 chemicals,  $0.85$  for MOA 4 chemicals and  $0.83$  for MOA 1 and MOA 2 chemicals). Therefore when acute toxicity data on fish are available, they might give a reliable basis to extrapolate the chronic toxicity on fish as a first tier assessment or within a weight of evidence approach.

There is a correlation between chemicals with high ACR values or interspecies sensitivity ratios and the outliers identified in the above-mentioned relationships. When considering chemicals with a high interspecies sensitivity ratio, *Daphnia* being more sensitive than fish, several aniline derivatives and pesticides acting through cholinesterase inhibition were identified. When considering high interspecies sensitivity ratio chemicals for which *Daphnia* is less sensitive than fish, we found pesticides and known endocrine disruptors such as ethynil oestradiol and 17 $\beta$ -oestradiol. Extreme (i.e. <1 or > 100) interspecies sensitivity ratios were mainly evident for MOA 2, 4 and 5 chemicals. Regarding ACR for fish, around 50% of the chemicals in each MOA class have an ACR within a factor of 10; whereas 100% of MOA 3, 90.9% of MOA 2, 88.3% of MOA 4 and 85.5% of MOA 1 chemicals have an ACR within a factor of 100. Therefore, the safety factor of 100 commonly applied in environmental risk assessment does not seem to be equally protective for every MOA.

## 1. Introduction

The assessment of aquatic toxicity is an important component of the environmental hazard and risk assessment of all types of chemicals, and is therefore included in several pieces of EU chemicals legislation. These include the Regulation concerning the Registration, Evaluation, Authorisation and Restriction of Chemicals (REACH, EC, 2006), the Biocidal Products Regulation (EU, 2012), the Plant Protection Products Regulation (EC, 2009a) and data requirements (EU, 2013a, 2013b), pharmaceuticals (EMA, 2006, 2004), feed additives (EC, 2008a; EFSA, 2008) and others. In addition, the Cosmetics Regulation (EC, 2009b) states that environmental concerns of cosmetic ingredients and products should be addressed through REACH. Information requirements according to the different regulations are summarised in Annex I.

Aquatic toxicity refers to the effects of chemicals on organisms living in the water and is usually determined by testing on organisms representing the three trophic levels, i.e. plants (or algae), invertebrates (crustaceans such as *Daphnia* spp.) and vertebrates (fish). The information on aquatic toxicity is used for classification and labelling (EC, 2008b), the derivation of Predicted No Effect Concentration (PNEC) values for use in risk assessment, and for the assessment of PBT substances. In general, the lowest of the available toxicity values is used to define the hazard category, derive the PNEC or "Toxicity" criterion of the different trophic levels (fish, crustacean, algae or aquatic plants): 50% effective concentration [ $EC_{50}$ ] or 50% lethal concentration [ $LC_{50}$ ] for acute aquatic toxicity; x% effective concentration [ $EC_x$ ], Lowest Observed Effect Concentration [LOEC], No Observed Effect Concentration [NOEC] for chronic aquatic toxicity.

Whereas acute aquatic toxicity testing is a basic requirement in most pieces of EU chemicals legislation, chronic aquatic toxicity testing may be required when the outcome of the acute testing indicates a risk, or in the case that long-term exposure is expected.

Regulatory aquatic risk assessment schemes require toxicity testing of chemicals on a limited number of laboratory species; thus, extrapolation from the obtained toxic responses to all species representing that trophic level in the environment is a fundamental tenet of regulatory risk assessment. To derive the PNEC for aquatic toxicity, safety factors are applied to the laboratory data. These factors are intended to account for interspecies differences in sensitivity, extrapolation from acute to chronic effects, the physicochemical complexity of natural water versus laboratory test media, and the complexity of the ecosystem versus single species laboratory tests. The choice of the factor (10, 100 or 1000) depends on the quality and quantity of the available data.

Moreover, differences in species sensitivity regarding acute aquatic toxicity have been well described (Hoekzema et al., 2006; Hutchinson et al., 2003; Jeram et al., 2005; Tebby et al., 2011; Weyers et al., 2000). To address the question of whether it is possible to predict acute toxicity in fish from non-vertebrate species, Netzeva et al. (2007) reviewed several Quantitative Activity-Activity Relationships between species, the most relevant and reliable relationship for acute fish toxicity being between *Daphnia* and rainbow trout ( $n=360$ ), with an  $r^2$  value of 0.67. More recent studies have confirmed the good correlation between acute fish and *Daphnia* toxicity data, especially for organothiophosphates ( $0.74 < r^2 < 0.94$ ) and (benzo)triazoles ( $r^2=0.87$ ) (Cassani et al., 2013; Kar and Roy, 2010; Zhang et al., 2010; Zvinavashe et al., 2009). The correlation depends on both the bio-uptake process and the MOA of the chemical (Zhang et al., 2010), as well as its physicochemical properties (Tebby et al., 2011). The US EPA has also developed a specific tool, the Interspecies Correlation Estimation (ICE) tool (implemented as WebICE; Raimondo et al., 2010) to predict acute toxicity to three relevant fish species (fathead minnow, rainbow trout, and common carp) on the basis of *Daphnia* toxicity, with the strongest correlation being evident between *Daphnia* and rainbow trout ( $r^2=0.51$ ). However, there is little work on the potential to predict chronic fish toxicity from *Daphnia* data.

Therefore, and in the light of the EU Directive on the protection of animals used for scientific purposes (EU, 2010) and the *EURL ECVAM strategy to replace, reduce and refine the use of fish in aquatic toxicity and bioaccumulation testing* (EURL ECVAM, 2014) we have been exploring whether interspecies extrapolations and acute to chronic relationships can be used for supporting the waiving of chronic fish tests. For this purpose, data (LC<sub>50</sub>, NOEC) for *Daphnia* and fish have been extracted from various databases and analysed to identify possible relationships taking into consideration different mode of actions. We discuss these approaches in particular in the context of REACH since the REACH regulation states in §25 that testing on vertebrate animals shall be undertaken only as a last resort (EC, 2006).

## 2. Regulatory and scientific background

### 2.1. REACH requirements and guidance on aquatic toxicity testing

Chronic or long-term aquatic toxicity testing is required for chemicals covered under REACH Annexes VIII-X (10-100, 100-1000, >1000 tonnes per year), *if the chemical safety assessment according to Annex I indicates the need to investigate further the effects on aquatic organisms*. A potential environmental risk would be evident if the Predicted No Effect Concentration / Predicted Environmental Concentration ratio (PNEC/PEC) is greater than 1<sup>1</sup> (EC, 2006; ECHA, 2012) (see Appendix I).

REACH further states that *The choice of the appropriate test(s) depends on the results of the chemical safety assessment* (EC, 2006). The appropriate tests listed are long-term testing on invertebrates, preferable *Daphnia* (TG211; OECD, 2012) and fish, e.g. the Fish Early-Life Stage (FELS) Toxicity Test (TG210; OECD, 2013), the Fish Short-term Toxicity Test on Embryo and Sac-Fry Stages (TG212; OECD, 1998), and the Fish Juvenile Growth Test (TG215; OECD, 2000).

Independent of the tonnage level, long-term aquatic toxicity testing on invertebrates or fish may be preferred to acute aquatic tests if a substance is poorly water soluble.

The ECHA Endpoint Specific Guidance on Aquatic toxicity (ECHA, 2012) details on how the information requirements on aquatic toxicity for the purpose of REACH can be met. If information on long-term aquatic toxicity is needed but a NOEC value from long-term toxicity testing on fish is not available, several options can be considered:

1. The use of other available long-term toxicity data: Standard testing data might be substituted by other reliable experimental data on aquatic invertebrates or fish (e.g. data from non-standard studies or non-standard organisms).
2. The use of reliable QSAR results, although this document also states that "currently reliable QSAR models for chronic toxicity are rare and thus reliable QSAR results will be seldom available".
3. Reliable read-across from available experimental data on a structurally related substance.

When considering long-term testing in fish, the relative sensitivity of fish and other species should also be compared (ECHA, 2012). If there is "compelling evidence" to suggest that the fish value is likely to be at least a factor of about 10 less sensitive than invertebrates or algae, there are no further requirements for fish testing. If invertebrates are likely to be more sensitive than fish and algae then a testing proposal for a long-term toxicity study on *Daphnia* (OECD TG211) should be submitted to ECHA.

If fish are likely to be more sensitive than invertebrates and algae or the relative sensitivity of fish cannot be predicted, a testing proposal for a long-term study on fish should be submitted; alternatively, risk management measures reducing exposure and hence risk sufficiently might be considered (ECHA, 2012).

---

<sup>1</sup> i.e. if the LC<sub>50</sub> of the short-term fish test divided by 1000 is greater than the Predicted Environmental Concentration (PEC)

## 2.2. Scientific background

### 2.2.1. Importance of Mode of Action and mechanistic understanding in environmental risk assessment

Extrapolation has always been a key component of criteria setting and risk assessment, and became important in both the quantitative and qualitative relationships between effect measures and assessment endpoints (Solomon et al., 2008). Acute to Chronic Ratios (ACRs), which are empirically derived from a toxicological database, provide simple rules-of-thumb for extrapolating from acute to chronic effects. Being applicable to a broad range of substances, ACRs justify the application of standard assessment factors in the risk assessment process. Therefore, those practices allows the uses of already existing data on a chemical toxicity for a given endpoints and/or species to predict its potential toxicity to another given endpoints/species.

The Mode of Action (MOA) describes the understanding of selected key events that lead to toxic effects, whereas the mechanism of action refers to a comprehensive understanding of the entire sequence of events that results in toxicity (ECETOC, 2006). One complexity of environmental risk assessment is that it should address all the various components of the ecosystem, i.e. all the species with their respective sensitivity to the chemical. To achieve this, the most sensitive species is usually chosen, with the hypothesis that if the most sensitive species is protected, the other species would be protected as well.

In the context of a testing strategy, the mechanistic understanding could allow testing the chemical only on the most sensitive species when there is a shared MOA, i.e. *Daphnia* toxicity tests could be preferred to fish toxicity tests for ion channel mediated neurotoxicity of pyrethroids, knowing that *Daphnia* will likely be amongst the most sensitive aquatic organisms although the target protein is found in crustacean, fish, insects and molluscs. In contrast, a very different MOA specific to vertebrates (i.e. receptor mediated feminisation of fish by steroidal oestrogens) would indicate fish testing (ECETOC, 2007). For the same reason, interspecies sensitivity factors can also greatly depend on the MOA. For instance, the acute "base set" (*Daphnia*, fish and algae) interspecies sensitivity factor for cypermethrin was found to be nearly 463000, a value much higher than typical for MOA 1-3 chemicals, as the target protein is not found in algae.

### 2.2.2. Mode of Action, QSARs and grouping of chemicals

In order to develop and apply QSARs for specific MOAs, several attempts have been made to develop structure-based classification schemes. Based on Verhaar et al. (1992) and Enoch et al. (2008), the following classification scheme was proposed (Table 2.1):

**Table 2.1: Verhaar Classification Scheme for aquatic modes of action (MOA).**

Class 1	chemicals acting by nonpolar narcosis (or baseline toxicity)
Class 2	chemicals acting by polar narcosis (including (substituted) phenols and (substituted) anilines)
Class 3	chemicals with non-specific reactivity (including aldehydes and epoxides)
Class 4	chemicals with specific reactivity (including pesticides, polychlorinated biphenyls and some polycyclic aromatic hydrocarbon metabolites)
Class 5	all the other chemicals that have not been classify in the previous class

Strictly, the fifth class is not a distinct class, but rather a set of miscellaneous chemicals not covered by MOAs 1-4. It has to be noted that this scheme is based on acute toxicity.

Russom et al. (1997) further extended the Verhaar classification scheme and distinguished eight modes of toxic action: base-line narcosis or narcosis I, polar narcosis or narcosis II, ester narcosis or narcosis III, uncoupling of oxidative phosphorylation, respiratory inhibition, electrophile/proelectrophile reactivity, acetylcholinesterase inhibition, and central nervous system seizure.

QSAR models can be used for the prioritisation of chemicals (Salvito et al., 2002; Sanderson et al., 2004) and are useful in cases where data availability is incomplete for a test or when there are differences in test methods. In practice, QSAR predictions for acute aquatic toxicity and bioconcentration in fish are commonly used and widely accepted for regulatory purposes. Models for acute aquatic toxicity are especially useful for chemicals with the least reactive acute MOA (MOA 1 and MOA 2); highly reactive chemicals are more problematic. Netzeva et al. (2007) summarised the features of several fish acute toxicity QSARs and found that  $r^2$  values for most narcosis QSARs were around 0.9. The evidence for the predictive power of acute fish baseline or narcotic toxicity QSARs is therefore strong, and acute ecotoxicity QSARs for non-polar and polar narcotic compounds have been used frequently over the last decades. However, there are few examples of QSARs for chronic fish toxicity, with the exception of those developed for endocrine disrupting chemicals (Grindon et al., 2006). For instance, chronic fish toxicity models have been developed for predicting chronic narcosis to fish (Claeys et al., 2013) and sub-lethal NOEC values for non-polar narcotics (Austin and Eadsforth, 2014).

Several freely available and user-friendly tools and information sources are available, such as the OECD QSAR Application Toolbox, the JRC QSAR models database ([http://ihcp.jrc.ec.europa.eu/facilities/JRC\\_QSAR\\_Model\\_Database.htm](http://ihcp.jrc.ec.europa.eu/facilities/JRC_QSAR_Model_Database.htm)), Toxtree (<http://sourceforge.net/projects/toxtree/>), and US EPA's ECOSAR (<http://www.epa.gov/tsca-screening-tools/ecological-structure-activity-relationships-ecosar-predictive-model>). This tool, which aims at estimating short-term and chronic toxicity to aquatic organisms, including fish, has been shown to be reliable for neutral organics (de Haas et al., 2011). However, its reliability is not satisfactory if used with chlorinated anilines, although those are polar narcotics (Dom et al., 2012), or if simply used as a black box, which is widely done in the absence of clear guidance (de Roode et al., 2006).

## 3. Methodology

### 3.1. Extraction of data

#### 3.1.1. Databases

In a first step, organic chemicals with chronic fish toxicity data were selected, and then data sets including acute fish toxicity data as well as acute and chronic toxicity data on *Daphnia* extracted from the Aquatic ECETOC, the Aquatic OASIS, the Aquatic Japan MoE databases, the ECHA database and the US EPA database, using the OECD QSAR Toolbox 2.3<sup>2</sup>.

#### 3.1.2. Species

*Daphnia (D.) magna* was the only species retained for *Daphnia* spp, while for fish preference was given to OECD freshwater fish species (zebrafish [*Danio rerio*], fathead minnow [*Pimephales promelas*], common carp [*Cyprinus carpio*], Japanese medaka [*Oryzias latipes*], guppy [*Poecilia reticulata*], bluegill [*Lepomis macrochirus*], rainbow trout [*Onchorhynchus mykiss*], three-spined stickleback [*Gasterosteus aculeatus*]) as well as the saltwater species sheepshead minnow (*Cyprinodon variegatus*). Data from other species of freshwater fish or from saltwater or estuarine species were considered only if there were no data available on the preferred species.

#### 3.1.3. Endpoint and duration of exposure

The LC<sub>50</sub> was chosen as the endpoint for acute toxicity tests. The duration of acute toxicity tests is typically 48h for *Daphnia* and 96h for fish. The acute effects taken into account in this study were immobilisation and mortality for *Daphnia*, and mortality for fish.

The NOEC was chosen as the endpoint for long-term toxicity tests. The minimum duration of exposure considered was 21 days for *Daphnia*; and 21, 28 or more days for fish. The chronic effects taken into account were mortality, reproduction, growth and development.

#### 3.1.4. Quality check

Data were checked for duplicates (i.e. if the same data were extracted from several databases), and it was checked that the toxicological value was below the solubility limits of the substances. Only data ranked Klimisch score 1 or 2 from the ECHA database were considered.

### 3.2. Treatment of data prior to analysis

#### 3.2.1. Lowest value and geometric mean approaches

If there was more than one value per endpoint of interest for a given chemical, either the lowest toxicological value was identified or the geometric mean was calculated for the given endpoint and used for the analysis. The use of the lowest value is often recommended in environmental risk assessment in order to favour a conservative approach; however when it comes to conclude on a relationship, the use of the geometric mean, by giving less weight to the extreme value, can improve the quality of the relationship.

---

<sup>2</sup> The extraction was made in July 2014 for the Aquatic ECETOC, the Aquatic OASIS, the Aquatic Japan MoE databases, and the US EPA database.

Both approaches were applied when analysing the whole set of chemicals (i.e. irrespective of the MOA), whereas only the geometric mean approach was applied to the chemicals subdivided according to their MOA (see below).

### **3.2.2. Assignment of Modes of Action**

The selected chemicals were profiled with the QSAR Toolbox and classified into five MOA classes according to the Verhaar classification scheme (see Table 2.1). To complete the classification they were further profiled with Toxtree and the post-processing filter encoded as a KNIME workflow (Ellison et al., 2015) that has recently been developed in order to improve the predictivity of the Toxtree scheme. In case of conflicting results or MOA 5 chemicals, other data from literature has been used to attribute a final MOA (Barron et al., 2015; ECETOC, 2007; Russom et al., 1997).

### **3.3. Analysis**

A simple linear regression on log10-transformed data was used (GraphPad Prism 5) to obtain the interspecies relationships and the corresponding  $r^2$  values (measures of goodness-of-fit) for the whole set of chemicals or subsets of chemicals according to their MOA. The Robust regression and Outlier removal (ROUT) method to identify potential outliers was used (Motulsky and Brown, 2006); the  $r^2$  given are the  $r^2$  after elimination of the outlier(s). For the interpretation of regression data it was assumed that a regression coefficient  $r^2$  of  $>0.6$  corresponds to a moderate correlation whereas a value of about 0.8 and more  $r^2$  corresponds to a reliable prediction.

The interspecies sensitivity ratio for each chemical ( $SR_{acute} = LC_{50Daphnia}/NOEC_{Fish}$ ;  $SR_{ch} = NOEC_{Daphnia}/NOEC_{Fish}$ ) and the acute to chronic ratio for fish ( $ACR_{fish}$ ) were also calculated to compare the respective sensitivity of fish and *Daphnia*. The statistical result was characterised by the median and 90%-ile value as well as by their minimum and maximum values. The distribution of the sensitivity ratio ( $<10$ ;  $10 < <100$ ;  $100 < <1000$ ;  $>1000$ ) were also analysed, taking the MOA into account.

## 4. Results and discussion

This chapter describes our own derivation of interspecies extrapolation and interspecies sensitivity ratio in 4.1; and focuses on acute to chronic relationship based on fish toxicity data and ACRs for fish in 4.2. We further discuss possible reasons for outliers.

### 4.1. Interspecies extrapolation (quantitative activity-activity relationships)

Chronic fish toxicity data have been compared with acute and chronic *Daphnia* data, in order to better characterise the relationship existing between those data. The ratio sensitivity has also been calculated for each chemical to compare the respective sensitivity of the two species.

#### 4.1.1. *Daphnid to chronic fish relationships – irrespective of the MOA*

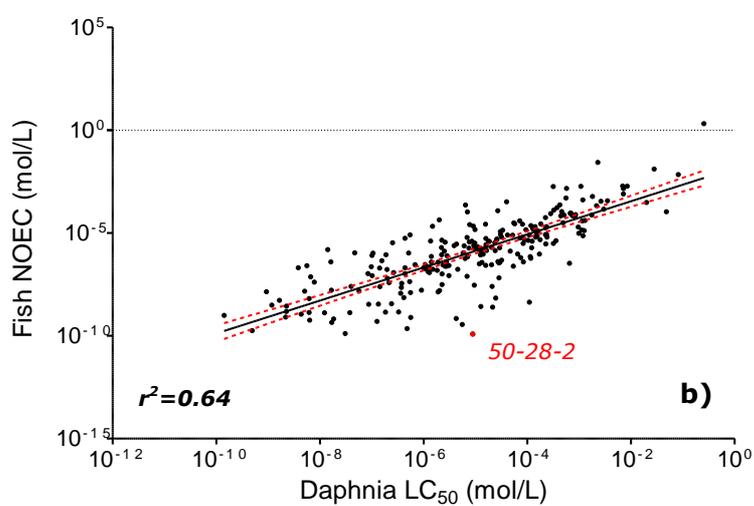
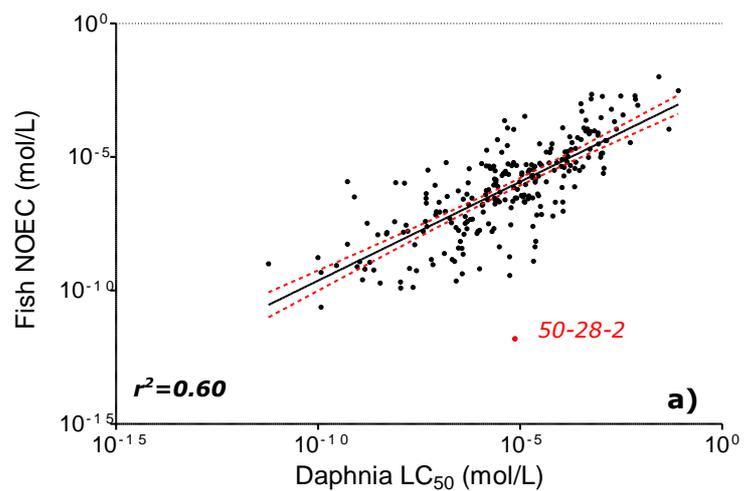
##### 4.1.1.1. Relationships

There were 246 substances with matched *Daphnia* acute data (LC<sub>50</sub> 48h) and chronic fish data (NOEC >21 days); and 193 substances with matched *Daphnia* chronic data (NOEC - 21 days), and chronic fish data. There was a significant relationship between *Daphnia* data and chronic fish toxicity data with both approaches ( $p < 0.0001$ ). However, in both cases (acute *Daphnia* and chronic *Daphnia* data), the goodness-of-fit was improved by using geometric means instead of the lowest values ( $r^2$  of logged data = 0.64 and 0.61 respectively for acute *Daphnia* data; 0.59 and 0.57 respectively for the chronic *Daphnia* data) (see

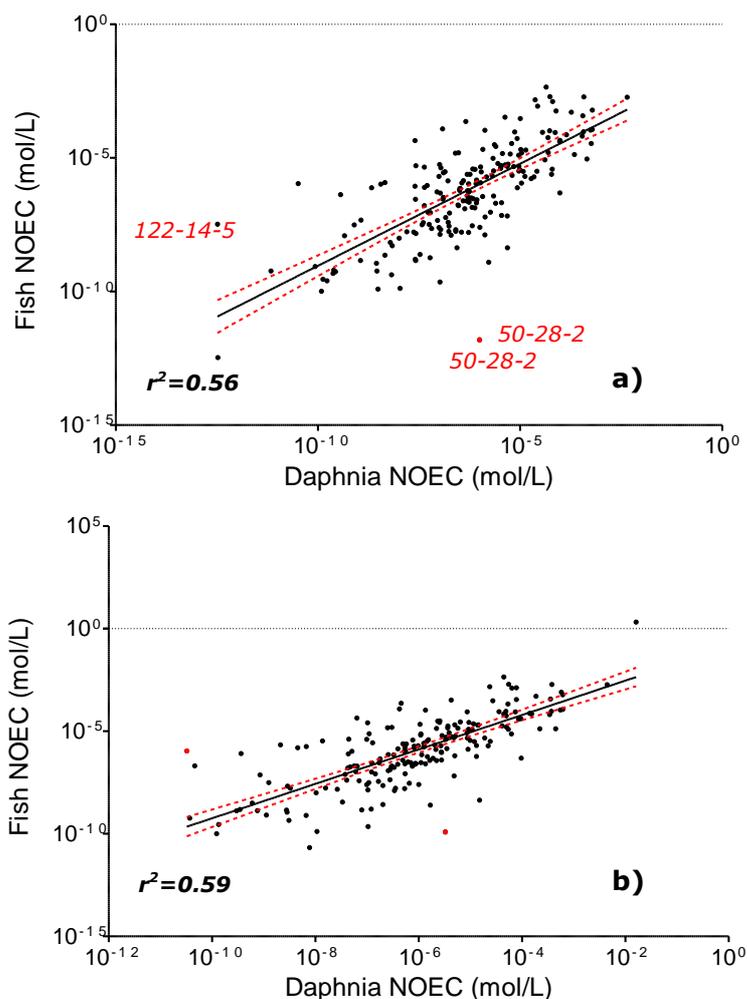
Table 4.1). Data are plotted in Figure 4.1 and Figure 4.2.

**Table 4.1: Linear relationships between *Daphnia* acute or chronic toxicity data and fish chronic toxicity data.**

	<b>Lowest value</b>	<b>Geometric mean</b>
<b>LC<sub>50</sub> <i>Daphnia</i></b>		
<b><i>n</i></b>	246	
<b><i>p</i> value</b>	< 0.0001	< 0.0001
<b><i>r</i><sup>2</sup></b>	0.60	0.64
<b>Outlier</b>	1	1
	50-28-2	50-28-2
<b>NOEC <i>Daphnia</i></b>		
<b><i>n</i></b>	193	
<b><i>p</i> value</b>	< 0.0001	< 0.0001
<b><i>r</i><sup>2</sup></b>	0.56	0.59
<b>Outlier</b>	1	2
	50-28-2	122-14-5; 50-28-2



**Figure 4.1: Relationships between *Daphnia* 48h LC<sub>50</sub> and fish NOEC values from the OECD QSAR Toolbox 2.3 (n=246). a: Lowest value approach, b: Geometric mean approach. The dashed lines represent the 95% confidence interval. *Italics*: CAS number of the outliers.**



**Figure 4.2: Relationships between *Daphnia* 21-days NOEC and fish NOEC values from the OECD QSAR Toolbox 2.3 (n=193). a: Lowest value approach, b: Geometric mean approach. The dashed lines represent the 95% confidence interval. *Italics*: CAS number of the outliers.**

#### 4.1.1.2. Discussion of outliers

Regarding acute *Daphnia* to chronic fish relationships, only 17 $\beta$ -oestradiol is identified as outlier, with both approaches (Figure 4.1). Regarding chronic *Daphnia* data to chronic fish relationship, 17 $\beta$ -oestradiol is also identified as an outlier with both approaches and a second chemical, fenitrothion, is identified with the geometric mean approach (.

Table 4.2).

**Table 4.2: Name and MOA of outliers**

<b>Lowest value approach</b>	<b>Geometric mean</b>	<b>Name of chemical</b>	<b>MOA</b>
<b>Acute <i>Daphnia</i> data</b>			
50-28-2	50-28-2	17 $\beta$ -oestradiol	2
<b>Chronic <i>Daphnia</i> data</b>			
50-28-2	50-28-2	17 $\beta$ -oestradiol	2
	122-14-5	Fenitrothion	4

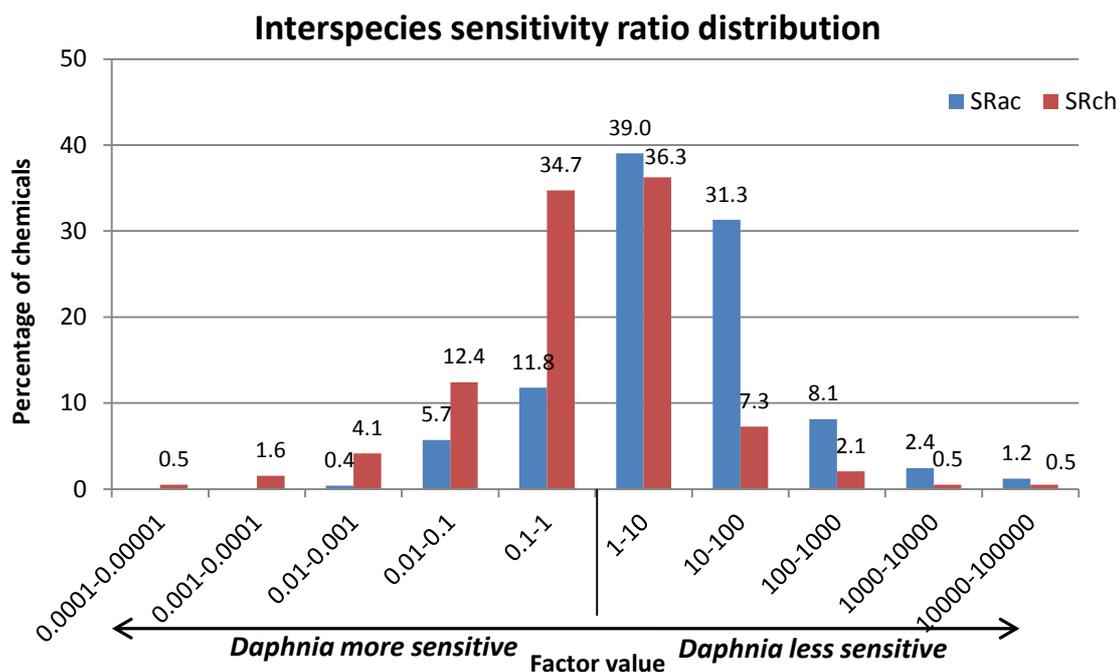
17 $\beta$ -oestradiol is a steroid and an oestrogen sex hormone found in most vertebrates as well as many crustaceans, insects, fish, and other animal species. It is a well-studied chemical characterised by a high quantity of data in the various databases (118 entries after extraction, 42 entries in our final database). The NOEC fish values reported display a high variability and range between 0.00001 and 0.001. This can be due, amongst other factors, to the fact that the guidelines used for chronic fish tests cover different life stages. The impact of this variability is decreased by the use of the geometric mean approach, which gives less weight to extreme values. Nevertheless, 17 $\beta$ -oestradiol remains an outlier due to its high toxicity to fish. Although being classified as MOA 2, 17 $\beta$ -oestradiol acts primarily as an agonist of the oestrogen receptor (ER). The result of ER activation is a modulation of gene transcription and expression in ER-expressing cells which is the predominant mechanism by which 17 $\beta$ -oestradiol mediates its biological effects. However, it also acts as an agonist of membrane oestrogen receptors via which it can mediate a variety of rapid, non-genomic effects (Pinto et al., 2014). Therefore, 17 $\beta$ -oestradiol acts by a rather specific and non-narcotic MOA, at least when it comes to chronic toxicity; however, this is not captured in the Verhaar scheme being a classification scheme based on acute toxicity.

Fenitrothion is a phosphorothioate insecticide acting through acetyl cholinesterase inhibition. It appears as one of the most chronically toxic chemicals to *Daphnia* in our database, whilst having a medium toxicity to fish. However, there was only one *Daphnia* NOEC value available.

#### 4.1.1.3. Interspecies sensitivity ratio

The sensitivity ratios of the chemicals have been calculated and their distribution is shown in Figure 4.3 ( $SR_{ac} = LC_{50Daphnia} / NOEC_{Fish}$ ;  $SR_{ch} = NOEC_{Daphnia} / NOEC_{Fish}$ ). A ratio of >1 indicates that *Daphnia* are less sensitive compared to fish and a ratio of <1 indicates that *Daphnia* are more sensitive compared to fish. Quite expectedly, acute *Daphnia*  $LC_{50}$  values are generally higher than chronic fish NOEC values (81.6% of the chemicals), and usually within a factor 10 for 39% of the latter chemicals.

When using *Daphnia* NOEC values, the distribution is shifted to the left by a factor of 10. For 69.9% of the chemicals, the  $SR_{ch}$  is within a factor of 10; amongst which *Daphnia* is less sensitive for 35.7% of the chemicals, and more sensitive for 34.5% of the chemicals. As a whole, *Daphnia* is more sensitive for 53% of the chemicals, whereas fish is more sensitive for 47% of the chemicals. Therefore, none of these two trophic levels (i.e. fish and daphnids) seems to be more sensitive than the other in chronic testing, which is in line with some previous work (May and Hahn, 2014).



**Figure 4.3: Sensitivity ratio derived with acute *Daphnia* data versus chronic fish data (blue bars) and chronic *Daphnia* data versus chronic fish data (red bars) in % of chemicals of the whole dataset; i.e. 34.7% of the chemicals demonstrates a higher chronic toxicity in *Daphnia* compared to NOEC fish and values range within a factor of 10; for 39% of the chemicals, the acute *Daphnia* LC<sub>50</sub> indicate a lower sensitivity compared to chronic NOEC fish and values range within a factor of 10.**

In 90.7% of the cases, the chronic SR<sub>ch</sub> is within a factor 100. For the remaining 9.3% corresponding to 19 chemicals, (see Table 4.3) a high SR<sub>ch</sub> was calculated.

These 19 chemicals are representing pharmaceuticals, pesticides, aromatic amines and hormones. With the exception of ibuprofen being classified as MOA 1, they are mostly specifically acting chemicals (seven MOA 4) and polar narcotics (seven MOA 2). One is classified as MOA 3 and two are classified as MOA 5.

Although the exact mechanism of action of ibuprofen is not known, this nonsteroidal anti-inflammatory pharmaceutical is a nonselective inhibitor of cyclooxygenase, an enzyme involved in prostaglandin synthesis via the arachidonic acid pathway. Its pharmacological effects are believed to be due to inhibition of cyclooxygenase-2 (COX-2) which decreases the synthesis of prostaglandins involved in mediating inflammation, pain, fever, and swelling<sup>3</sup>. Norethindrom is a progestin hormone, whereas ethinyl oestradiol and 17β-oestradiol are oestrogen hormones.

<sup>3</sup> <http://www.drugbank.ca/drugs/DB01050#>, version 4.3 (22/06/2015), retrieved 24/02/2016

**Table 4.3: Substances with a high SRch**

		<b>CAS number</b>	<b>Name</b>	<b>MOA</b>	<b>Ratio</b>
<b>Daphnia more sensitive</b>	<b>0.00001/0.0001</b>	122-14-5	Fenitrothion	4	0.00003
	<b>0.0001/0.001</b>	333-41-5	Diazinon	4	0.000224
		298-04-4	Disulfoton	4	0.000443
		298-46-4	Carbamazepine	5	0.000978
	<b>0.001/0.01</b>	108-42-9	3-chloroaniline	2	0.001455
		591-27-5	3-amino-phenol	2	0.002
		298-00-0	Methyl parathion	4	0.002919
		95-53-4	o-methylaniline	2	0.003508
		624-92-0	Dimethyl disulphide	3	0.003757
		106-49-0	p-methylaniline	2	0.003939
		63-25-2	Carbaryl	4	0.004048
		62-53-3	Aniline	2	0.007719
	<b>Daphnia less sensitive</b>	<b>100/1000</b>	15687-27-1	Ibuprofen	1
57-63-6			Ethinyl oestradiol	2	359
1897-45-6			Chlorothalonil	4	466
68-22-4			Norethindrone	5	670
<b>1000/10000</b>		138261-41-3	Imidacloprid	4	3505
<b>10000/100000</b>		50-28-2	17 $\beta$ -oestradiol	2	26912

Amongst other chemicals for which *Daphnia* is far less sensitive than fish, is chlorothalonil, a broad spectrum non-systemic fungicide also used as a wood protectant, pesticide, acaricide, and to control mold, mildew, bacteria and algae. It acts via the formation of substituted chlorothalonil-reduced glutathione derivatives in cells, which trigger inhibition of specific NAD thiol-dependent glycolytic and respiratory enzymes (Tillman et al., 1973). This action on basic glycolytic and respiratory pathways is conserved across biological taxa and toxicity is evident in multiple organisms.

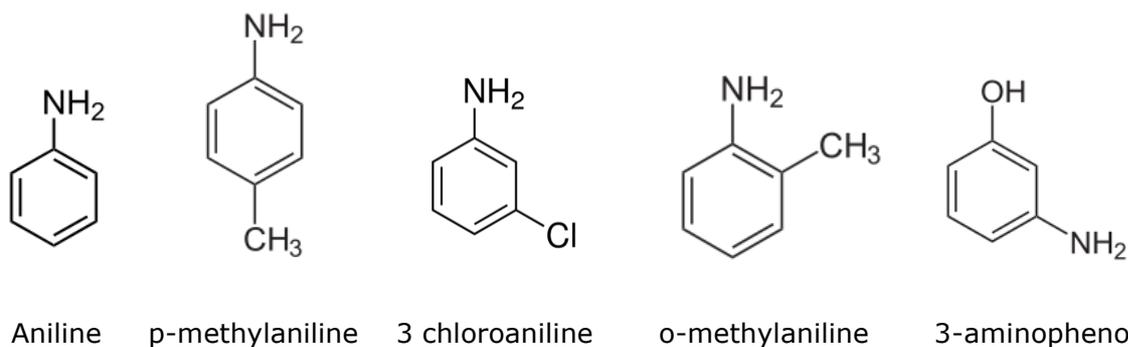
Imidacloprid is a neonicotinoid, a systemic insecticide which acts as an insect neurotoxin, by blocking nicotinic acetylcholine receptors and preventing acetylcholine from transmitting impulses between nerves, which result in the insect's paralysis and eventual death. Quite unexpectedly, it appears to be much more toxic to fish than to *Daphnia*. There was one entry for chronic fish toxicity in the database, from the general literature. When going back to the original study, it appears that the NOEC corresponds to the concentration measured in the field when assessing the chronic endpoint 27 days after contamination of the field (1.1  $\mu\text{g/L}$ ) (Sanchez-Bayo and Goka, 2005). As there was no effect, this quite low concentration appears as a NOEC, although other sources mention a much higher fish NOEC, i.e. the EFSA conclusion on imidacloprid retains a chronic NOEC value of 9.02 mg/L (EFSA, 2014). This means that this high chronic sensitivity ratio is probably methodologically biased.

Regarding chemicals for which *Daphnia* is much more sensitive than fish, we found mainly pesticides acting through acetylcholinesterase inhibition and aromatic amines. Diazinon, disulfoton and methylparathion are organophosphate insecticides, whilst

carbaryl is of the carbamate family, and fenitrothion is a phosphorothioate. All of them act through acetylcholinesterase inhibition.

Carbamazepine is a pharmaceutical, an anticonvulsant, which works by decreasing nerve impulses that cause seizures and pain.

Finally, aniline and its derivatives, p-methylaniline, 3-chloroaniline, 2-methylaniline and 3-amino-phenol show chronic sensitivity factors ranging between 0.001 and 0.01, which means that *Daphnia* is at least 100-fold more sensitive than fish.



**Figure 4.4: Aniline and its derivatives.**

It has to be highlighted that the Verhaar classification scheme is built on acute fish toxicity, and that chemicals are attributed to the four MOA classes based on structural characteristics. However, acute toxicity mechanisms are different from chronic toxicity mechanisms. This can explain why chemicals acting via endocrine disruption mechanisms are not identified as "specifically acting chemicals" (MOA 4) but as, for example, MOA 2 in the case of ethinyl oestradiol and 17 $\beta$ -oestradiol. In the same way, aniline and its derivatives are classified as polar narcotics which are characterised by a slightly higher toxicity to fish than the baseline toxicity of MOA 1 chemicals, although its toxicity to *Daphnia* is much higher.

### 4.1.2. *Daphnid to chronic fish relationships by MOA*

To check whether or not the goodness-of-fit depends on the MOA, the linear regression was performed for each MOA class, using the geometric mean approach. The results are shown in Table 4.4, Figure 4.5 (acute toxicity), and Figure 4.6 (chronic toxicity)

#### 4.1.2.1. Relationships

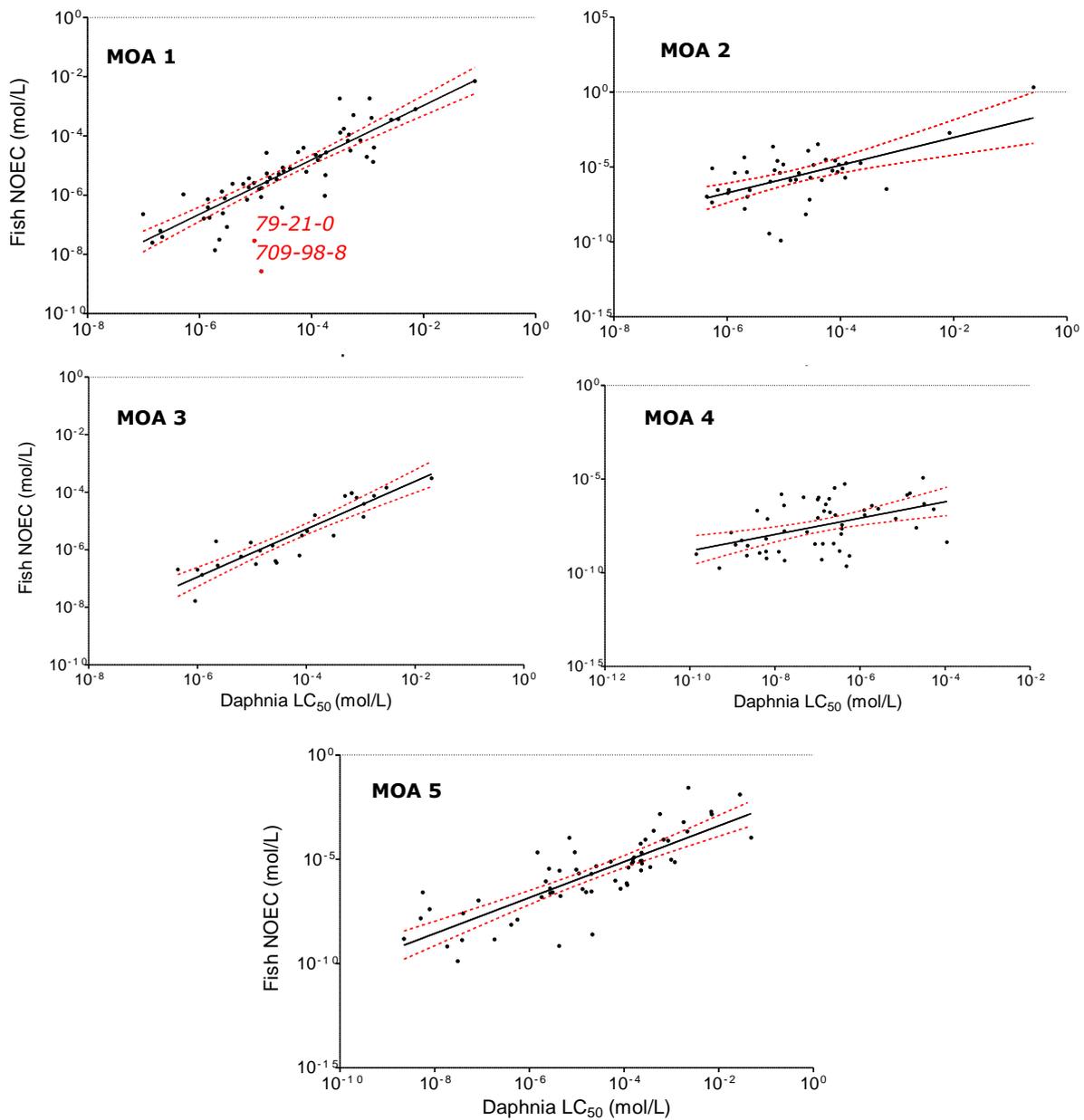
The relationships are highly significant ( $p \leq 0.0001$ ) for four classes of MOA; i.e. MOAs 1, 2, 3, and 5. For MOA 4 chemicals the relationships are not significant with  $p = 0.0004$  for the relationship based on acute *Daphnia* data and  $p = 0.4982$  for the relationship based on chronic *Daphnia* data. This means that chronic *Daphnia* toxicity would be a very poor predictor of chronic fish toxicity for MOA 4 chemicals, at least if all the MOA 4 are taken together. This makes sense as this is the group of chemicals with specific MOA which are not necessarily present in all taxa. It would therefore be interesting to divide further this MOA 4 group and to investigate the relationship of specific mechanisms (e.g. AchE inhibition, neurotoxicant, respiratory blocker, etc).

**Table 4.4: Linear relationships between *Daphnia* acute or chronic toxicity and fish chronic toxicity data depending on the MOA.**

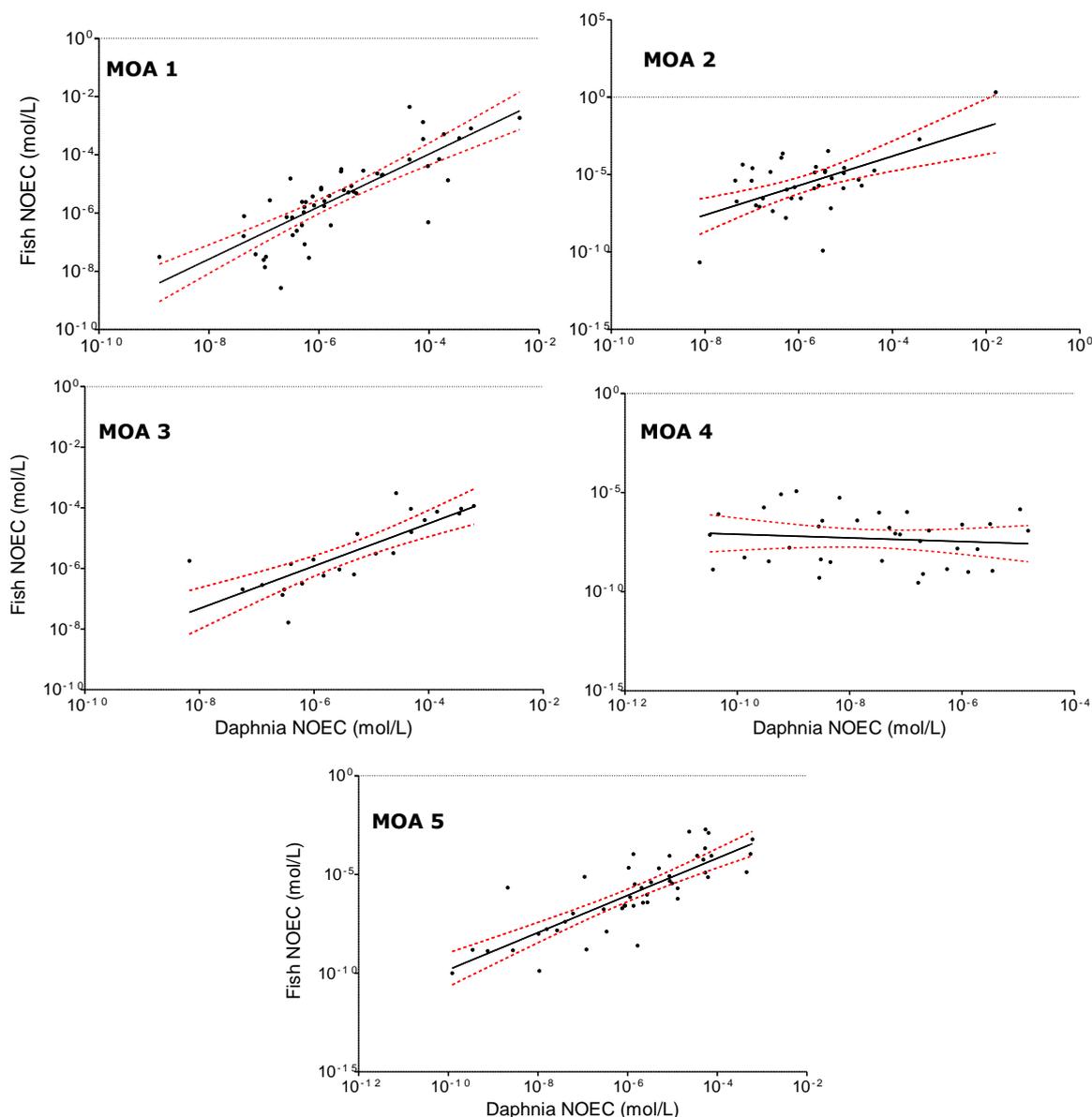
	<b>MOA 1</b>	<b>MOA 2</b>	<b>MOA 3</b>	<b>MOA 4</b>	<b>MOA 5</b>
<b>LC<sub>50</sub> <i>Daphnia</i></b>					
<b>n</b>	62	42	27	52	63
<b>p value</b>	< 0.0001	< 0.0001	< 0.0001	0.0004	< 0.0001
<b>r<sup>2</sup></b>	0.79	0.36	0.83	0.22	0.66
<b>Outlier</b>	2	0	0	0	0
	709-98-8; 79-21-0				
<b>NOEC <i>Daphnia</i></b>					
<b>n</b>	52	36	23	34	48
<b>p value</b>	< 0.0001	0.0001	< 0.0001	0.5168	< 0.0001
<b>r<sup>2</sup></b>	0.66	0.35	0.65	0.01	0.67
<b>Outlier</b>	0	0	0	0	0

In both cases, the  $r^2$  of logged data is higher for MOAs 1 and 3 (respectively, 0.79/0.83 for MOA 1/MOA 3 based on acute *Daphnia* data, and 0.66/0.65 for MOA 1/MOA 3 based on chronic *Daphnia* data), and much lower for MOAs 2 and 4 chemicals (respectively, 0.36/0.22 for MOA 2/MOA 4 based on acute *Daphnia* data and 0.35/0.01 for MOA 2/MOA 4 based on acute *Daphnia* data). The strongest relationship is therefore obtained with acute *Daphnia* data, and not with chronic *Daphnia* data.

The  $r^2$  for MOA 5 chemicals is rather high. This could suggest that many MOA 1 or 3 chemicals might be present in the MOA 5 group knowing that the MOA 5 class is a mix of all the chemicals that have not been allocated to MOA 1 to 4 classes, i.e. a mix of MOAs 1, 2, 3 and 4 chemicals that have not been identified as such.



**Figure 4.5: Relationships between *Daphnia* LC<sub>50</sub> and fish NOEC values by MOA. The dashed lines represent the 95% confidence interval. *Italics*: CAS number of the outliers.**

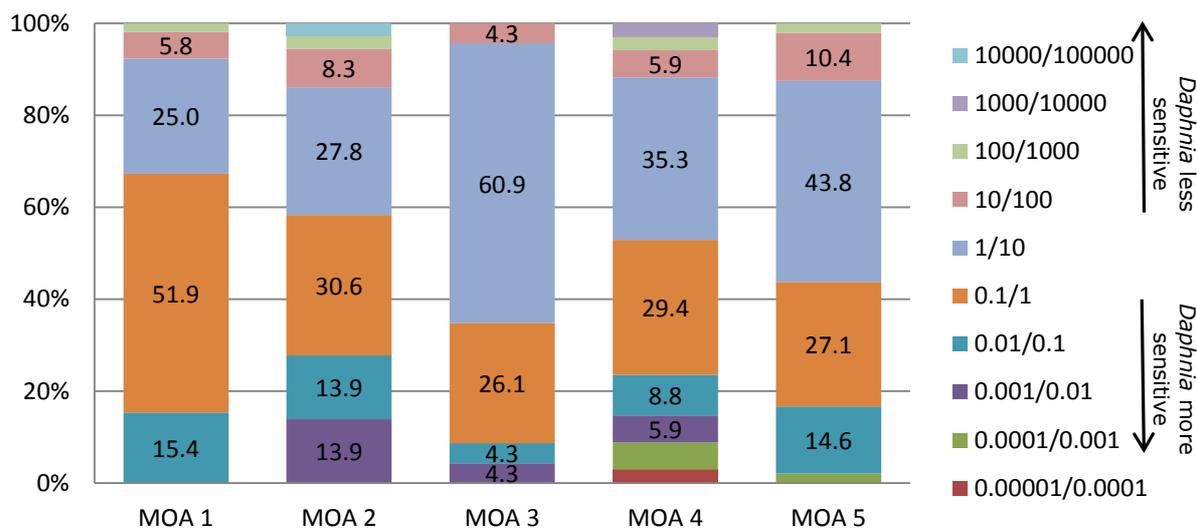


**Figure 4.6: Relationships between *Daphnia* NOEC and fish NOEC values by MOA. The dashed lines represent the 95% confidence interval. *Italics*: CAS number of the outliers.**

#### 4.1.2.2. Outliers

Only two outliers have been identified, both belong to the MOA 1 class and are present in the relationship based on acute *Daphnia* data. The two outliers are propanil (CAS No: 709-98-8) and peracetic acid (CAS No: 79-21-0). Propanil is a widely used contact herbicide which mainly acts against weeds by inhibiting their photosynthesis and CO<sub>2</sub> fixation, and which shows a very high chronic toxicity toward fish compared to the other MOA 1 chemicals. Peracetic acid is used as an antimicrobial. Only one NOEC value for fish was available for peracetic acid in the ECHA database (0.0022 mg/L) representing a nominal value in the middle of the range of concentrations tested (0.2, 0.7, 2.2, 7.5, 22.4 µg peracetic acid/L).

### 4.1.2.3. Sensitivity ratios



**Figure 4.7: Distribution of the chronic sensitivity ratios between *Daphnia* NOEC and fish NOEC according to their MOA.**

The distribution of the sensitivity ratios according to their MOAs (Figure 4.7) shows that 87% of the MOA 3 chemicals are within a chronic sensitivity ratio *Daphnia*/fish of 10, and 95.6% are within a factor of 100. However, for the majority of the MOA 3 chemicals, *Daphnia* seems less sensitive than fish (60.9% of the MOA 3 chemicals have a sensitivity ratio between 1-10). Regarding MOA 1 chemicals, 76.9% of the chemicals have a sensitivity ratio within 10, *Daphnia* being more sensitive than fish for 51.9% of them; and 98.1% of the MOA 1 chemicals are within a sensitivity ratio of 100.

For MOA 4, the sensitivity ratios cover a broader range, i.e. only 64.7% of the MOA 4 chemicals have a chronic sensitivity ratio within a factor of 10, *Daphnia* being more sensitive for only 29.4% of the chemicals. 79.4% of these chemicals have a sensitive ratio within 100; which means that 20% of the MOA 4 chemicals have very high values of sensitivity ratio. MOA 2 chemicals also follow this pattern with only 58.4% of chemicals within a sensitivity ratio of 10 and 80.6% within a factor 100.

## 4.2. Acute to chronic fish data

### 4.2.1. Acute to chronic fish relationships – irrespective of the MOA

#### 4.2.1.1. Relationships

There were 240 substances with acute fish toxicity data and chronic fish data. There was a significant relationship between acute and chronic fish toxicity data with both approaches ( $p < 0.0001$ ), but as previously, the goodness-of-fit was improved by using geometric means instead of the lowest values ( $r^2$  of logged data = 0.82 and 0.87 respectively for the lowest value approach and the geometric mean approach, see Table 4.5). Data are plotted in Figure 4.8.

Table 4.5: Linear relationships between acute fish data and chronic fish data.

	<b>Lowest value</b>	<b>Geometric mean</b>
<b>n</b>	240	
<b>p value</b>	< 0.0001	< 0.0001
<b>r<sup>2</sup></b>	0.82	0.87
<b>Outlier</b>	3	4
	1897-45-6; 709-98-8; 298-00-0	1897-45-6; 56- 38-2; 709-98- 8; 135-19-3

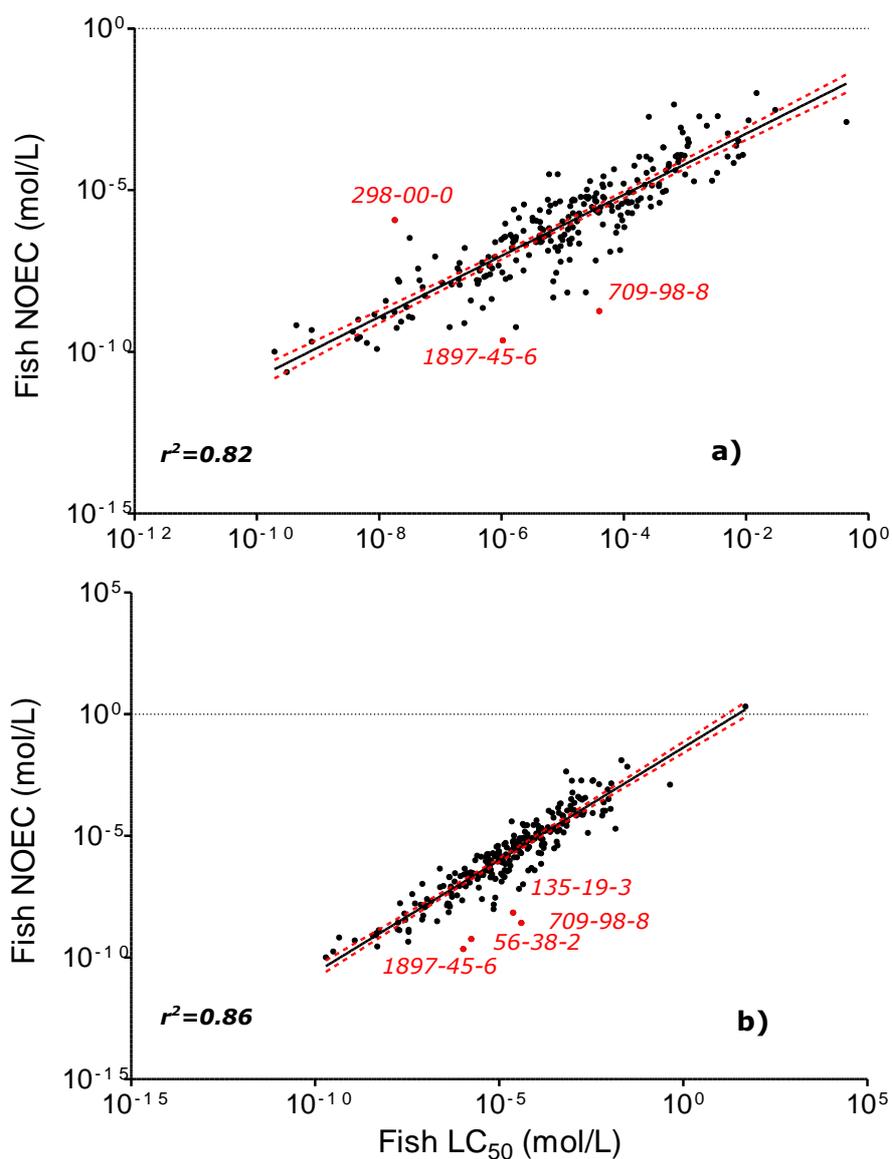


Figure 4.8: Relationships between acute LC<sub>50</sub> fish data and fish NOEC values from the OECD QSAR Toolbox 2.3 (n=240). a: Lowest value approach, b: Geometric mean approach. The dashed lines represent the 95% confidence interval. *Italics*: CAS number of the outliers.

#### 4.2.1.2. Discussion of outliers

With either of the two approaches, two outlier chemicals are identical whereas the other outlier chemicals depend on the approach used (Figure 4.8). The name and the MOA of three outliers are presented in Table 4.6.

**Table 4.6: Name and MOA of outliers of the acute to chronic relationships on fish data.**

<b>Name of chemical</b>	<b>CAS Number</b>	<b>MOA</b>	<b>Lowest value approach</b>	<b>Geometric mean</b>
Methyl parathion	298-00-0	4	+	
Propanil	709-98-8	1	+	+
Chlorothalonil	1897-45-6	4	+	+
2-naphthol	135-19-3	2		+
Ethyl-parathion	56-38-2	4		+

Two chemicals, propanil and chlorothalonil, are identified as outliers in both approaches, because their chronic fish toxicity is higher than expected when considering their acute fish toxicity. Propanil is a widely used contact herbicide, which acts against weeds by inhibiting their photosynthesis and CO<sub>2</sub> fixation. According to Russom et al. (1997), it is classified as a MOA 1 chemical acting as a non-polar narcotic in acute toxicity to fish. However, the OECD QSAR Toolbox identified this chemical as a MOA 5 chemical. Chlorothalonil is a broad spectrum non-systemic fungicide (see chapter 4.1.1.3) which acts via the formation of substituted chlorothalonil-reduced glutathione (GSH) derivatives in cells, which trigger inhibition of specific nicotinamide adenine dinucleotide (NAD) thiol-dependent glycolytic and respiratory enzymes (Tillman et al., 1973). It is classified as a specifically acting chemical (MOA 4).

With the geometric mean approach, two other chemicals are identified as outliers because of a higher chronic toxicity to fish, 2-naphthol, a widely used intermediate for the production of dyes and other compounds, classified as a MOA 2, and ethyl-parathion, an organophosphate pesticide (MOA 4 chemical).

With the lowest value approach, methyl parathion is also identified as an outlier because of its lower chronic fish toxicity in comparison to its acute toxicity to fish. This organophosphate pesticide was already mentioned because of a high interspecies sensitivity ratio between *Daphnia* and fish (see chapter 4.1.1.3).

#### 4.2.1.3. Acute to chronic ratio

The average ACR for fish over the 240 chemicals was 141.8, but the median is 8.9, which means that there are extremely high values which influence this average value. This is in line with some previous work reporting a median ACR for fish of 10.5 (Ahlers et al., 2006). Quite unexpectedly, for seven chemicals the ACR was below 1 (but within a factor of 10, the lowest ACR being 0.15), which means that the chronic toxicity (NOEC) value in the database was higher for those chemicals than the acute toxicity value. This might be due to the fact that different endpoints and different species have been considered in the database; and that the variability between species and/or endpoints might overcome the acute to chronic ratio. Amongst those seven chemicals, there are three MOA 1, two MOA 5, one MOA 4, and one MOA 2 (See Table 4.7). Usually MOA 4 chemicals are characterised by higher ACR due to their specific MOA; however, ziram, a pesticide, has an ACR of 0.54 according to our database.

**Table 4.7: Name and MOA of chemicals with ACR < 1.**

	<b>CAS number</b>	<b>Name</b>	<b>MOA</b>	<b>Ratio</b>
<b>ACR &lt; 1</b>	4747-21-1	N-methylisopropylamine	1	0.15
	108-18-9	Diisopropylamine	1	0.51
	137-30-4	Ziram	4	0.54
	106-93-4	1,2-dibromoethane	1	0.54
	70124-77-5	Flucythrinate	5	0.67
	107-51-7	Octamethyltrisiloxane	5	0.77
	110-86-1	Pyridine	2	0.91

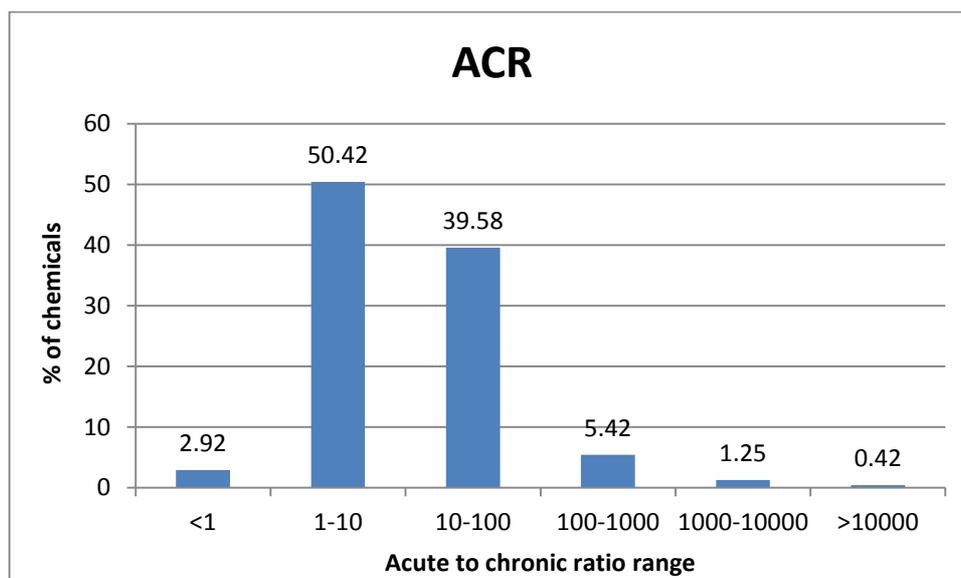
For ziram, there are two fish LC<sub>50</sub> (0.0097 and 0.57 mg/L) and two NOEC values (0.101 and 0.189 mg/L) available in our database. They have been extracted from the ECHA database and are considered reliable data. It was not possible to go back to the original reference of the pyridine data; however, the very low toxicity (one LC<sub>50</sub> value of 99.6 mg/L and one NOEC value of 110 mg/L) probably means that this higher NOEC is due to the experimental design.

Regarding flucythrinate, a pyrethroid pesticide, two LC<sub>50</sub> values of 0.0002 mg/L and one NOEC value of 0.0003 mg/L on two different species were extracted from the Aquatic ECETOC database. For octamethyltrisiloxane, an ectoparasiticides drug, all data were extracted from the ECHA database: one LC<sub>50</sub> value of 0.0194 mg/L on *Oncorhynchus mykiss* along with two NOEC values in the same range (0.027 mg/L on *Oncorhynchus mykiss* for growth, survival and hatching, and 0.021 mg/L for survival on *Pimephales promelas*). We have some doubts on the value reported as LC<sub>50</sub> since this value is also given as NOEC value of the same study.

The ECHA database also reported two values for n-methylisopropylamine: one LC<sub>50</sub> 96h on *Danio rerio* with mortality initiating from 43.3 mg/L (acute fish toxicity test, according to the OECD TG203, carried out in 2008) the calculated LC<sub>50</sub> was 48.5 mg/L; and one NOEC value >323 mg/L after 28 days of exposure (Juvenile growth test carried out in 2011 according to the OECD TG215) on the same species. There was no explanation for the difference between those two values. Regarding diisopropylamine, there were six LC<sub>50</sub> values on four different species (37, 40, 42, 75, 196 and 798 mg/L, which is a rather broad and unusual range (factor of 21 between the higher and the lower; LC<sub>50</sub> reported) and only one NOEC value of 187 mg/L. The highest LC<sub>50</sub> of 798 mg/L and the NOEC value were obtained on the same species, *Gasterosteus aculeatus*, which does not seem to be very sensitive to this chemical. All data were extracted from the ECHA database in which they were ranked as reliable with a Klimisch score of 1 or 2.

Regarding 1,2 bromoethane, one fish LC<sub>50</sub> value (32.1 mg/L) and two NOEC values (5.81 and 9.62 mg/L for growth and mortality obtained with *Oryzia latipes*) were extracted from the Aquatic ECETOC database. In addition to these data, the ECHA database contains two more LC<sub>50</sub> values derived with *Oncorhynchus mykiss* (1.13 and 1.86 mg/L) and labelled with a Klimisch score of 1. This corresponds to a factor 28 between the lowest and the highest LC<sub>50</sub> value.

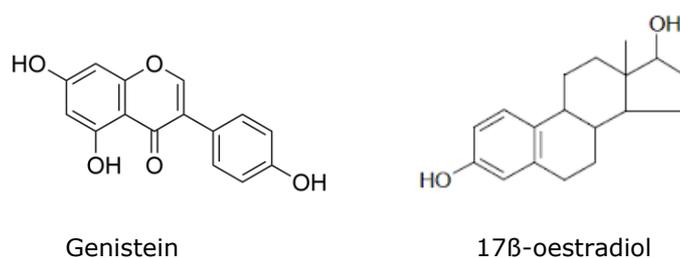
The range of ACR is quite broad (up to 14906) but 53.34% of the chemicals have an ACR below 10 and 92.9% have an ACR below 100 (see Figure 4.9). The maximum value of 14906 is therefore far above the maximum ACR of 2450 found by Ahlers et al. (2006) The 90<sup>th</sup> percentile value is 70.85. If we apply the Ahlers et al. (2006) value of 30 to discriminate between low and high ACR, 190 chemicals (79.2%) have a low and 50 (20.8%) a high ACR.



**Figure 4.9: Acute to chronic fish toxicity ratio distribution.**

The 17 chemicals with ACR above 100 are presented in Table 4.8. Quite unexpectedly, six (35%) of these high ACR are MOA 1 chemicals, five (29%) are MOA 4 chemicals, three (17%) are MOA 2 chemicals and three (17%) are MOA 5 chemicals. Ahlers et al. (2006) previously found that MOA 1 was a good predictor that a substance would have a lower ACR, although other MOAs were not necessarily associated with higher ACRs. The MOA 1 are genistein, monoethanolamine, dimethylamine, peracetic acid and diuron.

Genistein is a phytoestrogen and belongs to the category of isoflavones. Due to its structure similarity to 17 $\beta$ -oestradiol (Figure 4.10) it has been shown to interact with animal and human oestrogen receptors (Green, 2015; Patisaul et al., 2002), but also to bind to and transactivate all three PPAR isoforms,  $\alpha$ ,  $\delta$ , and  $\gamma$  (Wang et al., 2014) and to possibly inhibit the DNA methyltransferase (Fang et al., 2007). Only one LC<sub>50</sub> value is reported for genistein (1.9 mg/L), along with five NOEC values coming from the same study (0.0013 to 0.0042 mg/L). All studies have been carried out with *Danio rerio*.



**Figure 4.10: Structural similarity between genistein and 17 $\beta$ -oestradiol.**

**Table 4.8: Name and MOA of chemicals with ACR >100.**

	<b>CAS number</b>	<b>Name</b>	<b>MOA</b>	<b>Ratio</b>
<b>100/1000</b>	181587-01-9	5-amino-1-[2,6-dichloro-4-(trifluoromethyl)phenyl]-4-(ethylsulfinyl)-1H-pyrazole-3-carbonitrile	4	134
	2212-67-1	Molinate	4	155
	330-54-1	Diuron	1	161
	124-40-3	Dimethylamine	1	192
	606-20-2	2,6-dinitrotoluene	5	193
	1912-24-9	Atrazine	5	232
	79-21-0	Peracetic acid	1	270
	67-68-5	Dimethylsulfoxide	5	340
	95-76-1	3,4-dichloro-aniline	2	430
	944-22-9	Fonofos	4	468
	98-54-4	4-tertbutylphenol	2	521
	446-72-0	Genistein	1	723
	141-43-5	Monoethanolamine	1	738
<b>1000/1000</b>	56-38-2	Ethyl parathion	4	2962
	135-19-3	2-naphthol	2	3458
	1897-45-6	Chlorothalonil	4	4646
<b>&gt;10000</b>	709-98-8	Propanil	1	14906

Monoethanolamine is a primary amine used as feedstock in the production of detergents, emulsifiers, polishes, pharmaceuticals, corrosion inhibitors, or chemical intermediates. One NOEC value of 1 mg/L (*Oryzias latipes*) was extracted from the ECHA database, along with four LC<sub>50</sub> values (170, 349, 2070 and 2120 mg/L, on four different species: *Cyprinus carpio*, *Carassius auratus*, *Pimephales promelas* and *Poecilia reticulata*). Dimethylamine is a secondary amine also characterised by high fish LC<sub>50</sub> values (17, 118, 120, 210, 396 mg/L, derived with three different species, *Oncorhynchus mykiss*, *Danio rerio* and *Poecilia reticulata*). One NOEC value of 0.6 mg/L is reported for egg mortality, which is the only one considered in our analysis as it is the only precise NOEC value reported (the other NOEC values are reported as >10 mg/L and >20 mg/L). Therefore, the difference in sensitivity here is due to the particular life stage considered for this study.

Regarding peracetic acid, nine LC<sub>50</sub> values were reported in the ECHA database, ranging from 0.078 to 2 mg/L and derived with three different species (*Lepomis macrochirus*, *Oncorhynchus mykiss*, *Danio rerio*), whereas only one NOEC value (0.0022 mg/L; *Danio rerio*) was reported. For diuron, a herbicide acting as photosynthesis inhibitor by blocking the plastoquinone binding site of photosystem II (Metz et al., 1986). For diuron, two LC<sub>50</sub> values were reported (14 mg/L and 14.7 mg/L, on *Pimephales promelas* and *Oncorhynchus mykiss*, respectively) and five NOEC values (from 0.029 to 0.41 mg/L). The lowest values (0.0269 and 0.0334 mg/L) were extracted from the US EPA Ecotox database, from a single study carried out in 1987 on *Pimephales promelas*. The highest value was extracted from the ECHA database (0.41 mg/L) from a study carried out in 1993 on *Oncorhynchus mykiss*.

The herbicide propanil (also previously identified as an outlier, see chapter 4.1.2.2 and 4.2.1.2) has the highest ACR (14906). This is in line with previous work reporting an ACR of 18100 (Kenaga, 1982). This ACR is based on one LC<sub>50</sub> entry (8.68 mg/L) and five NOEC entries from two studies, ranging from 0.0004 to 0.0012 mg/L. Other data

sources, as the US EPA in the Reregistration Eligibility Decision retain a NOEC value of 0.0091 mg/L (236 days), and a LC<sub>50</sub> of 2.3 mg/L. This would result in an ACR of 252 being still considered as a high ACR but much lower than the previous one derived from our database.

Chlorothalonil was also previously identified as an outlier (see chapter 4.1.1.3 and 4.2.1.2). Its high ACR is based on one fish LC<sub>50</sub> (0.278 mg/L) and one NOEC value (0.00006 mg/L). This high chronic toxicity to fish, extracted from the ECOTOX US EPA database, was found in the literature (Teather et al., 2005), and is related to hatching survival of fertilised eggs after 36 days of exposure. However, when going back to the original study, it appears that 0.00006 mg/L was the only concentration tested (i.e. the concentration for which we start to observe an effect might be higher) and that the three duration schemes were 7 days, 3 weeks and 5 months and not 36 days. Therefore this data might not be reliable.

The two chemicals with the next highest ACR are the outliers previously identified with the geometric mean approach: the organophosphate pesticide ethylparathion and 2-naphthol, a widely used intermediate for the production of dyes and other compounds (MOA 2). Only one NOEC value is reported for ethylparathion (0.17 µg/L, on growth), along with two LC<sub>50</sub> values (0.5 and 0.51 mg/L). All these data come from the same study<sup>4</sup>, however, it was not possible to access the original reference. Regarding 2-naphthol, only one LC<sub>50</sub> and one NOEC value are available (3.46 and 0.001 mg/L, respectively), both extracted from the ECHA database. However, it was not possible to retrieve more details that could explain these high ACR.

3,4 dichloroaniline is a data-rich chemical with 24 NOEC values, ranging from 0.0011 to 0.23 mg/L and derived with five different fish species and 20 LC<sub>50</sub> value, ranging from 1.94 to 13.3 mg/L. It was not possible to go back to all of the original studies but high ACR (i.e. 1200) has already been reported for this chemical in the literature (Call et al., 1987).

Fonofos ACR is based on one LC<sub>50</sub> value (1.09 mg/L on *Pimephales promelas*) and one NOEC value (0.00233 mg/L on *Lepomis macrochirus*) (Fairchild et al., 1992). In the same study, a LC<sub>50</sub> of 5.3 µg/L (0.0053 mg/L) was reported for this organothiophosphate insecticide (*Lepomis macrochirus*, 96h) which was not included in our database. Regarding terbutylphenol, the ACR is also based on only two data entries, one LC<sub>50</sub> value (5.21 mg/L) and one NOEC value (0.01 mg/L), on the same species, *Pimephales promelas*. The NOEC value comes from the ECHA database from an early life stage toxicity study (2008, ranked Klimisch score 1). The ECHA database also reports a LC<sub>50</sub> value >1 mg/L.

Atrazine is another data-rich chemical with 12 NOEC values ranging from 0.0302 to 0.21 mg/L, on two different species (*Pimephales promelas* and *Lepomis macrochirus*) and different life stages (juvenile, larvae and adults). Two LC<sub>50</sub> values are reported (20.5 and 18.8 mg/L) from two studies on two different species (*Onchorynchus mykiss* and *Cyprinus carpio*). Atrazine is a triazine herbicide which acts by binding to the plastoquinone-binding protein in photosystem II. Atrazine has also been shown to have endocrine disrupting effects (Mizota and Ueda, 2006; Prossnitz et al., 2007) including on wildlife (Hayes et al., 2003, 2002)

Dinitrotoluene is a nitroaromatic chemical used in the production of flexible polyurethane foams and as a plasticiser, deterrent coating and burn rate modifier in propellants. It is known to be toxic by converting haemoglobin into methaemoglobin and to be

---

<sup>4</sup> Acute and Chronic Parathion Toxicity to Fish and Invertebrates, 1981, Spacie, A. Vilkas, A. G. Doebbler, G. F. Kuc, W. J. Iwan, G. R. Technical report. Environmental Protection Agency Office of research and development, EPA-68-01-0155

carcinogenic. Only two LC<sub>50</sub> values (34 and 18.6 mg/L) and one NOEC value were reported (0.13 mg/L), for which it was not possible to go back to the original study.

## 4.2.2. Acute to chronic fish relationship by MOA

### 4.2.2.1. Relationships

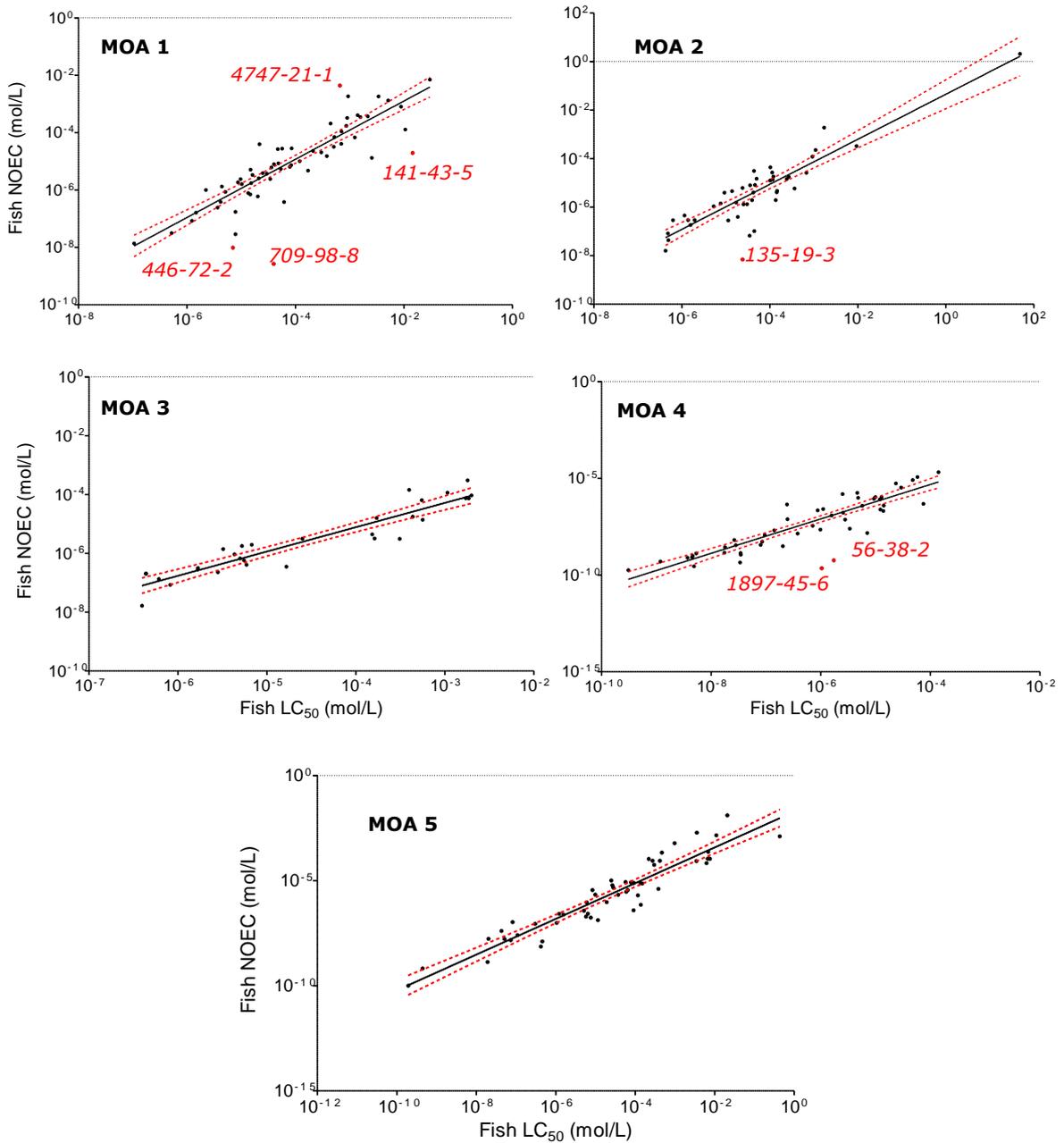
To check whether or not the goodness-of-fit depends on the MOA, the linear regression was performed for each MOA class. The results are presented in Table 4.9 and plotted in Figure 4.11. There were highly significant relationships between acute and chronic toxicity for each MOA class ( $p < 0.0001$ ), with high  $r^2$  ( $> 0.80$ ) for every MOA. The highest  $r^2$  is obtained with MOA 3 and 5 chemicals ( $r^2 = 0.88$ ).

**Table 4.9: Linear relationships between fish LC<sub>50</sub> and fish chronic toxicity data depending on the MOA.**

	<b>MOA 1</b>	<b>MOA 2</b>	<b>MOA 3</b>	<b>MOA 4</b>	<b>MOA 5</b>
<b>n</b>	62	44	30	50	54
<b>p value</b>	< 0.0001	0.0001	< 0.0001	< 0.0001	< 0.0001
<b>r<sup>2</sup></b>	0.83	0.83	0.88	0.85	0.88
<b>Outlier</b>	4	1	0	2	0
	141-43-5; 446-72-0; 709-98-8; 4747-21-1	135-19-3		1897-45-6; 56-38-2	

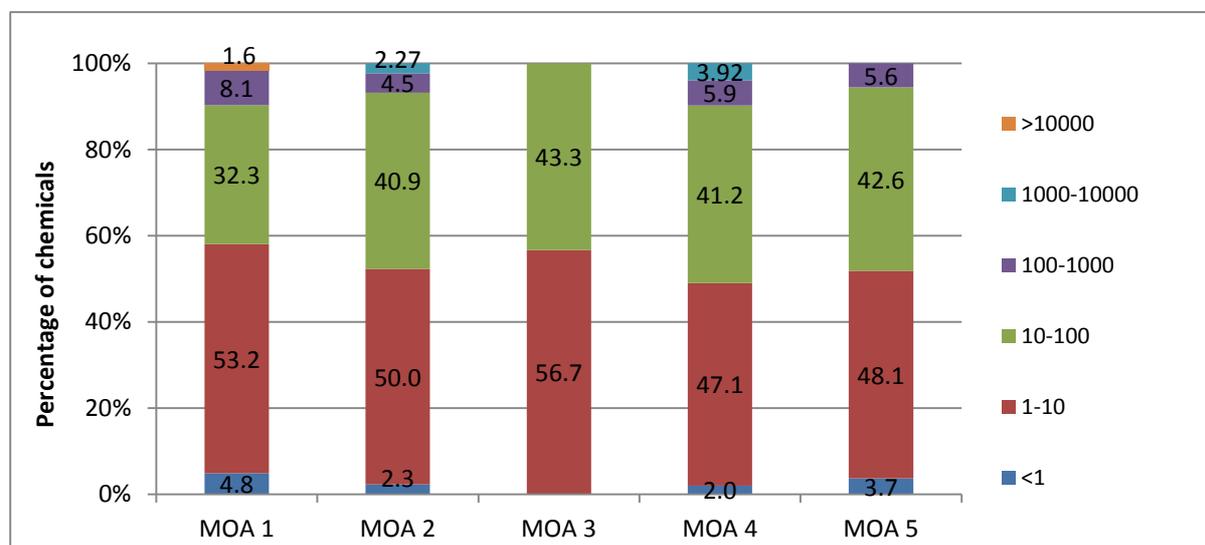
### 4.2.2.2. Discussion of outliers

Except for methyl parathion, all the chemicals previously identified as outliers in the relationship based on the whole set of chemicals are also identified as outliers in their MOA groups, with the exception of methyl parathion: propanil (MOA 1), 2-naphthol (MOA 2), ethylparathion and chlorothalonil (MOA 4) See chapter 4.2.1.2 for more details. In addition, in the MOA 1 chemicals group, three other chemicals are outliers: monoethanolamine (CAS No: 141-43-5), genistein (CAS No: 446-72-0) and n-methylisopropylamine (CAS No: 4747-21-1). All of them have already been mentioned because of their high or low (for n-methylisopropylamine) ACR (see chapter 4.2.1.3).



**Figure 4.11: Relationships between fish LC<sub>50</sub> and fish NOEC values by MOA. The dashed lines represent the 95% confidence interval. *Italics*: CAS number of the outliers.**

### 4.2.2.3. Acute to chronic ratio and MOA



**Figure 4.12: Distribution of ACR (acute to chronic fish toxicity ratio) of chemicals according to their MOA.**

The distribution of the ACR according to MOA (see Figure 4.12) shows that 100% of the MOA 3 chemicals have an ACR within a factor of 100, and 56.7% within a factor of 10. 90.9% of the MOA 2 chemicals have an ACR within a factor of 100, with 50% of chemicals having an ACR below a factor of 10. 88.2% of the MOA 4 chemicals have an ACR within a factor of 100, of which 47% have an ACR below 10. However, only 85.5% of the MOA 1 chemicals are within an ACR of 100, with 53.2% chemicals with an ACR below 10.

Overall, these results show that an ACR of 100 when applied to acute fish toxicity data for substances with MOA 3 and 2, as defined by the Verhaar classification scheme, will protect against the chronic effects of industrial substances in respectively 100 and 90% of the cases. That is in line with some previous work that show that an ACR of 100, as implied by the European guidance (ECHA, 2008), is protective for >90% of the industrial chemicals (May and Hahn, 2014). However for MOA 4 and MOA 1 chemicals, an ACR of 100 would be sufficiently protective for only 88 and 85% of the chemicals.

## 5. Conclusions and recommendations

When several toxicological data are available for a given chemical, the lowest value is usually used in environmental risk assessment in the interest of being as protective as possible. This is especially important for the setting of PNEC, for example, when it is derived from a single toxicological value in a given species but aims to protect all the other species. However, in the context of this report, it has been shown that when it comes to the derivation of predictive relationships by regression modelling, the use of the geometric mean, calculated across the full range of empirical data available rather than just the most sensitive value, allows more stable and representative estimates of chronic fish toxicity, and improves the goodness-of-fit of the linear regressions. Therefore, this approach decreases the statistical influence of outliers and takes into account the whole set of toxicological information available for each chemical.

Our results show that the best fitted relationships for predicting chronic fish toxicity are based on acute fish data ( $r^2=0.87$ ). Unexpectedly, acute *Daphnia* data ( $r^2=0.64$ ) also gives a stronger relationship with chronic fish data than chronic *Daphnia* data ( $r^2=0.59$ ) when dealing with the whole set of chemicals regardless of the MOA. When considering the MOA according to the Verhaar scheme, MOA 3 and MOA 1 chemicals give the strongest acute *Daphnia* to chronic fish relationship and chronic *Daphnia* to chronic fish relationship. However, the relationships obtained with acute *Daphnia* data show a much better fit ( $r^2=0.83$  and  $0.69$  for MOA 3 and MOA 1, respectively) than the one obtained with chronic *Daphnia* data ( $r^2=0.66$  and  $0.65$  for MOA 1 and MOA 3, respectively). When considering acute fish data, all the MOA give highly fitted relationships ( $r^2=0.88$  for MOA 3 and MOA 5 chemicals,  $0.85$  for MOA 4 chemicals and  $0.83$  for MOA 1 and MOA 2 chemicals).

On the basis of these results it can be concluded that when acute fish toxicity data are available, they might give a reliable basis to extrapolate to chronic fish toxicity as a first tier assessment or within a weight of evidence approach. When no acute fish toxicity data are available, acute *Daphnia* toxicity data could be used as a surrogate at least for MOA 3 and MOA 1 chemicals. For MOA 4 chemicals, it has to be highlighted that this group brings together chemicals with highly specific toxicity and particular MOA; i.e. it considers as a whole chemicals acting by various and different MOAs such as neurotoxicants, AchE inhibitors and respiratory inhibitors. Moreover, the more specific is a MOA, the more uncertain will be the extrapolation of toxicity data to other species, especially if the mode of action is not conserved across taxa. This can explain why the relationships obtained with MOA 4 are not significant and/or not well-fitted. It would be interesting to further breakdown this group to look at each specific MOA individually.

There is a correlation between chemicals with high ACR values or interspecies sensitivity ratio and the outliers identified in the above-mentioned relationships. When considering chemicals with a high interspecies sensitivity ratio, *Daphnia* being more sensitive than fish, several aniline derivatives and pesticides acting through cholinesterase inhibition were identified. When considering high interspecies sensitivity ratio chemicals for which *Daphnia* is less sensitive than fish, we found pesticides and known endocrine disruptors such as ethynil oestradiol and  $17\beta$ -oestradiol.

Extreme interspecies sensitivity ratios were mainly MOA 2, 4 and 5 chemicals. Regarding the ACR between acute and chronic fish toxicity data, around 50% of the chemicals of each MOA class have an ACR within a factor of 10, whereas 100%, 90.9%, 88.3% and 85.5% of the MOA 3, MOA 2, MOA 4 and MOA 1 chemicals respectively have an ACR within a factor of 100. Therefore, the safety factor of 100 commonly applied in environmental risk assessment does not seem to be equally protective for every MOA. Thus, the ACR can be used as a first tier approach, whereas regression models provide more accurate, MOA-specific extrapolations from acute to chronic toxicity. Thus, when acute toxicity data are available, regression equations seem to provide a reliable,

chemical-specific means of extrapolation from acute to chronic toxicity data, and could be used in an integrated testing strategy in order to waive long-term toxicity testing on fish.

In the future, new tools could help to strengthen extrapolation approaches. For example Adverse Outcome Pathways (AOPs) could help to identify where cross-species extrapolation is appropriate (Ankley et al., 2010; Burden et al., 2015) based on the conservation of the toxicological pathway between species. AOPs could also help in the grouping of chemicals based upon a common Molecular Initiating Event (MIE) or other key event (KE) to refine and improve relationships by restricting them to more specific subsets of chemicals. For data-poor chemicals, another option would be the use of the threshold of toxicological concern (TTC) approach, which aims to establish a level of exposure for chemicals below which there would be no appreciable risk. This concept is well established for assessing human safety and has been reapplied for a wide range of structural classes. Use of the TTC in environmental safety is just beginning, and key questions focus on hazard extrapolation of diverse taxa across trophic levels, importance of mode of action, and whether safe concentrations for ecosystems estimated from acute or chronic toxicity data are equally useful and in what context (Belanger et al., 2015).

In conclusion, the prediction of chronic fish toxicity can be based on acute fish toxicity data irrespective of MOA, as well as on acute *Daphnia* data, for MOA 3 and MOA 1 chemicals. MOA 5 chemicals are a heterogeneous group and need to be better characterised in terms of their MOA in order to develop additional MOA-based regression models. A tiered approach in which each tier allows for greater accuracy of prediction could be based on the sequential use of the TTC, ACR and interspecies ratio values, and MOA-based regression models that generate chemical-specific predictions of chronic toxicity.

## References

- Ahlers, J., Riedhammer, C., Vogliano, M., Ebert, R.-U., Kühne, R., Schüürmann, G., 2006. Acute to chronic ratios in aquatic toxicity-variation across trophic levels and relationship with chemical structure. *Environ. Toxicol. Chem.* 25, 2937–2945. doi:10.1897/05-701r.1
- Ankley, G.T., Bennett, R.S., Erickson, R.J., Hoff, D.J., Hornung, M.W., Johnson, R.D., Mount, D.R., Nichols, J.W., Russom, C.L., Schmieder, P.K., Serrano, J.A., Tietge, J.E., Villeneuve, D.L., 2010. Adverse outcome pathways: A conceptual framework to support ecotoxicology research and risk assessment. *Environ. Toxicol. Chem.* 29, 730–741. doi:10.1002/etc.34
- Austin, T.J., Eadsforth, C. V., 2014. Development of a chronic fish toxicity model for predicting sub-lethal NOEC values for non-polar narcotics. *SAR QSAR Environ. Res.* 25, 147–60. doi:10.1080/1062936X.2013.871577
- Barron, M.G., Lilavois, C.R., Martin, T.M., 2015. MOAtox: A comprehensive mode of action and acute aquatic toxicity database for predictive model development. *Aquat. Toxicol.* 161C, 102–107. doi:10.1016/j.aquatox.2015.02.001
- Belanger, S.E., Sanderson, H., Embry, M.R., Coady, K., DeZwart, D., Farr, B.A., Gutsell, S., Halder, M., Sternberg, R., Wilson, P., 2015. It is time to develop ecological thresholds of toxicological concern to assist environmental hazard assessment. *Environ. Toxicol. Chem.* 34, 2864–2869. doi:10.1002/etc.3132
- Burden, N., Sewell, F., Andersen, M.E., Boobis, A., Chipman, J.K., Cronin, M.T.D., Hutchinson, T.H., Kimber, I., Whelan, M., 2015. Adverse Outcome Pathways can drive non-animal approaches for safety assessment. *J. Appl. Toxicol.* 35, 971–975. doi:10.1002/jat.3165
- Call, D.J., Poirier, S.H., Harting, M.L.K.S.L., Lindberg, C.A., 1987. Toxicity of 3, 4-Dichloroaniline to fathead minnows, *Pimephales promelas*, in acute and early life-stage exposures. *Bull. Environ. Contam. Toxicol.* 352–358.
- Cassani, S., Kovarich, S., Papa, E., Roy, P.P., van der Wal, L., Gramatica, P., 2013. Daphnia and fish toxicity of (benzo)triazoles: Validated QSAR models, and interspecies quantitative activity-activity modelling. *J. Hazard. Mater.* 258-259, 50–60. doi:10.1016/j.jhazmat.2013.04.025
- Claeys, L., Iaccino, F., Janssen, C.R., Van Sprang, P., Verdonck, F., 2013. Development and validation of a quantitative structure-activity relationship for chronic narcosis to fish. *Environ. Toxicol. Chem.* 32, 2217–25. doi:10.1002/etc.2301
- de Haas, E.M., Eikelboom, T., Bouwman, T., 2011. Internal and external validation of the long-term QSARs for neutral organics to fish from ECOSAR™. *SAR QSAR Environ. Res.* 22, 545–559. doi:10.1080/1062936X.2011.569949
- de Roode, D., Hoekzema, C., de Vries-Buitenweg, S., van de Waart, B., van der Hoeven, J., 2006. QSARs in ecotoxicological risk assessment. *Regul. Toxicol. Pharmacol.* 45, 24–35. doi:10.1016/j.yrtph.2006.01.012
- Dom, N., Knapen, D., Blust, R., 2012. Assessment of aquatic experimental versus predicted and extrapolated chronic toxicity data of four structural analogues. *Chemosphere* 86, 56–64. doi:10.1016/j.chemosphere.2011.08.050
- EC, 2009a. Regulation (EC) No 1107/2009 concerning the placing of plant protection products on the market and repealing Council Directives 79/117/EEC and 91/414/EEC. OJ L309.
- EC, 2009b. Regulation (EC) No 1223/2009 of the European Parliament and of the Council on cosmetic products. OJ L 342.

- EC, 2008a. Regulation (EC) No 429/2008 on detailed rules for the implementation of Regulation (EC) No 1831/2003 of the European Parliament and of the Council as regards the preparation and the presentation of applications and the assessment and authorisation of feed. OJ L133/1.
- EC, 2008b. Regulation (EC) No 1272/2008 on classification, labelling and packaging of substances and mixtures. OJ L 353/1.
- EC, 2006. Regulation (EC) No 1907/2006 of the European Parliament and of the Council concerning the Registration, evaluation, Authorisation and Restriction of Chemicals (REACH). OJ L396.
- ECETOC, 2007. Intelligent Testing Strategies in Ecotoxicology: Mode of Action Approach for Specifically Acting Chemicals - Technical Report No. 102. Brussels, Belgium. doi:ISSN-0773-0872-102
- ECETOC, 2006. Toxicological Modes of action: Relevance for human risk assessment. Technical Report No. 99. Brussels, Belgium.
- ECHA, 2012. Guidance on information requirements and chemical safety assessment Chapter R . 7b: Endpoint specific guidance (version 1.2), Guidance for the implementation of REACH. European Chemicals Agency. Helsinki, Finland.
- ECHA, 2008. Guidance on information requirements and chemical safety assessment: QSARs and grouping of chemicals (Chapter R.6): QSARs and grouping of chemicals. European Chemicals Agency. Helsinki, Finland.
- EFSA, 2014. Conclusion on the peer review of the pesticide risk assessment for aquatic organisms for the active substance imidacloprid. EFSA J. 12, 1–49. doi:10.2903/j.efsa.2014.3835.Available
- EFSA, 2008. Technical Guidance for assessing the safety of feed additives for the environment - Prepared by the Panel on Additives and Products or Substances used in Animal Feed. Parma.
- Ellison, C.M., Madden, J.C., Cronin, M.T.D., Enoch, S.J., 2015. Investigation of the Verhaar scheme for predicting acute aquatic toxicity: Improving predictions obtained from Toxtree ver. 2.6. Chemosphere 139, 146–154. doi:10.1016/j.chemosphere.2015.06.009
- EMA, 2006. Guideline on the environmental risk assessment of medicinal. European Medicines Agency. London, UK.
- EMA, 2004. Environmental Impact Assessment for Veterinary Medicinal Products: Phase II Guidance. CVMP/VICH/790/03-FINAL. European Medicines Agency. London, UK.
- Enoch, S.J., Hewitt, M., Cronin, M.T.D., Azam, S., Madden, J.C., 2008. Classification of chemicals according to mechanism of aquatic toxicity: an evaluation of the implementation of the Verhaar scheme in Toxtree. Chemosphere 73, 243–8. doi:10.1016/j.chemosphere.2008.06.052
- EU, 2013a. Regulation (EU) No 284/2013 setting out the data requirements for Plant Protection Products, in accordance with regulation (EC) No 1107/2009 of the European Parliament and of the council concerning the placing of plant protection products on the market. OJ L93.
- EU, 2013b. Regulation (EU) No 283/2013 setting out the data requirements for active substances, in accordance with Regulation (EC) No 1107/2009 of the European Parliament and of the Council concerning the placing of plant protection products on the market. OJ L93.
- EU, 2012. Regulation (EU) No 528/2012 concerning the making available on the market and use of biocidal products. OJ L 167/1.
- EU, 2010. Directive 2010/63/EU of the European Parliament and of the Council of 22

- September 2010 on the protection of animals used for scientific purposes, Official Journal of the European Union. OJ L 276/33. doi:32010L0063
- EURL ECVAM, 2014. EURL ECVAM Strategy to replace, reduce and refine the use of fish in aquatic toxicity and bioaccumulation testing. Joint Research Centre, European Commission, Ispra, Italy. doi:10.2788/084219
- Fairchild, J.F., Little, E.E., Huckins, J.N., 1992. Aquatic hazard assessment of the organophosphate insecticide fonofos. *Arch. Environ. Contam. Toxicol.* 22, 375–379. doi:10.1007/BF00212556
- Fang, M., Chen, D., Yang, C.S., 2007. Dietary polyphenols may affect DNA methylation. *J. Nutr.* 137, 223S–228S.
- Green, S.E., 2015. In vitro comparison of estrogenic activities of popular women's health botanicals. Master Sci. Thesis. University of Illinois. doi:10.1017/CBO9781107415324.004
- Grindon, C., Combes, R., Cronin, M.T.D., Roberts, D.W., Garrod, J., 2006. Integrated Decision-tree Testing Strategies for of the EU REACH Legislation. *In Vitro* 651–664.
- Hayes, T., Haston, K., Tsui, M., Hoang, A., Haeffele, C., Vonk, A., 2003. Atrazine-induced hermaphroditism at 0.1 ppb in American leopard frogs (*Rana pipiens*): Laboratory and field evidence. *Environ. Health Perspect.* 111, 568–575. doi:10.1289/ehp.5932
- Hayes, T., Kelly Haston, M.T., Anhthu Hoang, C.H., Vonk, A., 2002. Feminization of male frogs in the wild. *Nature* 419, 895–896.
- Hoekzema, C.C., Murk, A.J., Van de Waart, B.J., Van der Hoeven, J.C.M., De Roode, D.F., 2006. Alternative approaches can greatly reduce the number of fish used for acute toxicity testing. *Environ. Toxicol. Chem.* 25, 1322–1325. doi:10.1897/05-293R2.1
- Hutchinson, T.H., Barrett, S., Buzby, M., Constable, D., Hartmann, A., Hayes, E., Huggett, D., Laenge, R., Lillicrap, A.D., Straub, J.O., Thompson, R.S., 2003. A strategy to reduce the numbers of fish used in acute ecotoxicity testing of pharmaceuticals. *Environ. Toxicol. Chem.* 22, 3031–3036. doi:10.1897/02-558
- Jeram, S., Riego Sintes, J.M., Halder, M., Baraibar Fentanes, J., Sokull-Klüttgen, B., Hutchinson, T.H., 2005. A strategy to reduce the use of fish in acute ecotoxicity testing of new chemical substances notified in the European Union. *Regul. Toxicol. Pharmacol.* 42, 218–224. doi:10.1016/j.yrtph.2005.04.005
- Kar, S., Roy, K., 2010. First report on interspecies quantitative correlation of ecotoxicity of pharmaceuticals. *Chemosphere* 81, 738–747. doi:10.1016/j.chemosphere.2010.07.019
- Kenaga, E.E., 1982. Predictability of chronic toxicity from acute toxicity of chemicals in fish and aquatic invertebrates. *Environ Toxicol Chem* 1:347-358. *Environ. Toxicol. Chem.* 1, 347–358.
- May, M., Hahn, S., 2014. Comparison of species sensitivity of *Daphnia* and fish in acute and chronic testing - Report UBA-FB 27448. Fraunhofer-Institut für Toxikologie und Experimentelle Medizin. Hannover, Germany.
- Metz, J.G., Pakrasi, H.B., Seibert, M., Arntzer, C.J., 1986. Evidence for a dual function of the herbicide-binding D1 protein in photosystem II. *FEBS Lett.* 205, 269–274. doi:10.1016/0014-5793(86)80911-5
- Mizota, K., Ueda, H., 2006. Endocrine disrupting chemical atrazine causes degranulation through Gq/11 protein-coupled neurosteroid receptor in mast cells. *Toxicol. Sci.* 90, 362–368. doi:10.1093/toxsci/kfj087
- Motulsky, H.J., Brown, R.E., 2006. Detecting outliers when fitting data with nonlinear

- regression - a new method based on robust nonlinear regression and the false discovery rate. *BMC Bioinformatics* 7, 1–20. doi:10.1186/1471-2105-7-123
- Netzeva, T.I., Pavan, M., Worth, A.P., 2007. Review of (Quantitative) Structure–Activity Relationships for Acute Aquatic Toxicity. *QSAR Comb. Sci.* 27, 77–90. doi:10.1002/qsar.200710099
- OECD, 2013. OECD 210 Fish Early Life Stage Toxicity Test, OECD Guidelines for the testing of chemicals. Organisation for Economic Co-operation and Development - Paris, France. doi:10.1787/9789264203785-en
- OECD, 2012. OECD TG 211: Daphnia magna Reproduction Test, OECD Guidelines for the Testing of Chemicals. Organisation for Economic Co-operation and Development - Paris, France. doi:10.1787/9789264070127-en
- OECD, 2000. OECD TG 215. Fish, juvenile growth test, OECD Guideline for the Testing of Chemicals. Organisation for Economic Co-operation and Development - Paris, France.
- OECD, 1998. OECD TG 212. Fish, Short-term Toxicity Test on Embryo and Sac-fry Stages, OECD Guideline for the Testing of Chemicals. Organisation for Economic Co-operation and Development - Paris, France.
- Patisaul, H.B., Melby, M., Whitten, P.L., Young, L.J., 2002. Genistein affects ER beta- but not ER alpha-dependent gene expression in the hypothalamus. *Endocrinology* 143, 2189–2197. doi:10.1210/endo.143.6.8843
- Pinto, P.I.S., Estêvão, M.D., Power, D.M., 2014. Effects of estrogens and estrogenic disrupting compounds on fish mineralized tissues. *Mar. Drugs* 12, 4474–4494. doi:10.3390/md12084474
- Prossnitz, E.R.P., Arterburn, J.B., Sklar, L.A., 2007. GPR30: a G protein-coupled receptor for estrogen. *Mol. Cell. Endocrinol.* 138–142. doi:10.1016/j.str.2010.08.012.Structure
- Raimondo, S., Vivian, D., Barron, M., 2010. Web-based interspecies correlation estimation (Web-ICE) for acute toxicity: User Manual Version 3.1. United States Environmental Protection Agency, Gulf Breeze, Florida, USA.
- Russom, C.L., Bradbury, S.P., Broderius, S.J., Hammermeister, D.E., Drummond, R.A., 1997. Predicting modes of toxic action from chemical structure: acute toxicity in the fathead minnow (*Pimephales promelas*). *Environ. Toxicol. Chem.* 16, 948–967.
- Salvito, D.T., Senna, R.J., Federle, T.W., 2002. A framework for prioritizing fragrance materials for aquatic risk assessment. *Environ. Toxicol. Chem.* 21, 1301–1308. doi:10.1897/1551-5028(2002)021<1301:AFFPFM>2.0.CO;2
- Sanchez-Bayo, F., Goka, K., 2005. Unexpected effects of zinc pyrethrin and imidacloprid on Japanese medaka fish (*Oryzias latipes*). *Aquat. Toxicol.* 74, 285–293. doi:10.1016/j.aquatox.2005.06.003
- Sanderson, H., Johnson, D.J., Reitsma, T., Brain, R.A., Wilson, C.J., Solomon, K.R., 2004. Ranking and prioritization of environmental risks of pharmaceuticals in surface waters. *Regul. Toxicol. Pharmacol.* 39, 158–183. doi:10.1016/j.yrtph.2003.12.006
- Solomon, K.R., Brock, T.C.M., de Zwart, D., Dyer, S.D., Posthuma, L., Richards, S.M., Sanderson, H., Sibley, P.K., van den Brink, P.J., 2008. *Extrapolation Practice for Ecotoxicological Effect Characterization of Chemicals*, SETAC Pres. ed.
- Teather, K., Jardine, C., Gormley, K., 2005. Behavioral and sex ratio modification of Japanese Medaka (*Oryzias latipes*) in response to environmentally relevant mixtures of three pesticides. *Environ. Toxicol.* 20, 110–117. doi:10.1002/tox.20084
- Tebby, C., Mombelli, E., Pandard, P., Péry, A.R.R., 2011. Exploring an ecotoxicity

- database with the OECD (Q)SAR Toolbox and DRAGON descriptors in order to prioritise testing on algae, daphnids, and fish. *Sci. Total Environ.* 409, 3334–3343. doi:10.1016/j.scitotenv.2011.05.029
- Tillman, R.W., Siegel, M.R., Long, J.W., 1973. Mechanism of action and fate of the fungicide chlorothalonil (2,4,5,6-tetrachloroisophthalonitrile) in biological systems: I. Reactions with cells and subcellular components of *Saccharomyces pastorianus*. *Pestic. Biochem. Physiol.* 3, 160–167. doi:10.1016/0048-3575(73)90100-4
- Verhaar, H.J.M., van Leeuwen, C.J., Hermens, J.L.M., 1992. Classifying environmental pollutants. 1: Structure-activity relationships for prediction of aquatic toxicity. *Chemosphere* 25, 471–491.
- Wang, L., Waltenberger, B., Pferschy-Wenzig, E.M., Blunder, M., Liu, X., Malainer, C., Blazevic, T., Schwaiger, S., Rollinger, J.M., Heiss, E.H., Schuster, D., Kopp, B., Bauer, R., Stuppner, H., Dirsch, V.M., Atanasov, A.G., 2014. Natural product agonists of peroxisome proliferator-activated receptor gamma (PPARalpha): A review. *Biochem. Pharmacol.* 92, 73–89. doi:10.1016/j.bcp.2014.07.018
- Weyers, A., Sokull-Klüttgen, B., Baraibar-Fentanes, J., Vollmer, G., 2000. Acute toxicity data: a comprehensive comparison of results of fish, *Daphnia*, and algae tests with new substances notified in the European Union. *Environ. Toxicol. Chem.* 19, 1931–1933.
- Zhang, X.J., Qin, H.W., Su, L.M., Qin, W.C., Zou, M.Y., Sheng, L.X., Zhao, Y.H., Abraham, M.H., 2010. Interspecies correlations of toxicity to eight aquatic organisms: Theoretical considerations. *Sci. Total Environ.* 408, 4549–4555. doi:10.1016/j.scitotenv.2010.07.022
- Zvinavashe, E., Du, T., Griff, T., Berg, H.H.J. van den, Soffers, A.E.M.F., Vervoort, J., Murk, A.J., Rietjens, I.M.C.M., 2009. Quantitative structure-activity relationship modeling of the toxicity of organothiophosphate pesticides to *Daphnia magna* and *Cyprinus carpio*. *Chemosphere* 75, 1531–1538. doi:10.1016/j.chemosphere.2009.01.081

## List of abbreviations and definitions

ACR	Acute to chronic ratio
AchE	Acetylcholinesterase
AOP	Adverse Outcome Pathway
CAS No	Chemical Abstracts Service Number
EC	European Commission
ECETOC	European Centre for Ecotoxicology and Toxicology of Chemicals
ECHA	European Chemicals Agency
ECx	The concentration of a chemical showing effect in x% of the test species during a given observation period
EMA	European Medicines Agency
EU	European Union
EURL ECVAM	European Union Reference Laboratory for Alternatives to Animal Testing
FELS	Fish Early-Life Stage test
ICE	Interspecies Correlation Estimation
JRC	Joint Research Centre
KE	Key Event
LC <sub>50</sub>	The lethal concentration of a chemical killing 50% of the test species during a given observation period
LOEC	Lowest Observed Effect Concentration
MIE	Molecular Initiating Event
MOA	Mode of Action
NOEC	No Observed Effect Concentration
OECD	Organisation for Economic Co-operation and Development
PEC	Predicted Environmental Concentration
PNEC	Predicted No Effect Concentration
QAARs	Quantitative Activity-Activity Relationships
QSARs	Quantitative Structures Activity Relationships
REACH	Registration, Evaluation, Authorisation and Restriction of Chemicals
ROUT	Robust regression and OUTlier removal
SR <sub>ac</sub>	Acute Sensitivity Ratio
SR <sub>ch</sub>	Chronic Sensitivity Ratio
SVHCs	Substances of Very High Concern
TG	Test Guideline
TTC	Threshold of Toxicological Concern
US EPA	US Environmental Protection Agency

## List of figures

Figure 4.1: Relationships between <i>Daphnia</i> 48h LC <sub>50</sub> and fish NOEC values from the OECD QSAR Toolbox 2.3 (n=246). a: Lowest value approach, b: Geometric mean approach. The dashed lines represent the 95% confidence interval. <i>Italics</i> : CAS number of the outliers. ....	12
Figure 4.2: Relationships between <i>Daphnia</i> 21-days NOEC and fish NOEC values from the OECD QSAR Toolbox 2.3 (n=193). a: Lowest value approach, b: Geometric mean approach. The dashed lines represent the 95% confidence interval. <i>Italics</i> : CAS number of the outliers. ....	13
Figure 4.3: Sensitivity ratio derived with acute <i>Daphnia</i> data versus chronic fish data (blue bars) and chronic <i>Daphnia</i> data versus chronic fish data (red bars) in % of chemicals of the whole dataset; i.e. 34.7% of the chemicals demonstrates a higher chronic toxicity in <i>Daphnia</i> compared to NOEC fish and values range within a factor of 10; for 39% of the chemicals, the acute <i>Daphnia</i> LC <sub>50</sub> indicate a lower sensitivity compared to chronic NOEC fish and values range within a factor of 10. ....	15
Figure 4.4: Aniline and its derivatives. ....	17
Figure 4.5: Relationships between <i>Daphnia</i> LC <sub>50</sub> and fish NOEC values by MOA. The dashed lines represent the 95% confidence interval. <i>Italics</i> : CAS number of the outliers. ....	19
Figure 4.6: Relationships between <i>Daphnia</i> NOEC and fish NOEC values by MOA. The dashed lines represent the 95% confidence interval. <i>Italics</i> : CAS number of the outliers. ....	20
Figure 4.7: Distribution of the chronic sensitivity ratios between <i>Daphnia</i> NOEC and fish NOEC according to their MOA. ....	21
Figure 4.8: Relationships between acute LC <sub>50</sub> fish data and fish NOEC values from the OECD QSAR Toolbox 2.3 (n=240). a: Lowest value approach, b: Geometric mean approach. The dashed lines represent the 95% confidence interval. <i>Italics</i> : CAS number of the outliers. ....	22
Figure 4.9: Acute to chronic fish toxicity ratio distribution. ....	25
Figure 4.10: Structural similarity between genistein and 17β-oestradiol. ....	25
Figure 4.11: Relationships between fish LC <sub>50</sub> and fish NOEC values by MOA. The dashed lines represent the 95% confidence interval. <i>Italics</i> : CAS number of the outliers. ....	29
Figure 4.12: Distribution of ACR (acute to chronic fish toxicity ratio) of chemicals according to their MOA. ....	30

## List of tables

Table 2.1: Verhaar Classification Scheme for aquatic modes of action (MOA). .....	7
Table 4.1: Linear relationships between <i>Daphnia</i> acute or chronic toxicity data and fish chronic toxicity data.....	11
Table 4.2: Name and MOA of outliers.....	13
Table 4.3: Substances with a high SRch.....	16
Table 4.4: Linear relationships between <i>Daphnia</i> acute or chronic toxicity and fish chronic toxicity data depending on the MOA.....	18
Table 4.5: Linear relationships between acute fish data and chronic fish data. ....	22
Table 4.6: Name and MOA of outliers of the acute to chronic relationships on fish data.....	23
Table 4.7: Name and MOA of chemicals with ACR < 1.....	24
Table 4.8: Name and MOA of chemicals with ACR >100. ....	26
Table 4.9: Linear relationships between fish LC <sub>50</sub> and fish chronic toxicity data depending on the MOA. ....	28

## Appendix I. Examples of EU regulatory information requirements for *Daphnia* and/or fish toxicity

Regulatory framework	Endpoints			
	Short-term aquatic toxicity		Long-term toxicity	
	Invertebrates ( <i>Daphnia spp</i> )	Fish	Invertebrates ( <i>Daphnia spp</i> )	Fish
<b>Industrial chemicals (REACH)</b> Regulation (EC) No. 1907/2006	> 1 t/year (Annex VII)  Not to be conducted if: <ul style="list-style-type: none"> <li>- aquatic toxicity is unlikely to occur, e.g. chemical highly insoluble in water or unlikely to cross biological membranes or</li> <li>- a long-term aquatic toxicity study on invertebrate is available or</li> <li>- adequate information for environmental classification and labelling is available</li> </ul> The long-term aquatic toxicity study on <i>Daphnia</i> (Annex IX, section 9.1.5) shall be considered if the substance is poorly water soluble.	> 10t/year (Annex VIII)  Not to be conducted if: <ul style="list-style-type: none"> <li>- aquatic toxicity is unlikely to occur, e.g. chemical highly insoluble in water or unlikely to cross biological membranes or</li> <li>- a long-term aquatic toxicity study on fish is available</li> </ul> In general applicants are invited to consider long-term aquatic toxicity testing as described in Annex IX <ul style="list-style-type: none"> <li>- if the chemical safety assessment according to Annex I indicates the need to investigate further effects on aquatic organisms.</li> <li>- if the substance is poorly water soluble.</li> </ul>	> 100t/year (Annex IX)  Long-term toxicity testing shall be proposed by the registrant if the chemical safety assessment according to Annex I indicates the need to investigate further the effects on aquatic organisms. The choice of the appropriate test(s) depends on the results of the chemical safety assessment.	Long-term toxicity testing on fish, (unless already provided as part of Annex VIII requirements). To be provided with one of the following tests: <ul style="list-style-type: none"> <li>- Fish early-life stage (FELS) toxicity test (OECD TG210);</li> <li>- Fish short-term toxicity test on embryo and sac-fry stages (OECD TG212) or</li> <li>- Fish, juvenile growth test (OECD TG215)</li> </ul>
<b>Cosmetic ingredients</b> Regulation (EC) No 1223/2009	Recital (5): The environmental concerns that substances used in cosmetic products may raise are considered through the application of Regulation (EC) No 1907/2006 of the European Parliament and of the Council of 18 December 2006 concerning the Registration, Evaluation, Authorisation and Restriction of Chemicals (REACH) and establishing a European Chemicals Agency(4) OJ L 396, 30.12.2006, p. 1. (4), which enables the assessment of environmental safety in a cross-sectoral manner.			

Regulatory framework	Endpoints			
	Short-term aquatic toxicity		Long-term toxicity	
	Invertebrates ( <i>Daphnia spp</i> )	Fish	Invertebrates ( <i>Daphnia spp</i> )	Fish
<b>Plant protection products data requirements - active substance</b> Regulation (EU) No 283/2013	<i>Active substance:</i> 24h or 48h acute toxicity with <i>Daphnia magna</i>  For active substances with an insecticidal mode of action or which show insecticidal activity a second species shall be tested, for example Chironomid larvae or Mysid shrimps (24 h or 48h acute toxicity)	<i>Active substance:</i> Threshold approach (OECD GD 126) or OECD TG203 Fish, acute toxicity (rainbow trout)	<i>Active substance:</i> A long-term or chronic toxicity study on one aquatic invertebrate species (e.g. <i>Daphnia magna</i> reproduction) - if exposure of surface water likely and the substance stable in water; If the active substance is an insect growth regulator, an additional study on chronic toxicity shall be carried out using relevant non-crustacean species such as <i>Chironomus spp.</i>	<i>Active substance:</i> Fish early life stage (FELS) toxicity test (OECD TG210) - if exposure of surface water likely and the substance stable in water; Fish full life cycle test (US EPA OCSP 850.1500) may be required
<b>Plant protection products data requirements - product</b> Regulation (EU) No 284/2013	<i>Product:</i> Above listed requirements apply only if: <ul style="list-style-type: none"> <li>- the toxicity cannot be predicted from the active substance(s)</li> <li>- the intended use includes direct application on water</li> <li>- extrapolation on the basis of available data for a similar plant protection product is not possible</li> </ul>		<i>Product:</i> Above listed requirements apply only if the toxicity cannot be predicted from the active substance(s) or the product is more acutely toxic than the active substance as manufactured by a factor of 10.  Necessary studies should be discussed with competent authority	
<b>Biocidal products</b> Regulation (EU) No 528/2012 <b>ECHA Guidance on data requirements (ECHA, 2013)</b>  Only to be carried out if exposure is likely.  <i>Note for Products:</i> Testing on the product itself only when valid data on components are missing or synergistic effects to be expected	<i>Active substance:</i> <i>Core data set</i> EC method C.2 - <i>Daphnia sp.</i> Acute Immobilisation Test) or the corresponding OECD TG202 - <i>Daphnia sp.</i> Acute Immobilisation Test  Testing may be omitted if results are available from any non-standard test protocols, also with a different invertebrate species.	<i>Active substance:</i> <i>Core data set</i> Short-term toxicity testing on fish - When short-term fish toxicity data is required the threshold approach (OECD GD126) should be applied.  The study does not need to be conducted if a valid long-term aquatic toxicity study on fish is available.	<i>Active substance:</i> <i>Additional data set</i> depending on the results of the studies on fate/behaviour and intended use, e.g. risk to the aquatic environment, and long-term exposure  Long term toxicity testing on invertebrates may be required, e.g. <i>Daphnia magna</i> Reproduction Test (OECD TG211 or other reproduction and growth tests on aquatic invertebrates	
			Long-term toxicity studies on fish may be required, one or two tests, OECD TGs 210, 212, 215 or fish full life cycle test	

Regulatory framework	Endpoints			
	Short-term aquatic toxicity		Long-term toxicity	
	Invertebrates ( <i>Daphnia spp</i> )	Fish	Invertebrates ( <i>Daphnia spp</i> )	Fish
<b>Veterinary Medicinal Products</b> Stepwise approach with initial screening (Phase I) to identify exposure, bioaccumulation, persistence. If given then studies are performed (Phase II). Phase II: CVMP/VICH/790/03-FINAL; EMEA, 2004) (corresponds to VICH GL38)	<i>Phase II:</i> Tier A – aquatic effect study OECD TG202 - <i>Daphnia</i> sp. acute immobilisation test	<i>Phase II:</i> Tier A – aquatic effect study - OECD TG203 Fish, acute toxicity	<i>Phase II:</i> Tier B - if risk quotient (PEC <sub>refined</sub> /PNEC) > 1 for <i>Daphnia</i> , OECD TG211 - <i>Daphnia magna</i> reproduction test	<i>Phase II:</i> Tier B - if risk quotient (PEC <sub>refined</sub> /PNEC) > 1 for fish, OECD TG210 - Fish early life stage test
<b>Human Medicinal Products</b> Stepwise approach with initial screening (Phase I) to identify exposure, bioaccumulation, persistence. If given then studies are performed (Phase II) Phase II: CHMP/SWP/4447/00 (EMA, 2006)	Not required	Not required	<i>Phase II:</i> Tier A – base set requirement OECD TG211 - <i>Daphnia</i> sp. reproduction test	<i>Phase II:</i> Tier A – base set requirement OECD TG210 – Fish early life stage test
<b>Feed additives</b> Regulation (EC) No 429/2008 (EC, 2008a) & (EFSA, 2008) Stepwise approach with initial determination (Phase I) whether a significant environmental effect of the additive is likely (based on estimated PEC). If likely, then studies are performed (Phase II)	<i>Phase IIA:</i> OECD TG202 - <i>Daphnia</i> sp. acute immobilisation test	<i>Phase IIA:</i> OECD TG203 Fish, acute toxicity	<i>Phase IIB:</i> If an environmental risk cannot be excluded following Phase IIA assessment, the refinement of the effect assessment for water/sediment may be based on chronic toxicity tests on the most sensitive aquatic/benthic organisms identified in Phase IIA assessment, e.g.  OECD TG211- <i>Daphnia</i> sp. reproduction test OECD TG210 – fish early life stage test	
<b>CLP Regulation</b> Regulation (EC) No 1272/2008 EC (2008b)	In the EU, the core classification system for aquatic environmental hazards consists of one acute (Acute 1) and three chronic (Chronic 1 – 3) hazard classification categories (EC, 2008). If adequate chronic toxicity data are not available, a combination of acute aquatic toxicity data and information on the environmental fate (degradability and bioaccumulation) is used to decide on the appropriate Chronic category. In addition, a "safety net" classification referred to as category Chronic 4 can be applied for substances which cannot be classified in any of the four categories but nevertheless raise some concerns.			



Europe Direct is a service to help you find answers to your questions about the European Union  
Free phone number (\*): 00 800 6 7 8 9 10 11  
(\* ) Certain mobile telephone operators do not allow access to 00 800 numbers or these calls may be billed.

A great deal of additional information on the European Union is available on the Internet.  
It can be accessed through the Europa server <http://europa.eu>

### **How to obtain EU publications**

Our publications are available from EU Bookshop (<http://bookshop.europa.eu>),  
where you can place an order with the sales agent of your choice.

The Publications Office has a worldwide network of sales agents.  
You can obtain their contact details by sending a fax to (352) 29 29-42758.

## JRC Mission

As the Commission's in-house science service, the Joint Research Centre's mission is to provide EU policies with independent, evidence-based scientific and technical support throughout the whole policy cycle.

Working in close cooperation with policy Directorates-General, the JRC addresses key societal challenges while stimulating innovation through developing new methods, tools and standards, and sharing its know-how with the Member States, the scientific community and international partners.

*Serving society  
Stimulating innovation  
Supporting legislation*

