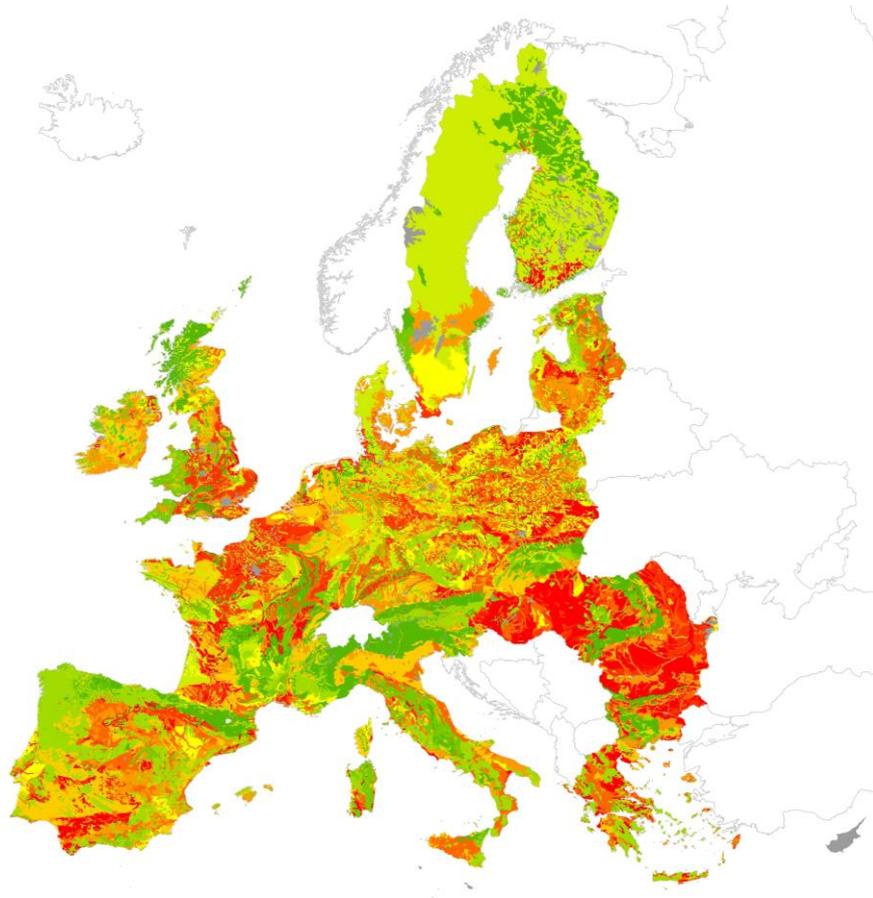


JRC TECHNICAL REPORTS

Mapping the storing and filtering capacity of European soils

András Makó, Mihály Kocsis,
Gyöngyi Barna, Gergely Tóth



2017

EUR 28392 EN

This publication is a Technical report by the Joint Research Centre (JRC), the European Commission's science and knowledge service. It aims to provide evidence-based scientific support to the European policymaking process. The scientific output expressed does not imply a policy position of the European Commission. Neither the European Commission nor any person acting on behalf of the Commission is responsible for the use that might be made of this publication.

JRC Science Hub

<https://ec.europa.eu/jrc>

JRC105121

EUR 28392 EN

PDF	ISBN 978-92-79-64939-4	ISSN 1831-9424	doi:10.2788/49218
Print	ISBN 978-92-79-64940-0	ISSN 1018-5593	doi:10.2788/156875

Luxembourg: Publications Office of the European Union, 2017

© European Union, 2017

The reuse of the document is authorised, provided the source is acknowledged and the original meaning or message of the texts are not distorted. The European Commission shall not be held liable for any consequences stemming from the reuse.

How to cite this report: Makó, A., Kocsis, M., Barna, GY., Tóth, G.; *Mapping the storing and filtering capacity of European soils*; EUR 28392; doi:10.2788/49218

All images © European Union 2017

Mapping the storing and filtering capacity of European soils

Authors

András Makó	Hungarian Academy of Sciences, Centre for Agricultural Research
Mihály Kocsis	University of Pannonia, Georgikon Faculty
Gyöngyi Barna	Hungarian Academy of Sciences, Centre for Agricultural Research
Gergely Tóth	European Commission, DG Joint Research Centre

Table of contents

Abstract.....	1
1. Introduction	2
2. Working approach and methodology	6
2.1 Identifying the groups of substances	6
2.2 The clay content of the soil profile.....	6
2.3 Humus content of the soil profile and the humus quality	7
2.4 Estimating the environmental factors	8
2.5 Estimating the cation exchange capacity and specific surface area of soils	8
2.6 Calculating the sorption factors	9
2.7 Storing and filtration capacity codes	10
2.7.1 The cation storing capacity	10
2.7.2 The cation filtering capacity	10
2.7.3 The anion storing capacity	11
2.7.4 The anion filtering capacity.....	11
2.7.5 The solids and pathogenic microorganisms storing capacity	11
2.7.6 The solids and pathogenic microorganisms filtering capacity	11
2.7.7 The non-polar organic chemicals storing capacity	12
2.7.8 The non-polar organic chemicals filtering capacity	12
2.7.9 The nonaqueous phase liquids (NAPLs) storing capacity	12
2.7.10 The nonaqueous phase liquids (NAPLs) filtering capacity	12
3. Presenting the soil storing and filtering capacity maps	15
4. Conclusions	29
References.....	30
List of abbreviations and definitions	33
List of figures	54

Abstract

The purpose of the current study was to develop an estimation method based on available soil map information (categorical data) and thereafter estimate and map across Europe the soil filtering and storing capacity of different substances, which support ecosystem services and those which present a hazard to the ecosystem functioning. Pollutants were grouped as follows: (1) elements in *cationic form*; (2) elements in *anionic form*; (3) *solids and pathogenic microorganisms*; (4) *non-polar organic chemicals* and (5) *nonaqueous phase liquids (NAPLs)*.

The spatial pattern of areas with high soil storing capacity are largely different depending on the stored substances. In general, those soils are characterized by good storing capability that have thick topsoil and subsoil layers and these layers are free from the effects of groundwater. As the content of clay and humus content increases and the stone or gravel content decreases, the storage capacity increases simultaneously. However, the effect of soil pH and the soil mineralogy is different depending on the various groups of pollutants (e.g. the calcareous soils with high swelling clay mineral content are able to bind more cations, whereas the acidic soils or the soils with high sesquioxide content have higher anion storing capacity).

The soil filtering capacity pattern of Europe in the case of different groups of substances are in part similar to the storing capacity pattern. The main difference between the two parameters was to take account of infiltration rate and the thickness of the filtration path up to the groundwater during the filtering capacity estimation.

1. Introduction

Soil supports ecosystem services by performing its function to filter and store substances and there is a requirement to increase awareness of the role of soil in preserving ecosystem services in Europe. Creating a map of the capacity for filtering and storing functions of different soils across the climatic conditions of Europe is one element of this action. In this first step, maps for the European Union were prepared, including area of its 25 Member States for which all data input was available (all EU Member States except Croatia, Cyprus and Malta).

The soil is a natural body. It has developed over a long time in a place through natural processes (weathering, humification, leaching etc.) under natural conditions (climate, organism, water, topography) from natural materials (parent materials). Soils however, continually change at different rates and along different pathways and are never stationary for more than a short periods of time. The changes are physical, chemical, and biological (biogeophysical, biogeochemical) and may and affect all soil phases. Despite these changes, the state of the soil always tends to remain at equilibrium with the environment it is exposed to. Thus, there are always complex interaction between the soil and environment [1–3].

When water, rain or wastewater falls on soil, some is evaporated and transpired, some runs off, and the most part infiltrates. Soil stores some water, but it is a leaky reservoir that loses water downward by seepage and upward by evaporation. Thus a considerable amount of water passes through the soil profile into the groundwater. During temporary storage and transmittance of this water in soil, the sediments may deposit in soil matrix, the possible suspended or dissolved contaminants may remove by reactions with soil constituents or exchange in the colloidal phase, and the purified water enters into the groundwater or into the surface waters. In a healthy soil ecosystem soil reduce the impacts of pollution by buffering, detoxifying, and decomposing potential. Quality of water in the watershed largely depends of the quality of soil around. So, soil acts as a natural filter medium, in which contaminants, pathogens and toxins that might otherwise foul the terrestrial environment are rendered harmless [4–6].

Modern society, however, generates ever greater quantities of potentially toxic materials and waste products, some of which do not occur naturally. Pollution is defined as the introduction of any substance to the environment that adversely affects the usefulness of a medium. Pollution and contamination are used mainly synonymously to mean the introduction into the environment by human of substances that are harmful or poisonous to people or ecosystems. Soil pollution is a global problem that threatens the life of microorganisms, plants, animals, and humans [7; 8].

Soil can filter pollutants (agrochemicals or other substances), whose properties and concentrations are potentially hazardous to the environment and/or human health. This filtering function of soil prevents hazardous materials from reaching groundwater. In the filtering process, substances are filtered out of percolation water and are bound by surfaces of solid phase (mainly by soil colloids: clay minerals, oxides, humic materials) in the soil. In the buffering process, which can be associated with the filtering, soil neutralizes acidifying impacts, thus preventing exposure of hazardous materials, eutrophication and a pH decrease. The extent to which substances can be deposited and bound is determined by soil *storing capacity*. Once the soil's *filtering and buffering capacity* is exceeded, the soil releases the stored pollutants, which end up in the groundwater or are uptaken by plants – and thus may enter the food chain [9; 10].

Soils vary greatly in their filtering, storing and buffering capacity. Under certain conditions, substances may take months or years to move from the land surface to the groundwater. In other cases, they can flow almost directly into the groundwater. The understanding of the interaction of soil characteristics (and related natural, e.g. climatic, hydrologic conditions) is essential in estimating the soil filtering, storing and buffering properties and determining the potential for groundwater (and connected surface waters) pollution in a given situation [11–14].

Soil pollution arises mainly from disposal of solid and liquid wastes in the ground; disposal of chemical wastes from the industries into the soil; indiscriminate use of agriculture chemicals, including fertilizers and pesticides; application sewage sludge and compost without judging metal or trace organic contamination; leakage of pipelines or

underground tanks into the soil; mining activity; irrigation with polluted water etc. Frequently, pollutants are introduced into the food chain via plants that have taken them up from soil or the atmosphere, and these pollutants often cause irreversible damage to individual organisms or to entire communities as a result of accumulation and exclusion processes. Therefore highest priority on the control of the influence on soil chemistry and microbial activities has to be given in the future [7; 15].

In recent decades interest has arisen in the possibilities and limitations of applying various waste materials to the land so as to utilize the soil's ability to filter, retain, buffer, immobilize, decompose, or otherwise mitigate the hazards of polluting agents. The soils have a significant but limited capacity to make harmless pollutants. The purification processes involved include physical transport phenomena, as well as complex chemical and microbiological interactions that take place primarily in the upper layer of soil [7].

The soil's ecological functions are among the others the substance storing, filtering, buffering and transformation capacity. These functions become increasingly important, because of the above mentioned increasing pollution effect. Soils may react through mechanical filtration, physical or chemical absorption and precipitation on its inner surfaces, or microbiological and biochemical mineralization and metabolization. As long as these filtering, storing, buffering and transformation capacities can be maintained, there is no danger to the groundwater or to the food chain. However, these capacities of soils are limited and vary according to specific soil and pollutant conditions [16].

The purpose of the current study is to assist with the implementation of the research programme on soil quality indicators, namely to facilitate the completion of a new soil quality indicator by supplying reliable spatial data on the soil's function to filter and store substances.

The objective of this study is to estimate and map the soil filtering and storing capacity of different substances across Europe.

Scope of the study includes the modelling and mapping of behaviour of those groups of substances which support ecosystem services (nutrients) and those which present a hazard to the ecosystem functioning (overused nutrients and pollutants). Literature review of this subject has shown that the following *problem substances* occur during the contamination of soil/subsurface/groundwater/surface water systems:

- Nutrients,
- Organics and Pesticides,
- Nonaqueous Phase Liquids (NAPLs),
- Pathogenic Microorganisms,
- Metals,
- Solids (Suspended and Dissolved).

Nutrients

Nitrogen- and phosphorus-containing compounds are often found in the water infiltrated into the soil. Farmers and homeowners apply fertilizers to the soil to promote plant growth. Nitrates result both from vehicular exhaust on the road and from fertilization of agricultural soil. Infiltrating and percolating water also contains phosphorus from motor oils, fertilizers, bird droppings, and animal remains [7; 8].

Organics and Pesticides

Although organics are also commonly found in infiltrating and percolating water of agricultural areas, runoff from *industrial areas* has been shown to contain higher concentrations of certain organics, such as pentachlorophenol, bis(2-ethylhexyl) phthalate or the polycyclic aromatic hydrocarbons.

Thousands of pesticide and pesticide formulations exist. They are classified on the basis of the organisms they attack (insecticides, herbicides, fungicides, nematocides, rodenticides etc.) and on the basis of the chemical structure (e.g. chlorinated hydrocarbons, organophosphates, carbamates etc.). The wide variation in chemical structure of pesticides results in the wide variation in their behaviour in soil and groundwater [8; 17].

Nonaqueous Phase Liquids (NAPLs)

In general, water is the primary liquid in soil. However, NAPLs released during spills, leaks, and tank or pipe ruptures may enter soil and replace water. The basic principles governing the migration of water in soil are generally the same ones governing of NAPLs. NAPLs are generally not miscible with water, therefore their distribution in soil in presence of water should be addressed. The term of NAPLs is used to refer to unrefined crude oil, refined petroleum (e.g. gasoline, kerosene), and petrochemicals (e.g. TCA, TCE) [8; 18; 19].

Pathogenic Microorganisms

Urban, agricultural or industrial runoff has been found to contain different pathogens. These pathogens include, but are not limited to, *Salmonella*, *Pseudomonas aeruginosa*, *Shigella*, or *enteroviruses*. *Shigella* species causing bacillary dysentery are one of the primary human-enteric-disease-producing bacteria present in water. *Pseudomonas* is reported to be the most abundant pathogenic bacteria organism in urban runoff and streams. Pathogenic *E. coli* can also be commonly found in urban runoff. *Viruses* also may be important pathogens, very small viral concentrations are capable of producing infections or diseases, especially when compared to the large numbers of bacteria organisms required for infection.

Infiltration will increase bacterial and viral penetration into the soil profile. Like the organics, the greatest chance for contamination occurs when the distance to the groundwater is small. Most, but not all, pathogens are usually filtered out or inactivated during percolation through the soil [20].

Metals

The heavy metals of most concern in infiltrating and percolating water are lead, zinc, copper, nickel, and chromium. Most of these heavy metals have very low solubilities at the typical pH of receiving waters. They either are removed by sediment adsorption or are organically complexed with other particulates and are easily removed during filtration. Metals in urban runoff originate both at industrial sites and on highways, etc., as part of the exhaust and other residue left by vehicular use. Metals seem to be more prevalent in runoff from industrial areas, although they are also commonly found in infiltrating and percolating water of agricultural areas [8; 21].

Solids

Suspended solids are of concern in runoff because of their ability to clog infiltration areas. During percolation, the suspended and colloidal particles that were not stopped at the surface travel downward until they are trapped by pores that are smaller than they are. Fine to medium textured soils remove essentially all of the suspended solids by straining, while coarse textured soils allow deeper penetration of these particles. *Dissolved solids* are in urban runoff due to the use of salt to de-ice roads in the winter and due to fertilizer and pesticide salts from the use of those items [10].

The relative magnitude of the filtering and storing capacity may vary considerably, based on a number of factors (Fig. 1). *Climate* (quantity, form and distribution of precipitation, temperature etc.), *topography*, *plant cover* and *land use* have been identified as important factors affecting infiltration and runoff. *Impervious surfaces*, such as compacted soil layers or roads, reduce infiltration of rainfall into the ground [8].

Filtration can be defined as an interaction between the solution (or suspension) and the filtering soil material. Pollutants are removed from the solution when they become attached to the media or to previously captured particles. In general, the *key properties* of the soils are *infiltration and percolation rate*; *liquid phase retention capacity*; *surface area (surface-substances interactions: physical straining ability, adsorption/ion-exchange ability)*; *aerobic/anaerobic condition*; *soil depth to the groundwater and the different (chemical and/or microbiological, biological) transformation processes*. The most important physical and chemical soil parameters that influence the above properties are *soil texture*

(clay content), *gravel or rock content*, *soil mineralogy* (especially type of clay minerals and oxides/hydrous oxides), *organic matter content and pH*. Due the lack of information about these properties or parameters there are often necessary to estimate them. Knowing the *soil types*, the topography or the geological origin there is possible to assess some missing information [22; 23].

The *liquid phase properties* can also affect the filtration and storing capacity. The most important properties are *polarity, viscosity, density* and the *type and concentration of substances* in liquid phase. Density and viscosity are both temperature dependent, and density will also depend upon the concentration of dissolved solids in the water [18; 19].

The above aspects of filtering and storing will be modelled. *Figure 1* summarises the most relevant soil processes, properties and natural conditions which are the factors of the soil filtration and storing capacity.

However, this study will not assess the effect of (chemical, biological and microbiological) *transformation* processes on the substances filtering and storing capacity of the soils.

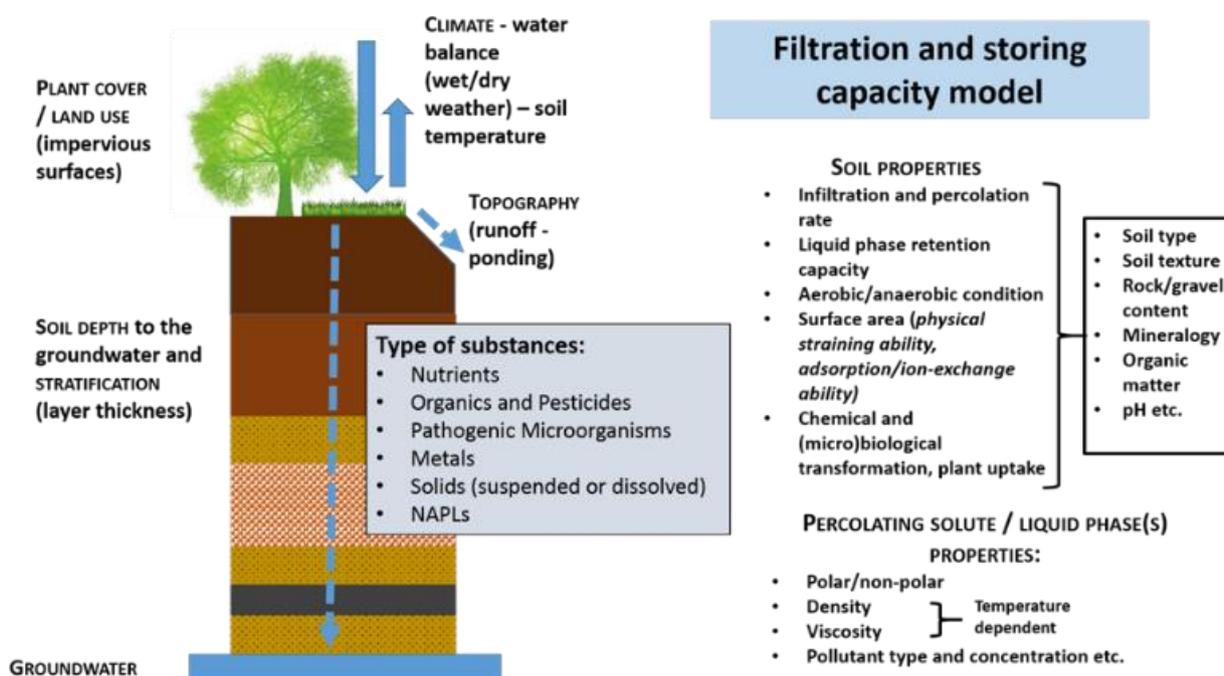


Figure 1. The proposed filtration and storing capacity model

2. Working approach and methodology

The geographical extent of the study includes 27 Member States of the European Union (EU). Data availability is one of the most important factors to estimate the planned soil function parameters. As far as the input data in soil maps, databases and related land use, climate etc. information are available and accurate, so can be used the developed estimation methods. To estimate the filtering and storing capacity the following European databases were available and usable as input data:

- *The European Soil Database v.2.0* [24];
- *Pedotransfer Rules database* (PTRDB) [25];
- *CORINE LC* database to obtain land use/cover data [26];
- *Climate data* (agro-climatic zones, grouped agro-climatic areas) [27].

The European Soil Database did not contain all soil properties that affect the substances filtering and storing capacity. Using the existing pedotransfer rules of the Pedotransfer Rules database and based on expert estimates the missing soil parameters (e.g. infiltration and percolation rate, soil mineralogy etc.) were calculated.

The steps of the estimation and mapping procedure associated with the soil storing and filtering capacity were as follow: in the first step the major groups of substances and the associated most relevant natural and soil properties were identified, together with their effects on filtering and storing capacity. In the next step of the capacity assessments an evaluation algorithm was created for each substances groups (identifying ranges of values of relevant soil properties and natural conditions and estimating the potential of soil types to substance filtering and storing). Finally, filtering and storing capacity maps of European soils were created for each substances groups.

2.1 Identifying the groups of substances

In the introduction the problem substances (occur during the contamination of soil/subsurface/groundwater/surface water systems) were summarised as follows: nutrients, organics and pesticides, nonaqueous phase liquids (NAPLs), pathogenic microorganisms, metals and solids. However, within these groups occur materials with different properties. Nutrients in anionic (phosphate or nitrate ions) and cationic (potassium ion) form behave quite differently in soils. Heavy metals also behave differently depending on the charging conditions. But the organics and pesticides form also a heterogeneous group, since they can also have a positive or negative charge or may be also non-polar. However, microbes and solid pollutants are similar in terms of soil storing and filtrating capacity. Therefore, the pollutants were regrouped:

- (1) elements in *cationic form* (e.g. K^+ , Na^+ , Ca^{2+} , Fe^{2+} , Fe^{3+} or heavy metals, as Ba^{2+} , Cd^{2+}) and organic chemicals with positive charges (e.g. organoamines, metallo-organics);
- (2) elements in *anionic form* (e.g. nutrients, as phosphate, nitrate; heavy metals, as molybdate, arsenate or other elements, as borate, sulfate, halides) and organic chemicals with negative charges (e.g. organic chemicals possess hydroxyl or carboxyl functional groups);
- (3) *solids and pathogenic microorganisms*;
- (4) *non-polar organic chemicals* and
- (5) *nonaqueous phase liquids* (NAPLs).

2.2 The clay content of the soil profile

The storing and filtering capacity of soils were estimated for the average 1.5 m depth soil profile. There were two exceptions to this rule: (1) the solid rock – as parent material – within 1.5 m and (2) water saturated soil layer within 1.5 m. (The solid rock is unable to perform the storing and filtering functions, because it does not pass through the infiltrating soil solution and the water saturated soil layer is continuously leached due to the groundwater which prevents the storage and filter properties of the soils.) Therefore firstly

the “effective soil depth” was estimated, i.e. the *maximum depth of the soil profile with storage and filter properties (MDSP)*.

Based on the above, it depends on the depth to the impermeable solid rock (DISR) and the depth to the groundwater (DG). DISR can be estimated on the basis of the soil parent material (PARMADO and PARMASE) and the depth to rock (DR) [28]. DG is predictable according to the soil classification (Full soil code of the soil taxonomy unit (STU) from the World Reference Base (WRB) for Soil Resources: WRBFU [29].

It was supposed, that the soil storing and filtering properties are generally dependent on the soil clay and humus content (mostly they represent the soil colloids) [7]. As a first step therefore the clay content of the soil profile (SPCC) was estimated. Prior to this, however, some auxiliary parameters had to be determined. Because of the soil profile can contain soil layers with different soil texture, we needed information about the stratification. The modified depth to a textural change code (MTDCH) shows the estimated depth where occurs a textural change between the topsoil and subsoil. The MTDCH can be determined according to the textural change information of ESDB (TEXTDEPCHG) and the “effective soil depth” (MDSP). (The textural change is taken into account in the case where MDSP was deeper as TEXTDEPCHG. Otherwise, the textural change was not considered.)

The *clay percentage of the topsoil (CPT) and subsoil (CPS)* were estimated on the basis of the dominant and secondary surface textural class of the STU (TEXTSRFDOM and TEXTSRFSEC) or on the basis of the dominant and secondary sub-surface textural class of the STU (TEXTSUBDOM and TEXTSUBSEC) [4]. In the case when the TEXTSRFDOM and TEXTSRFSEC or TEXTSUBDOM and TEXTSUBSEC did not match, weighted average code was calculated. If the secondary surface or sub-surface textural class of the STU was peat, it was not considered.

The *modified clay percentage of the topsoil (MCPT) and subsoil (MCPS)* was estimated from the CPT and CPS, the volume of stones (VS) and the topsoil or subsoil packing density (PD_TOP or PD_SUB). (The higher is the stone content and the lower is the packing density, the more reduced the relative clay amount per unit soil volume.) It was assumed that VS is unchanged throughout the soil profile [4; 7]. Finally, the **clay content of the soil profile (SPCC)** was estimated on the basis of the MCPT, MCPS, MTDCH and MDSP.

2.3 Humus content of the soil profile and the humus quality

In the next step the **humus content of the soil profile (SPHC)** was estimated. To do this, we had to determine previously the *thickness of the humus layer (HLT)*. This parameter was estimated on the basis of soil classification (WRB_GRP: Soil reference group code of the STU from the World Reference Base (WRB) for Soil Resources [29]. If the “effective soil depth” (MDSP) was lower than the characteristic humus layer thickness of a WRP group, the latter overwrote the HLT category code.

To estimate the humus content of the soil profile (SPHC) the following information of the ESDB were used: (1) OC_TOP = topsoil organic carbon content, (2) PEAT= peat, (3) VS = volume of stones and (4) PD_TOP = topsoil packing density. The clay percentage of the topsoil (CPT) and subsoil (CPS) provided additional information about the peaty layers of the soil profile. In case of peat soils or peaty soil layers high humus content were assumed. In other cases the humus content was estimated from the topsoil organic carbon content (OC_TOP).

The VS and the PD_TOP might alter the humus content: the higher is the stone content and the lower is the packing density, the more reduced the relative humus content per unit soil volume. (It was also assumed that VS is unchanged throughout the soil profile.) Finally, the SPHC code was estimated taking into account the HLT [7; 30]. The storing and filtering properties of soil humus content is highly dependent on the **humus quality (HQ)**. The quality of the humic substances of the soils is predictable on the basis of soil taxonomy (WRB_GRP). There were distinguished three categories: soils containing predominantly (1) fulvic acids (2) humic acids and (3) peaty, partially decomposed materials [7; 30].

2.4 Estimating the environmental factors

The storing and filtration capacity of soil depends on different **environmental factors**, such as climate and topography. The *water balance (WB) of the soil profile* is predictable on the basis of the average annual rainfall and temperature [4]. To perform this prediction the data of regrouped climatic zones of Europe [31] were applied. (The authors created these zones for continental scale soil quality assessment using the original 35 climatic areas of [27]). To evaluate the effects of slope conditions on the infiltration the *modified slope categories of the STUs (SLOPEM)* were first estimated. The weighted average was calculated from the SLOPE-DOM (Dominant slope class of the STU) and SLOPE-SEC (Secondary Slope class of the STU), considering twofold the dominant slope parameter.

Thereafter the *modified water balance code (WBM)* was estimated according to the WB, SLOPEM and the code for the grouped CORINE 2006 categories (CORGRP_CAT). Less water infiltration was assumed, if the slope is steeper and/or the soil is uncovered part of the year (arable and horticultural land use) [4; 32].

In the next step the **estimated leaching factor (LEACH_f)** was calculated from WBM. (The smaller the WBM code is, the higher the LEACH_f value, and vice versa.) To ascertain the filtration capacity of the soils the *water infiltration rate (IR)* is a decisive soil parameter [4; 6; 32]. To determine the soil IR code, first some auxiliary factors were estimated. *Topsoil (TPD_f) and subsoil (SPD_f) packing density factors* are dependent on PD_TOP and PD_SUB. The values of these factors are higher, if the packing density is low, and vice versa. Because of the infiltration is less affected by the subsoil compaction, the rate of change is less for the SPD_f.

The *impermeable layer factor (IL_f)* is dependent on the code for the presence of an impermeable layer within the soil profile of the STU (IL). The value of IL_f is higher (1) if no impermeable layer exists and the smaller, the closer is the impermeable layer to the ground surface.

Using the above parameters and the SPCC the IR was estimated. The infiltration rate is usually higher in sandy soils and lower in clayey soils, but this may change due to compaction or impermeable layer proximity. 10 grade **infiltration rate factor (IR_f)** was estimated according to the IR code. The **groundwater depth factor (DG_f)** was calculated from the estimated depth to the groundwater (DG) to take into account the depth to the groundwater during the filtration capacity estimation.

2.5 Estimating the cation exchange capacity and specific surface area of soils

The storing and filtration capacity of soil depends on the clay mineral and the organic matter surfaces and also on the charge of surfaces [4; 7; 33; 34]. The fixation and adsorption processes are dependent on the pH value or base saturation and on the aeration (redox potential) of the soil layers. The total amount of cations adsorbed by the negative charges of clay minerals and humus materials on a unit mass of soil is defined as the cation exchange capacity (CEC) of the soil. According to the literature [35–38], the average CEC values of soil can be estimated from the clay content, clay mineralogy, organic matter content and organic matter (humus) quality [7].

The *cation exchange capacity related to the humus content (CEC_HUM)* was estimated from the SPHC and HQ. The average CEC of fulvic acid, humic acid and peat materials were 1000, 500 and 150 cmol(+)/kg, respectively. The *cation exchange capacity related to the soil minerals (CEC_MIN)* was estimated from the SPCC and the topsoil and subsoil mineralogy (MIN_TOP and MIN_SUB). The average CEC of KX (1/1 Min. + Oxy. & Hydroxy.), KQ (Minerals + Quartz), MK (2/1 & 1/1 Minerals), M (2/1 & 2/1/1 non swel. Min.), NA (Not applicable), TO (Andic Minerals), TV (Vitric Minerals) and MS (Swel. & non swel. 2/1 Min.) were 7, 9, 17, 25, 25, 25, 25 and 83 cmol(+)/kg, respectively.

The **cation exchange capacity related to soil minerals and humus content (CEC_SUM)** is the simple average of the CEC_HUM and CEC_MIN codes. The calculated

values were converted to a 10-point scale, where the code 1 represents the lowest and the code 10 the highest CEC values.

One of the key soil properties affecting the ability of the storing and filtering is the specific surface area (SSA). SSA is highly correlated with the soil's physical and chemical properties, such as different adsorption processes. The average SSA values of the soil can be estimated from the clay content, clay mineralogy, organic matter content and organic matter (humus) quality, based on the relevant literature [7; 30; 39].

The *specific surface area related to the humus content (SSA_HUM)* was estimated from the SPHC and HQ. The average SSA of fulvic acid, humic acid and peat materials were 5000, 2000 and 1000 m²/kg, respectively. The *specific surface area related to the soil minerals (SSA_MIN)* was estimated from the SPCC) and the MIN_TOP and MIN_SUB. The average SSA of KX (1/1 Min. + Oxy. & Hydroxy.), KQ (Minerals + Quartz), MK (2/1 & 1/1 Minerals), M (2/1 & 2/1/1 non swel. Min.), NA (Not applicable), TO (Andic Minerals), TV (Vitric Minerals) and MS (Swel. & non swel. 2/1 Min.) were 32, 15, 74, 83, 83, 700, 700 and 416 m²/kg, respectively. The ***specific surface area related to soil minerals and humus content (SSA_SUM)*** is the simple average of the SSA_HUM and SSA_MIN codes. The calculated values were converted to a 10-point scale, where the code 1 represents the lowest and the code 10 the highest SSA values.

2.6 Calculating the sorption factors

The *base saturation of the soil profile (BASE_SAT)* was estimated from the base saturation ESDB code of the topsoil and subsoil (BS_TOP and BS_SUB). Because there is no other information about the acidity of the soils, the soil pH was evaluated according to this parameter. Base saturation generally increases with an increase in soil pH (low BASE_SAT code means soils with low soil pH, and vice versa) [40–42].

The *aeration code (AER)* was used to describe the aerobic/anaerobic conditions (redox status) of the soils. It refers to the presence or absence of electrons in soil. The fixation of elements in soil is dependent upon the redox and pH status of the soil system. E.g. acidic (low base saturation code) and relative anaerobic (low AER code) soil condition means that most elements in cationic form do not exhibit a propensity to exist in immobile hydroxide, oxide, oxyhydroxide forms, they prefer to exist in mobile, dissolved forms. Therefore using the BASE_SAT and AER codes we are able to qualitatively estimate the fixation potential of the elements or chemicals with different charges.

The *cation sorption factor (CATS_f)* was estimated from the **BASE_SAT** and **AER** codes. The total concentration of substances (elements or chemicals) in soils consists of three parts:

- (1) concentration of fixed substances comprising part of the structure of solid phase
- (2) concentration of substances adsorbed onto the surface of soil minerals and onto organic matter and
- (3) concentration of substances in soil water or groundwater [8].

The separation of the fixation and adsorption in soil-water system is not an easy task if we have few information about the soil condition, therefore we used the "sorption" term, which includes both soil processes [7]. (The concentration of substances solved in water phase and retained in soil capillary pores only indirectly, as the clay content dependent property was considered.) The cation sorption factor (**CATS_f**) simplifies the complex soil mechanisms. It was assumed that the sorption of elements in cationic form (and organic chemicals with positive charges) are higher at high base saturation (**BASE_SAT**) and at aerobic condition (**AER**) [43]. The calculated values were converted to a 10-point scale, where the factor 0.1 the worst and the code 1.0 the optimum cation adsorption conditions represents.

The *anion sorption factor (ANS_f)* was estimated also from the **BASE_SAT** and **AER** codes. There are different types of surfaces responsible for accumulating anions in soil. The most important are the oxides, the edges of alumino-silicate clay minerals (especially the 1:1 clay minerals) and the soil organic materials. The net positive charge of these

materials is highly dependent on the soil pH. If the pH is low, then will develop a positive surface charge. However, the fixation processes of substances in anionic form depends on the aerobic/anaerobic condition of the soil system, too [7; 44]. Therefore the anion sorption factor (**ANS_f**) simplifies the complex soil mechanisms as follows: the sorption of elements in anionic form (and organic chemicals with negative charges) are higher at low base saturation (**BASE_SAT**) and at aerobic condition (**AER**). The calculated values were converted to a 10-point scale, where the factor 0.1 the worst and the factor 1.0 the optimum anion adsorption conditions represents.

The *mineral factor* (**MIN_f**) was calculated from the topsoil and subsoil mineralogy (**MIN_TOP** and **MIN_SUB**) to take into account the oxide surfaces during the anion sorption estimation.

2.7 Storing and filtration capacity codes

The storing and filtering capacity codes were estimated based on the above derived parameters. The calculated values were converted to a 10-point scale, where the code 1 the lowest and the code 10 the highest capacities represents. To transform the estimated values (continuous scale variables) into limited number (10) of distinct categories the Visual Binning method was used (SPSS, Transform, Visual Binning, Equal Percentiles Based on Scanned Cases). This method generates binned categories with an equal number of cases in each bin using the empirical (empirical distribution function with averaging) algorithm for percentiles.

2.7.1 The cation storing capacity

The *cation storing capacity* (**STOR_CAPCA**) was estimated from the soil mineral cation exchange capacity (**CEC_MIN**), cation exchange capacity related to humus content (**CEC_HUM**), cation sorption factor (**CATS_f**) and leaching factor (**LEACH_f**). It represents the soil storing capacity for elements in cationic form (e.g. K^+ , Na^+ , Ca^{2+} , Fe^{2+} , Fe^{3+} or heavy metals, as Ba^{2+} , Cd^{2+}) and organic chemicals with positive charges (e.g. organoamines, metallo-organics). It was assumed that **CEC_HUM** is pH dependent, but only part of the **CEC_MIN** depends on soil pH (base saturation). The adsorption was calculated accordingly. The fixation of these materials however, is dependent on the soil pH (base saturation) and aerobity (e.g. the Fe^{2+} or FeS are mobile forms and Fe^{3+} or $Fe(OH)_3$, $Fe(CO_3)$ are immobile forms of iron) (**CAT_f**). The higher the leaching (**LEACH_f**), the less the storing capacity of the soils is.

2.7.2 The cation filtering capacity

The *cation filtering capacity* (**FILT_CAPCA**) was estimated from the soil mineral cation exchange capacity (**CEC_MIN**), cation exchange capacity related to humus content (**CEC_HUM**), cation sorption factor (**CATS_f**), infiltration rate factor (**IR_f**) and groundwater depth factor (**DG_f**). It represents the soil filtration capacity for elements in cationic form and organic chemicals with positive charges. It was assumed that the higher the cation sorption capacity of the soil (dependent on **CEC_MIN**, **CEC_HUM** and **CAT_f**), the higher the soil cation filtration capacity is. But this parameter also depends on the infiltration rate (**IR_f**): soils with high water infiltration rate, are assigned a low filtering capacity (no time enough for the physicochemical processes), and less permeable soils, with low infiltration rate, are assigned a high capacity. The filtration capacity of a soil indicates however, its capacity to bind substances in the soil and not let them reach the groundwater. The thickness of the filtration path up to the groundwater was considered also under this method (**DG_f**).

2.7.3 The anion storing capacity

The *anion storing capacity* (**STOR_CAPAN**) was estimated from the specific surface area related to soil minerals (**SSA_MIN**), mineral factor (**MIN_f**), specific surface area related to humus content (**SSA_HUM**), anion sorption factor (**ANS_f**) and leaching factor (**LEACH_f**). It represents the soil storing capacity for elements in anionic form (e.g. nutrients, as phosphate, nitrate; heavy metals, as molybdate, arsenate or other elements, as borate, sulfate, halides) and organic chemicals with negative charges (e.g. organic chemicals possess hydroxyl or carboxyl functional groups). It was assumed that the higher the specific surface of soil, the higher the possibility of the appearance of positive charges. The formation of positive charges at humic materials (**SSA_HUM**) is pH dependent, but only part of the mineral surfaces (**SSA_MIN**) have pH (base saturation) dependent positive charges. It was taken into account that sesquioxides have particularly large amount of positive charges (**MIN_f**). The anion adsorption was calculated accordingly. The fixation of these materials however, is dependent on the soil pH (base saturation) and aerobity (e.g. the mobility of Cr^{3+} , HCrO_4^- , CrO_4^{2-} , Cr_2O_3 and $\text{Cr}(\text{OH})_3$ forms of chrome are different) (**ANS_f**). The higher the leaching (**LEACH_f**), the less the anion storing capacity of the soils is [45].

2.7.4 The anion filtering capacity

The *anion filtering capacity* (**FILT_CAPAN**) was estimated from the specific surface area related to soil minerals (**SSA_MIN**), mineral factor (**MIN_f**), specific surface area related to humus content (**SSA_HUM**), anion sorption factor (**ANS_f**), infiltration rate factor (**IR_f**) and groundwater depth factor (**DG_f**). It represents the soil filtration capacity for elements in anionic form and organic chemicals with negative charges. It was assumed that the higher the anion sorption capacity of the soil (dependent on **SSA_MIN**, **SSA_HUM** and **ANS_f**), the higher the soil anion filtration capacity is. The filtering capacity dependency on the infiltration rate (**IR_f**) and groundwater depth (**DG_f**) was taken into account similarly as in the case of cation filtering capacity.

2.7.5 The solids and pathogenic microorganisms storing capacity

The *solids and pathogenic microorganisms storing capacity* (**STOR_CAPSO**) was estimated from the specific surface area related to soil minerals and humus content (**SSA_SUM**) and leaching factor (**LEACH_f**). It was assumed that the sorption of different solid materials and microorganisms is dependent mainly on the soil surfaces (**SSA_SUM**). The higher the leaching (**LEACH_f**), the less the solids and pathogenic microorganisms storing capacity of the soils is.

2.7.6 The solids and pathogenic microorganisms filtering capacity

The *solids and pathogenic microorganisms filtering capacity* (**FILT_CAPSO**) was estimated from the specific surface area related to soil minerals and humus content (**SSA_SUM**), infiltration rate factor (**IR_f**) and groundwater depth factor (**DG_f**). It was assumed that the higher the solids and pathogenic microorganisms sorption in the soil (dependent on **SSA_SUM**), the higher the soil solids and pathogenic microorganisms filtration capacity is [10; 46–48]. The filtering capacity dependency on the groundwater depth (**DG_f**) was taken into account similarly as in the case of cation filtering capacity. But the estimated effect of the infiltration rate is opposite. In this case the suspended dirt and pollutant particles are mechanically fixed to the soil. (The term Mechanical Filtering Capacity means capacity of the soil to mechanically clarify a suspension [49]).

The filtering capacity of a soil was characterized by the amount of water being able to pass the respective soil in a given time unit [50]. The higher the filtering capacity is, the more water can pass. So if the residence time of the water is shorter, the higher the filtering capacity is. It means that soils with a high infiltration rate (**IR_f**) have a high solids and pathogenic microorganisms filtering capacity [10; 51].

2.7.7 The non-polar organic chemicals storing capacity

The *non-polar organic chemicals storing capacity* (**STOR_CAPNP**) was estimated from the specific surface area related to soil minerals and humus content (**SSA_SUM**) and leaching factor (**LEACH_f**). It was assumed that the sorption of different non-polar organic chemicals is dependent mainly on hydrophobic surfaces. The bigger soil surfaces (**SSA_SUM**), the higher the hydrophobic part of these surfaces is [8]. The higher the leaching (**LEACH_f**), the less the non-polar organic chemicals storing capacity of the soils is.

2.7.8 The non-polar organic chemicals filtering capacity

The *non-polar organic chemicals filtering capacity* (**FILT_CAPNP**) was estimated from the specific surface area related to soil minerals and humus content (**SSA_SUM**), infiltration rate factor (**IR_f**) and groundwater depth factor (**DG_f**). It was assumed that the higher the non-polar organic chemicals sorption in the soil (dependent on **SSA_SUM**), the higher the non-polar organic chemicals filtration capacity is. The filtering capacity dependency on the infiltration rate (**IR_f**) and groundwater depth (**DG_f**) was taken into account similarly as in the case of cation filtering capacity [52; 53].

2.7.9 The nonaqueous phase liquids (NAPLs) storing capacity

The *nonaqueous phase liquids (NAPLs) storing capacity* (**STOR_NAPL**) was estimated from the clay content of the soil profile (**SPCC**), specific surface area related to soil minerals and humus content (**SSA_SUM**) and leaching factor (**LEACH_f**). According to the literature (e.g. [18; 54–58]) the sorbed and retained nonaqueous phase liquids amount depends on the pore size distribution (it is dependent on **SPCC**) and on the area of hydrophobic surfaces (it is dependent on the **SSA_SUM**) of the soils. The higher the leaching (**LEACH_f**), the less the nonaqueous phase liquids storing capacity of the soils is.

2.7.10 The nonaqueous phase liquids (NAPLs) filtering capacity

The *nonaqueous phase liquids (NAPLs) filtering capacity* (**FILT_NAPL**) was estimated from the specific surface area related to soil minerals and humus content (**SSA_SUM**), infiltration rate factor (**IR_f**) and groundwater depth factor (**DG_f**). It was quite difficult to define the NAPLs filtering capacity. This parameter was measured as the capacity of soils to clarify the infiltrating water contains dissolved and emulsified NAPL components. It was assumed that the higher the nonaqueous phase liquids component (mainly non-polar organics) sorption in the soil (dependent on **SSA_SUM**), the higher the NAPLs filtration capacity is. The filtering capacity dependency on the infiltration rate (**IR_f**) and groundwater depth (**DG_f**) was taken into account similarly as in the case of cation filtering capacity [59; 60].

The flow chart of the storing and filtering capacity estimation procedure is presented by the *Figure 2*.

The GIS tasks were performed with the software ESRI ArcGIS 9.3. The capacity codes of all the ESDB polygons were calculated using the key variables soil mapping units (SMUs) and soil taxonomy units (STUs). Then we mapped the storing and filtration capacity of substances for the SMUs across the European Union after assigning colors to a continuous scale of grade 10 capacity code system, where 1 represents the lowest and the code 10 the highest capacities. Thereafter we calculated the percentage area (PA) of each categories across the European Union and within each Biogeographical Region (BGR). A weighted capacity index (WCI) was calculated by the following formula:

$$WCI = \sum_{i=1}^{10} PA_i \times C_i,$$

where C_i was the capacity code and PA_i the percentage area of the i th category within the given BGR. The WCI values were normalized to ten-stage scale (WCI_{norm}).

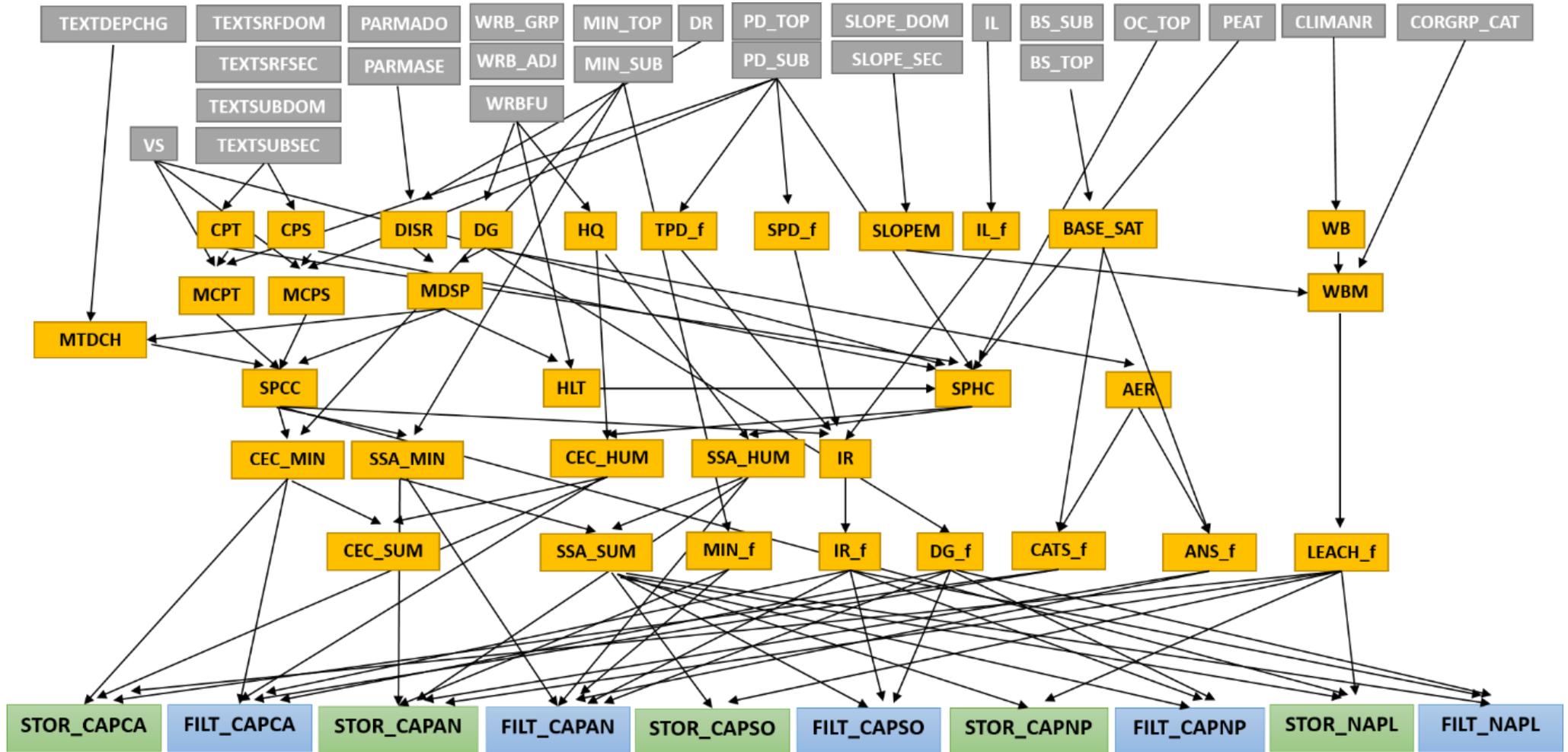


Figure 2. Diagram for the evaluation of the soil storage and filtering functions

3. Presenting the soil storing and filtering capacity maps

Results of soil storing and filtration capacity evaluation for fields of the European Union, prepared according to the above algorithm are presented in Fig. 3–12.

The spatial pattern of areas with high soil storing capacity (Fig. 3–7) are largely different depending on the stored substances. In general, those soils are characterized by good storing capability that have thick topsoil and subsoil layers and these layers are free from the effects of groundwater. As the content of clay and humus content increases and the stone or gravel content decreases, the storage capacity increases simultaneously. However, the effect of soil pH and the soil mineralogy is different depending on the various groups of pollutants (e.g. the calcareous soils with high swelling clay mineral content are able to bind more cations, whereas the acidic soils or the soils with high sesquioxide content have higher anion storing capacity).

The soil filtering capacity maps (Fig. 8–12) of different groups of substances are in part similar to the storing capacity maps, since both are based on the estimated CEC or SSA of soils. The main difference between the two mapped parameters was to take account of infiltration rate (IR_f) and the thickness of the filtration path up to the groundwater (DG_f) during the filtering capacity estimation. Moreover assessing the impact of infiltration rate depending on the quality of the infiltrating material were different (generally the filtering capacity increased in less permeable soils, but in the case of the solids and pathogenic microorganisms the estimated effect of the infiltration rate was opposite). These factors influenced primarily the spatial pattern of filtering capacity maps of substances in anionic forms, the NAPLs and most significantly the solids and pathogenic microorganisms.

Cation storing capacity
(elements in cationic form and organic chemicals with positive charges)

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

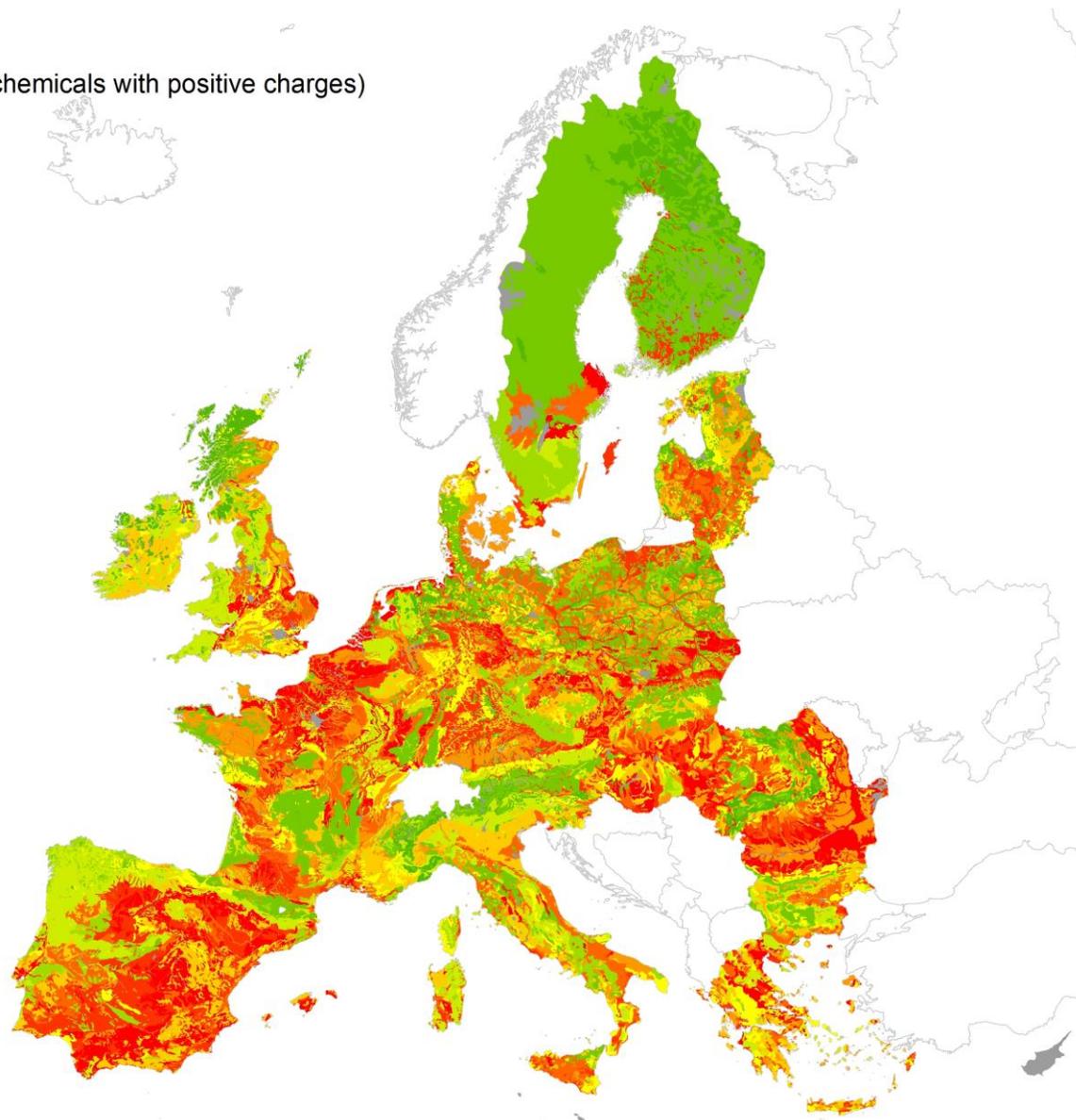
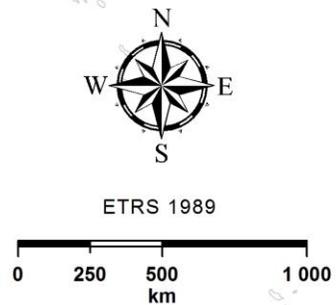


Figure 3. Map of cation storing capacity

Anion storing capacity
(elements in cationic form and organic chemicals with positive charges)

Legend

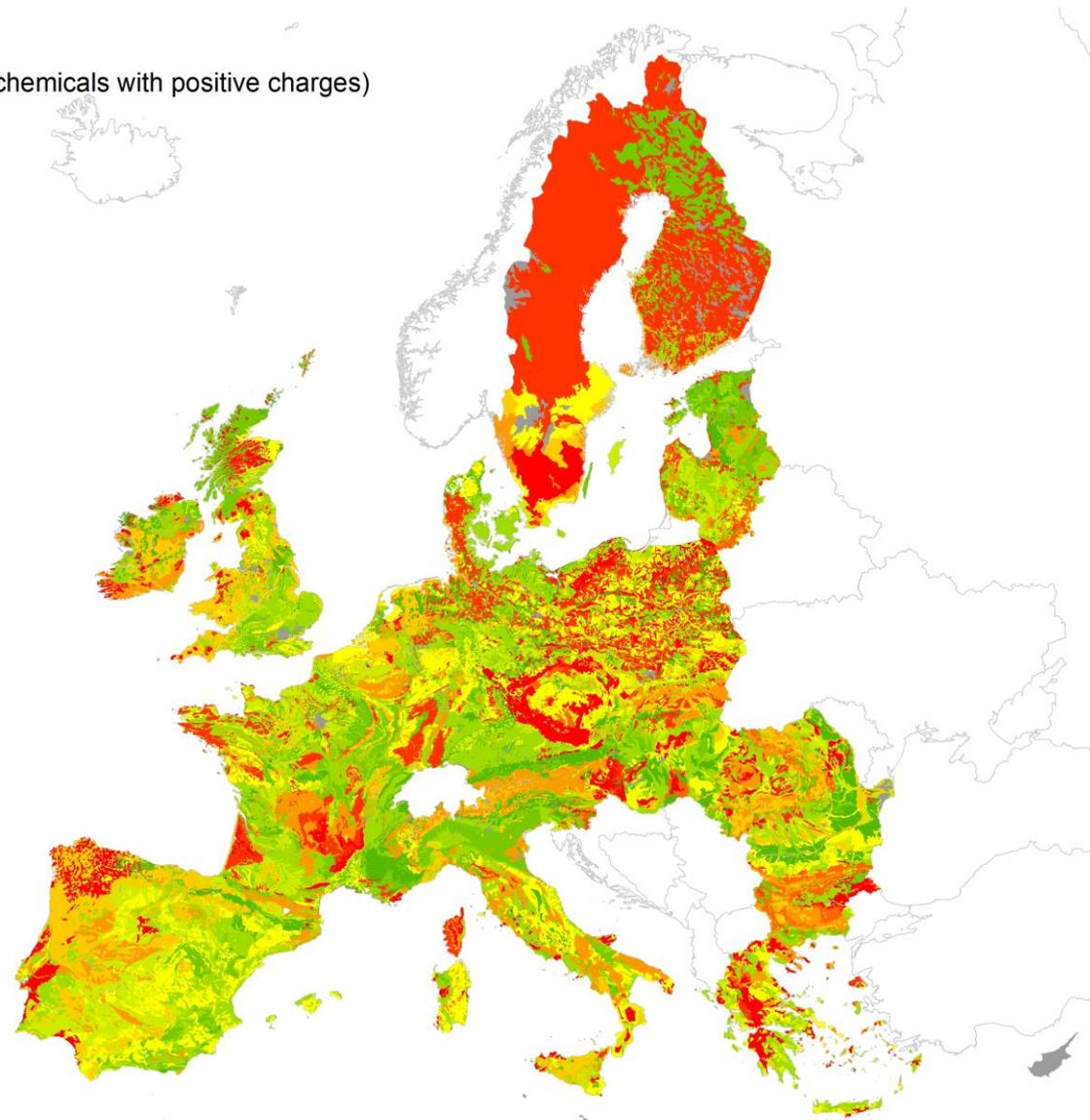
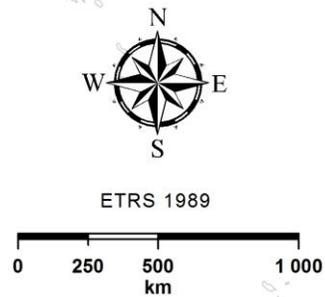


Figure 4. Map of anion storing capacity

Solids and pathogenic microorganisms storing capacity

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

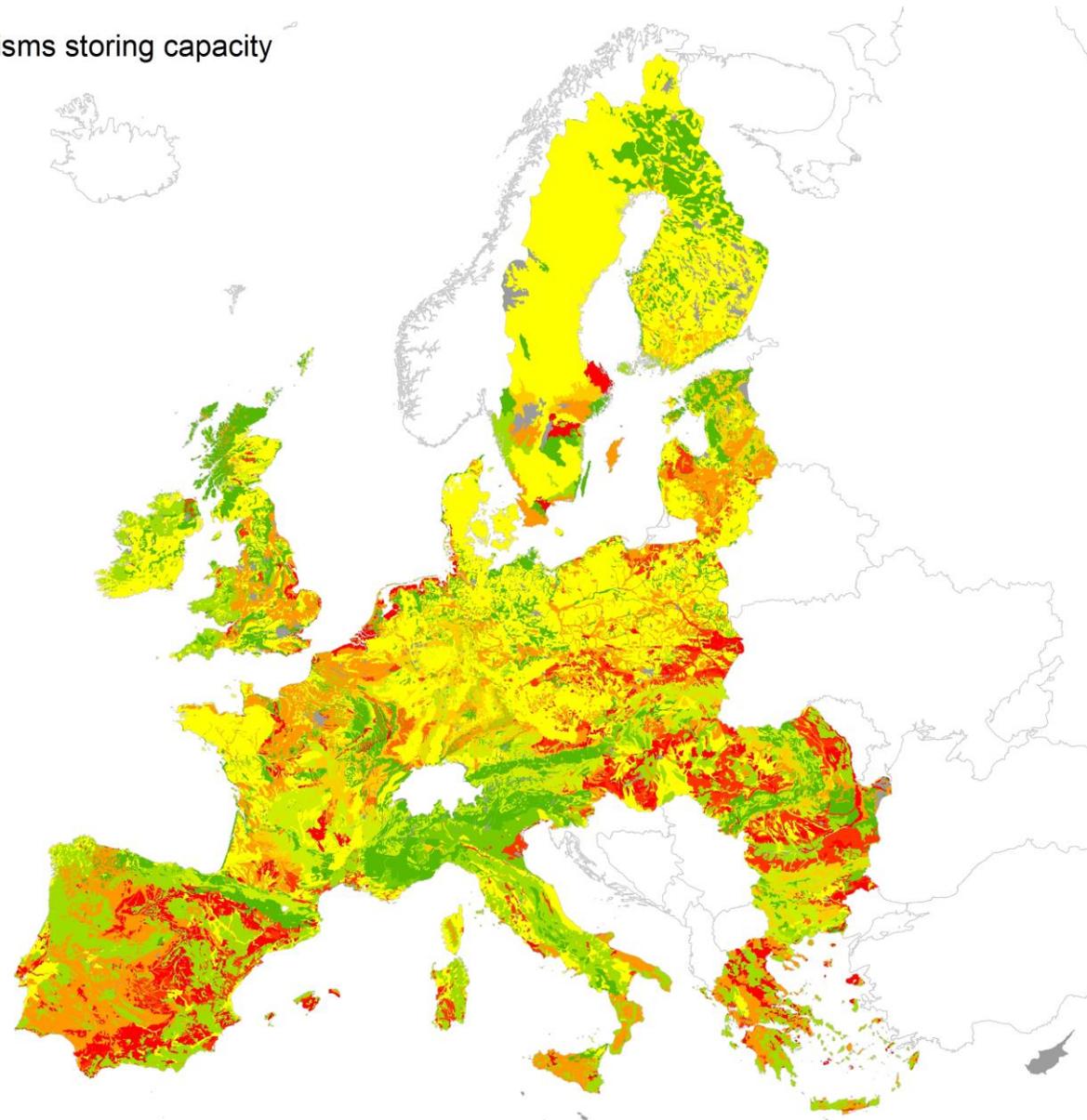
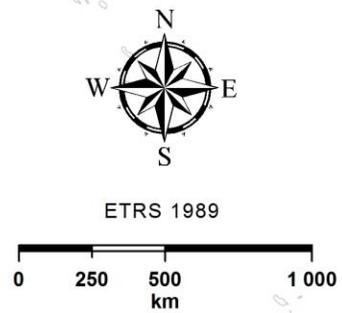


Figure 5. Map of solids and pathogenic microorganisms storing capacity

Non-polar organic chemicals storing capacity

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

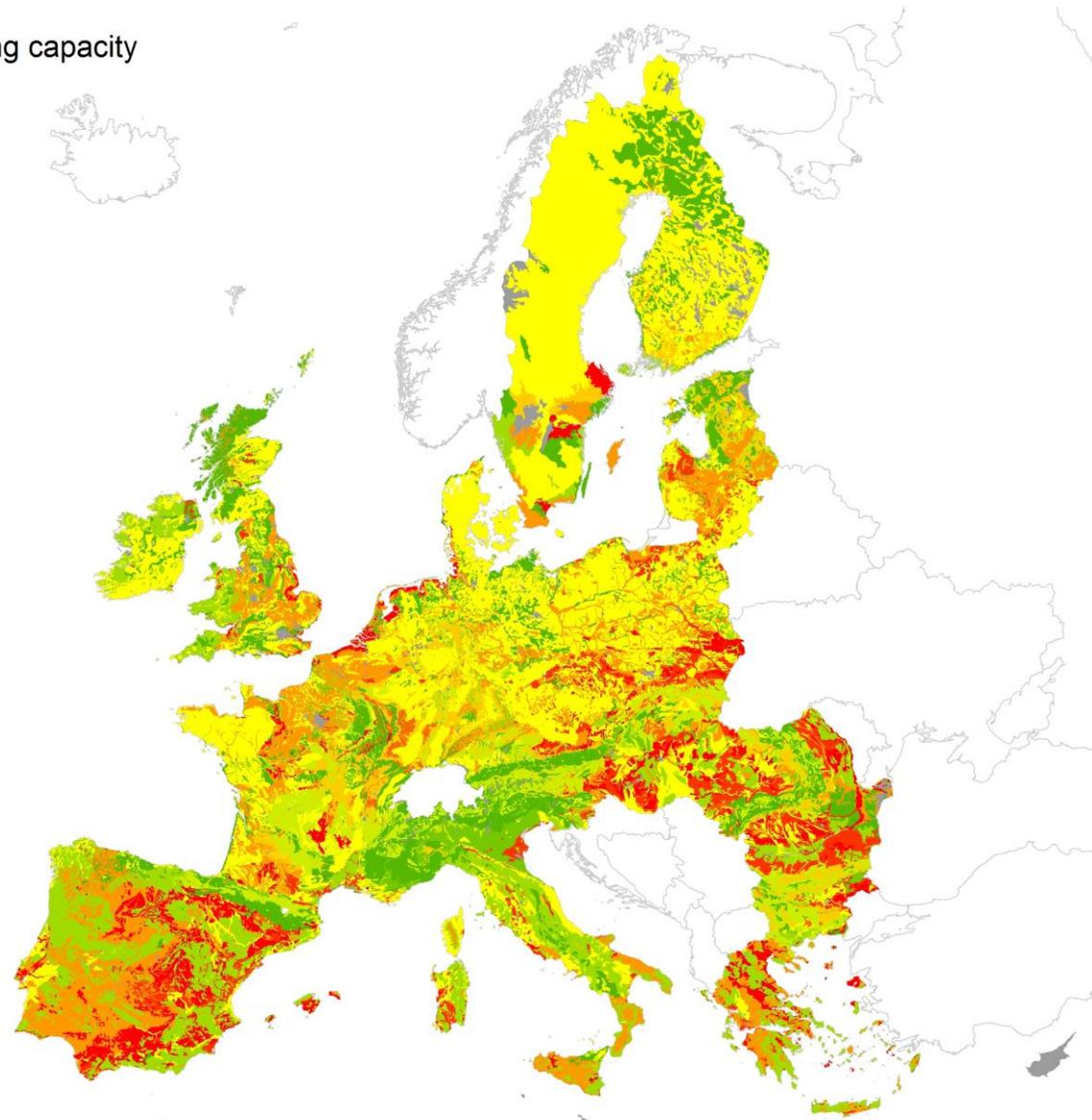
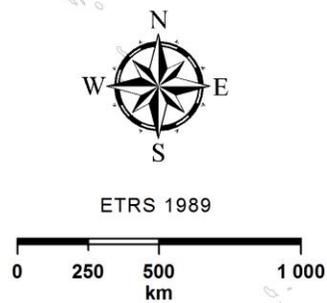


Figure 6. Map of non-polar organic chemicals storing capacity

Nonaqueous Phase Liquids (NAPLs) storing capacity

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

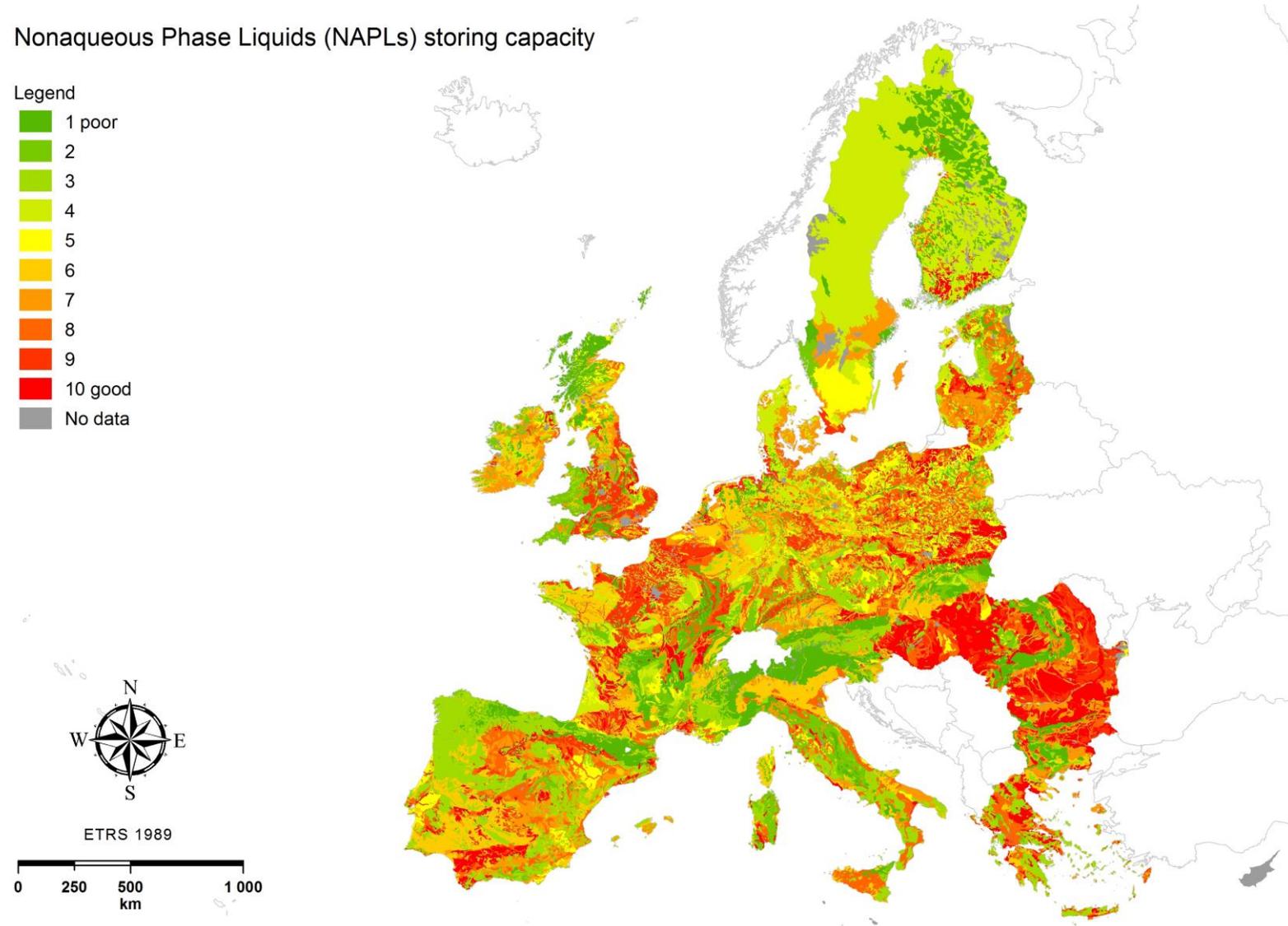


Figure 7. Map of nonaqueous phase liquid (NAPL) storing capacity

Cation filtering capacity
(elements in cationic form and organic chemicals with positive charges)

Legend

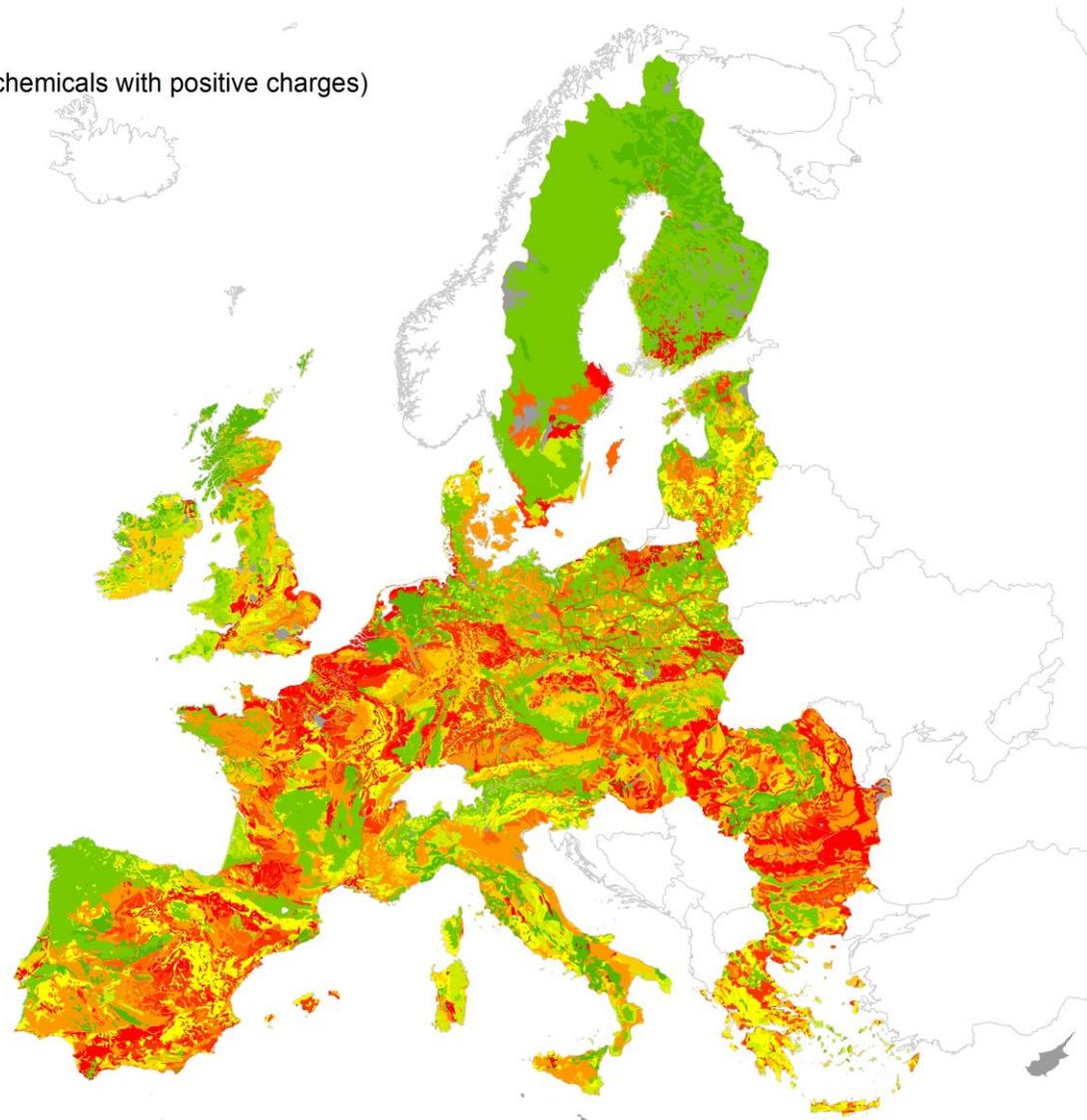
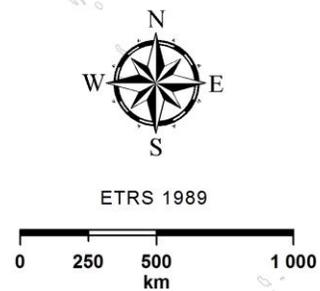


Figure 8. Map of cation filtering capacity

Anion filtering capacity
(elements in cationic form and organic chemicals with positive charges)

Legend

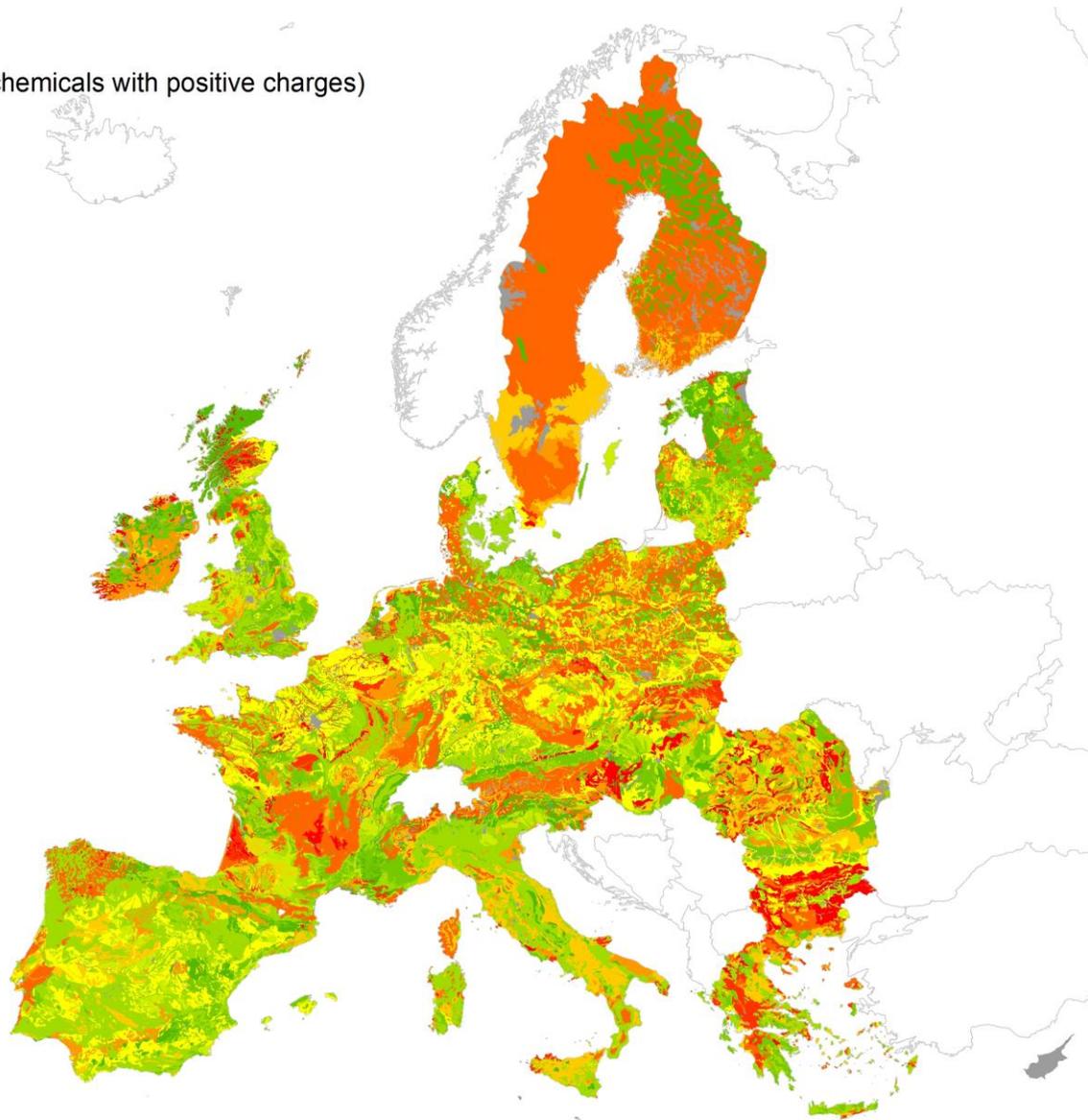
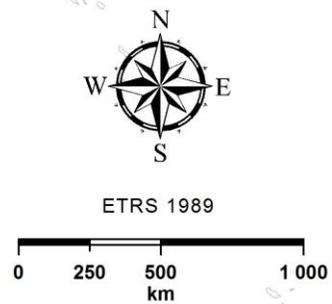


Figure 9. Map of anion filtering capacity

Solids and pathogenic microorganisms filtering capacity

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

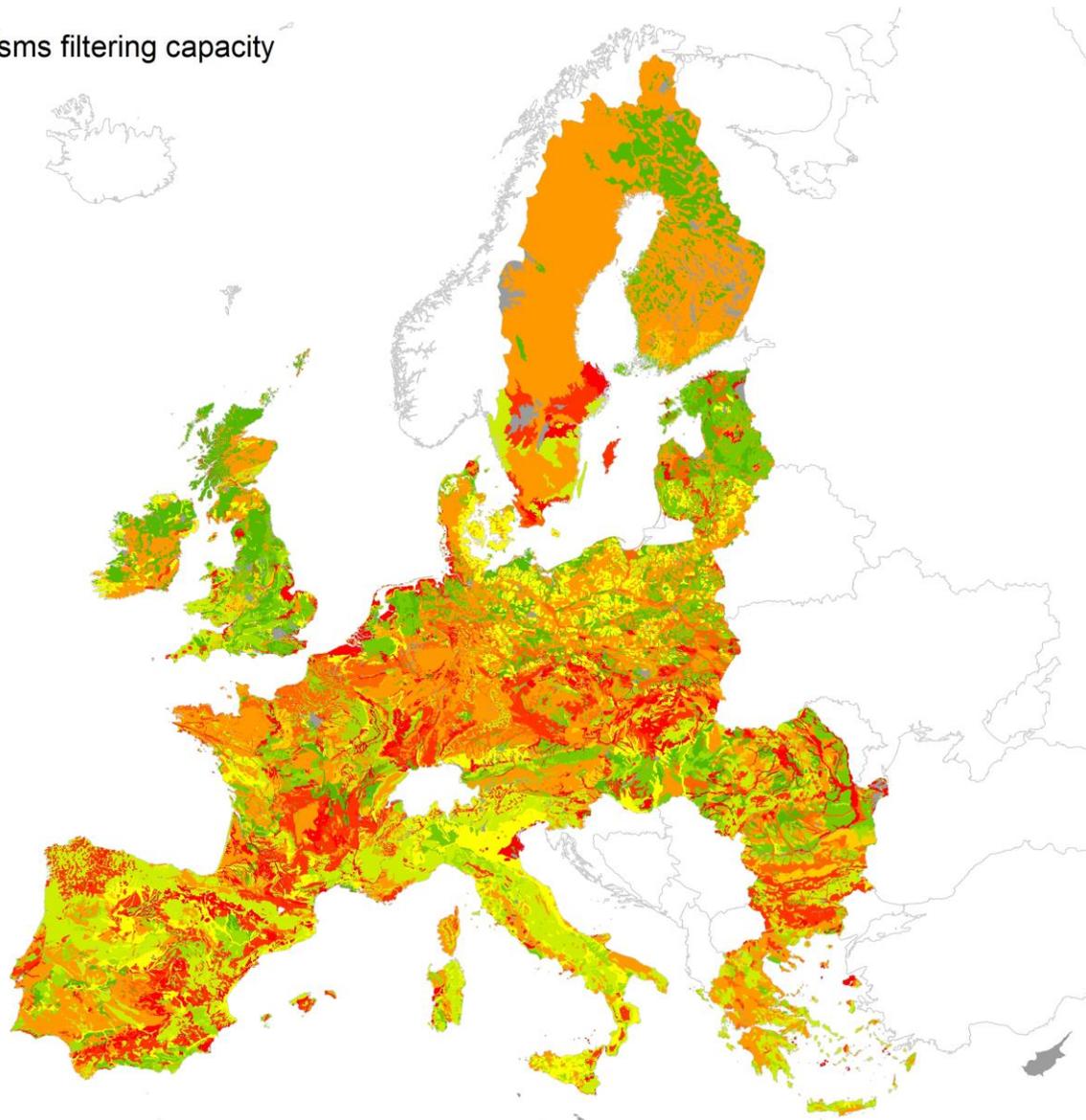
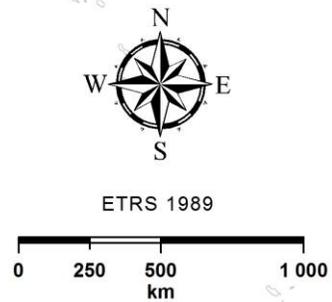


Figure 10. Map of solids and pathogenic microorganisms filtering capacity

Non-polar organic chemicals filtering capacity

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

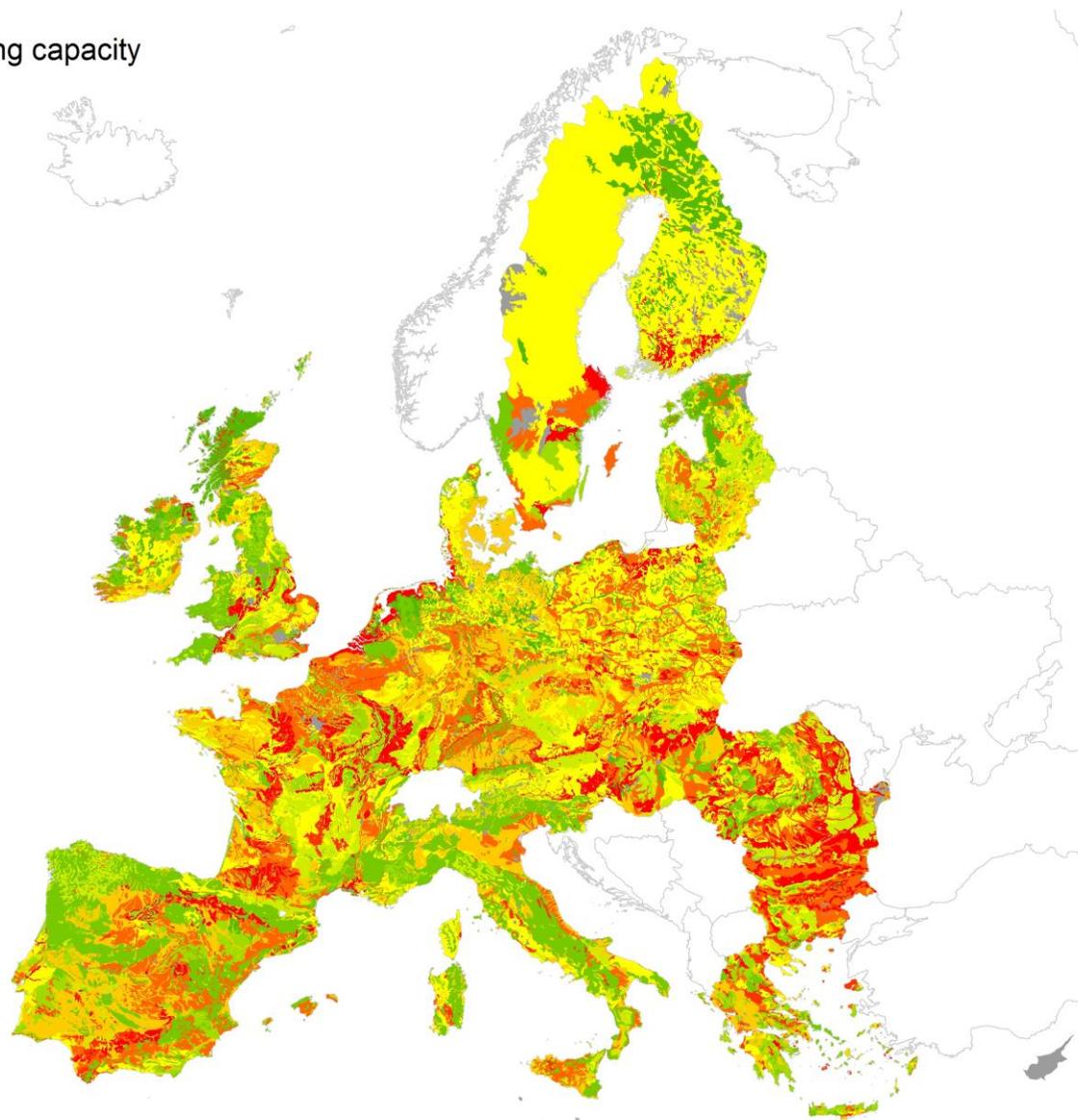
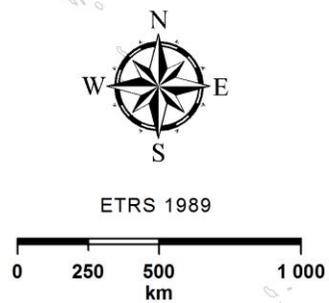


Figure 11. Map of non-polar organic chemicals filtering capacity

Nonaqueous Phase Liquids (NAPLs) filtering capacity

Legend

- 1 poor
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10 good
- No data

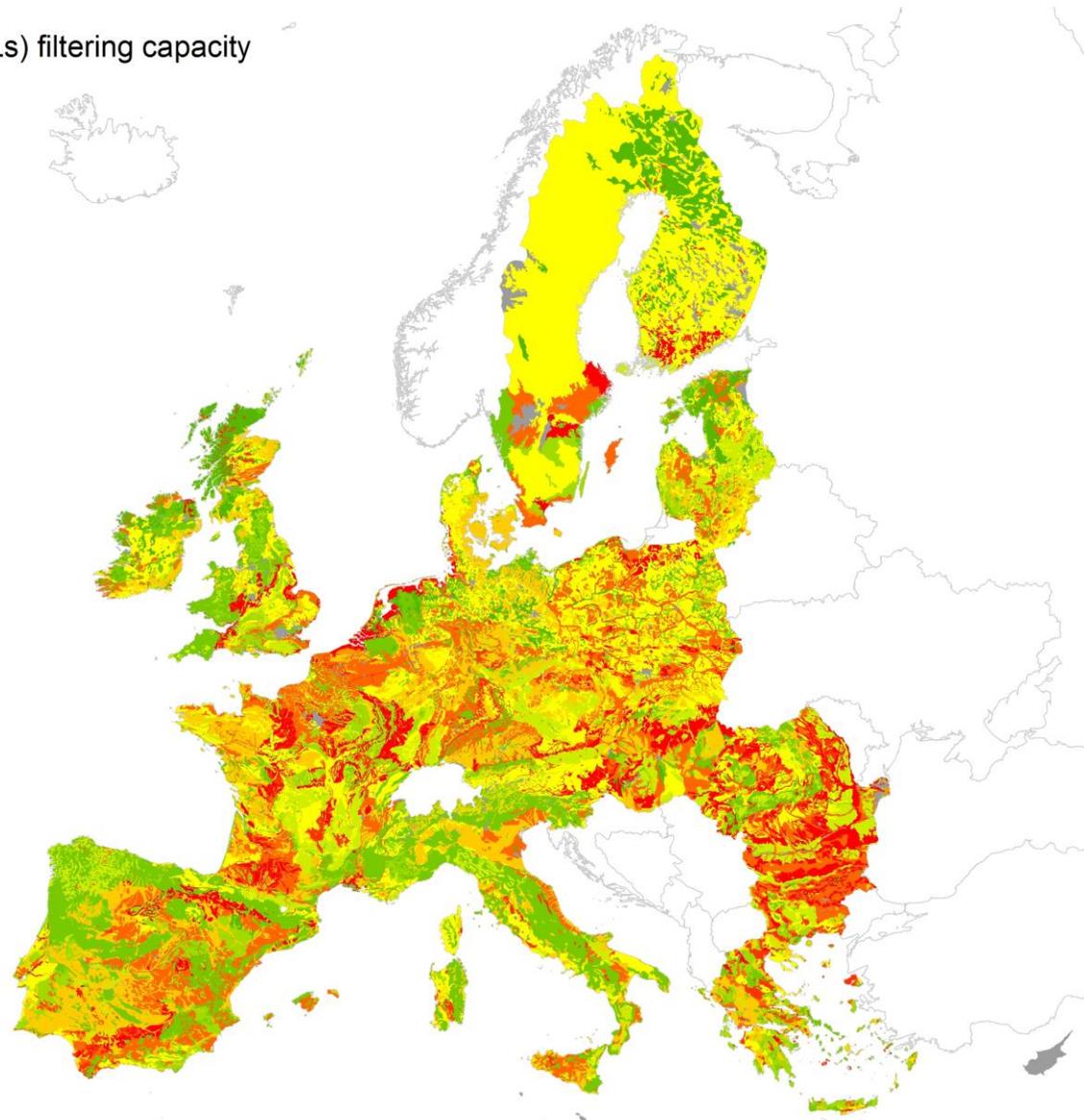
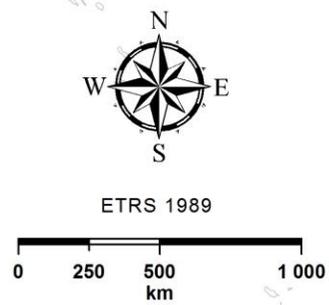


Figure 12. Map of nonaqueous phase liquid (NAPL) filtering capacity

As a next step we examined the distribution of storing and filtering properties of soils by European BGR (Fig. 13). Using the calculated normalized ten-stage capacity indices (WCI_{norm}) we characterized the storing and filtering ability of soils at a particular group of substances in a given region. Comparing the storing and filtering properties in Fig. 14 and 15 is visible that the Steppic, Pannonian and Black sea regions have the highest storing and filtering capacity in the case of substances in cationic forms and NAPLs, while the Boreal and Alpine regions can be characterized by the best ability to store and filter the anionic substances. The storing and filtering capability of regions are quite different in the case of non-polar organics, and similar differences were observed in the case of solids and pathogenic microorganisms.

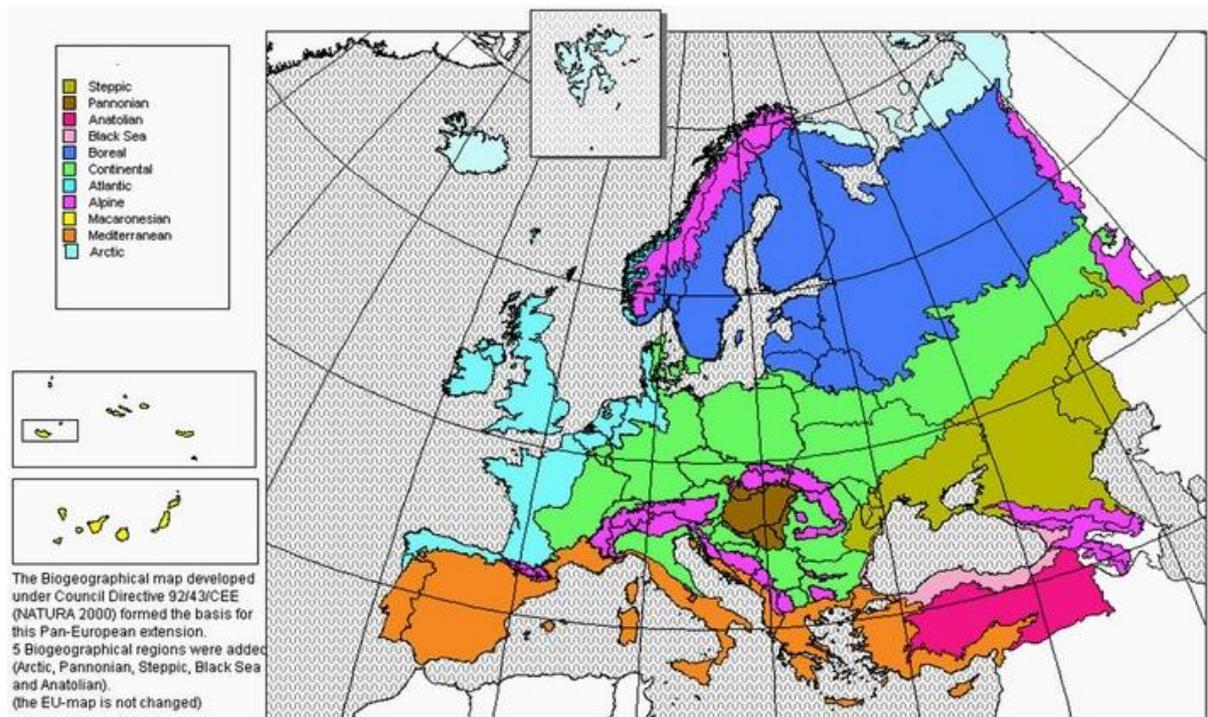


Figure 13. The Biogeographical Regions map of Europe [61]

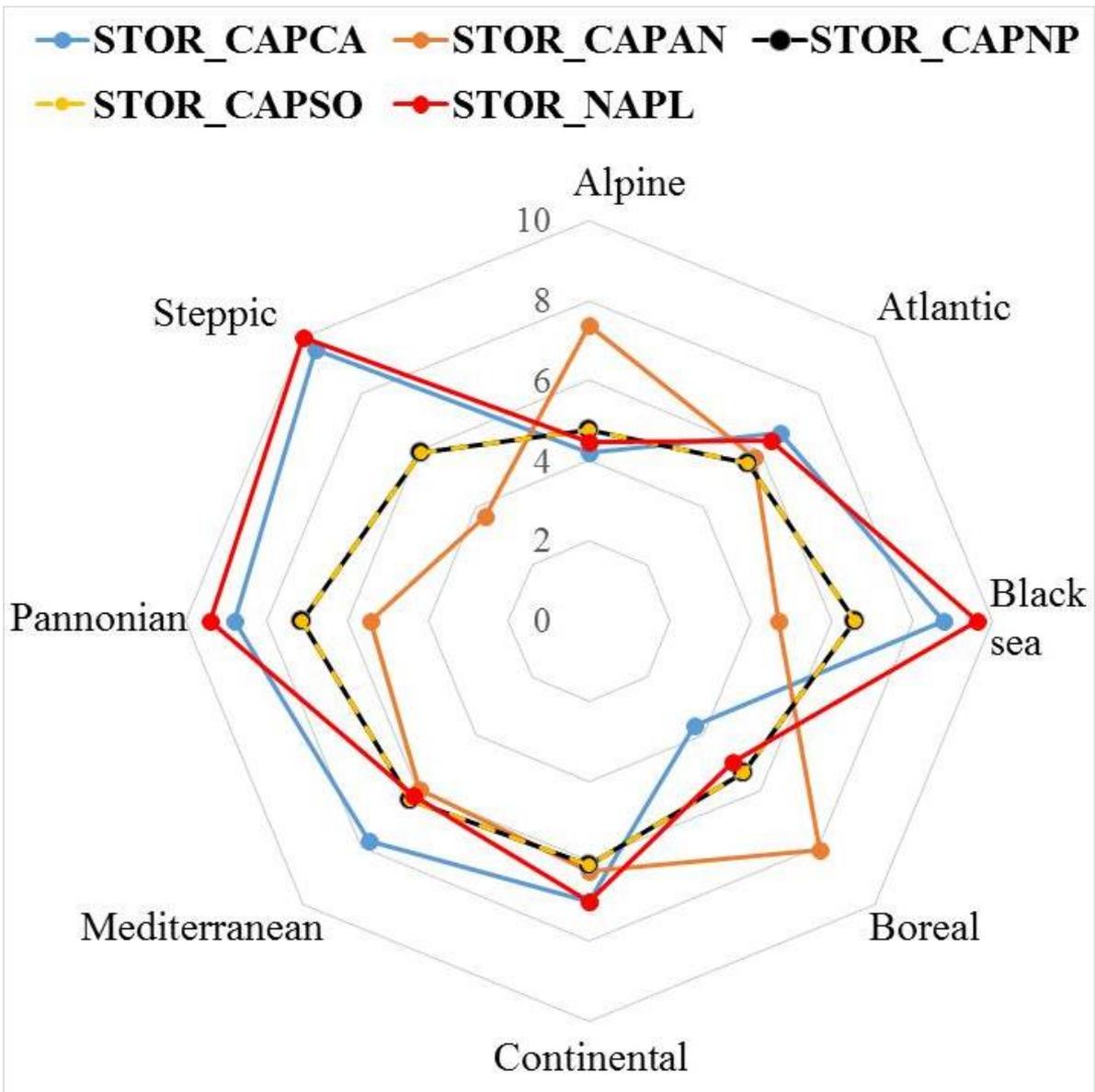


Figure 14. Comparison of WCI_{norm} values of European Biogeographical Regions to evaluate the spatial pattern of storing capacity of different substances (1: poor; 10: good)

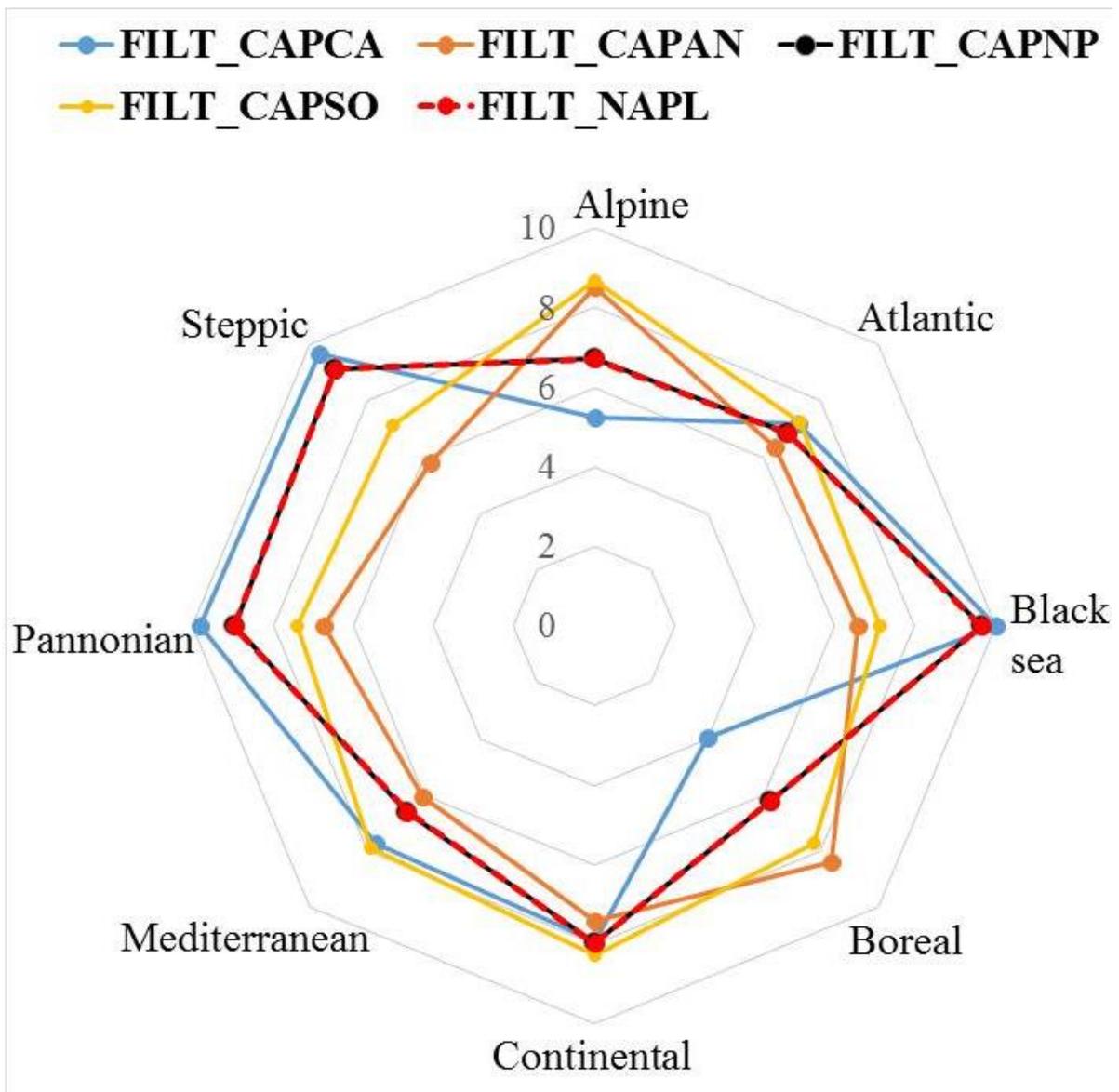


Figure 15 Comparison of WCI_{norm} values of European Biogeographical Regions to evaluate the spatial pattern of filtering capacity of different substances (1: poor; 10: good)

4. Conclusions

The presented algorithm to estimate the soil storing and filtering capacity based on large-scale soil map data is an important contribution to obtain basic Europe-wide environmental information. The knowledge of soil storing and filtering capacity for different substances is very useful during the environmental planning and modelling. These soil parameters (and the related sub-parameters) can usually not easily be determined or calculated by scientists without continent-wide extensive data access. With the publication of this study, the users will be able to download European datasets (storing and filtering capacity categories of soils in the case of different groups substances and furthermore the estimated CEC and SSA categories of soils at European scale) from the European Soil Data Centre.

The proposed algorithm provides a framework for the digital soil mapping of the soil storing and filtering functions at continental scale. The presented maps delineate areas where soil ability to store and filter a given type of substance is low and therefore the vulnerability of ecosystem in the case of soil pollution with this substance is high.

The study also suggests the possibility of improvement of the output datasets. As shown in the estimation algorithm above, a very complex estimation chain leads to the results, due to lack of basic spatial data needed. The development of European soil database, including the enlargement of soil monitoring systems will serve the growth of accuracy and reliability of soil function estimation methods.

References

- [1.] Blum, W.E.H., 2002. Environmental protection through sustainable soil management, a holistic approach. In: Pagliali, M. & Jones, R.: Sustainable land management – Environmental protection. A soil physical approach. Advances in geology. 35. IUSS. Catena Verlag GMBH. Reiskirchen. 1–8.
- [2.] White, R.E., 2005. Principles and practice of soil science: the soil as a natural resource. 4th ed. Wiley-Blackwell.
- [3.] Yong, R.N., Nakano, M. and Pusch, R., 2012. Environmental soil properties and behavior. Boca Raton, CRC Press.
- [4.] Hillel, D., 1998. Environmental soil physics. Academic Press. New York
- [5.] Lal, R. and Shukla, L., 2004. Principles of Soil Physics. Books in Soils, Plants, and the Environment Series. Marcel Dekker Inc.
- [6.] Glinski, J., Horabik, J. and Lipiec, J. (eds.), 2011. Encyclopedia of Agrophysics. Springer. Dordrecht.
- [7.] Dragun, J., 1998. The soil chemistry of hazardous materials. Farmington Hills, Michigan. Amherst. Massachusetts.
- [8.] Yaron, B., Dror, I. and Berkowitz, B., 2012. Soil-subsurface change. Chemical pollutant impacts. Springer. Heidelberg.
- [9.] Beck, T., Bos, U., Wittstock, B., Baitz, M., Fischer, M. and Sedlbauer, K., 2003. LANCA ® Land use indicator value calculation in life cycle assessment – Method Report. Fraunhofer Verlag. Echterdingen.
- [10.] Clark, S. E., 2000. Urban stormwater filtration: optimization of design parameters and a pilot-scale evaluation. Ph.D. Dissertation, University of Alabama at Birmingham, Birmingham, AL.
- [11.] Allen, S., Brown, P., McKay, G. and Flynn, O., 1992. An evaluation of single resistance transfer models in the sorption of metals ions by peat. Journal of Chemical Technology and Biotechnology. 54. 271–276.
- [12.] Pitt, R.E., Field, R., Lalor, M., Brown, M., 1995. Urban stormwater toxic pollutants: assessment, sources, and treatability. Water Environment Research. 67 (3): 260–275.
- [13.] Pitt, R., Clark, S. and Field. R., 1999a. Groundwater contamination potential from stormwater infiltration practices. Urban Water. 1 (3): 217–236.
- [14.] Pitt, R., Lantrip, J., Harrison, R., Henry, C. and Hue, D., 1999b. Infiltration through disturbed urban soils and compost-amended soil effects on runoff quality and quantity. U.S. Environmental Protection Agency, Water Supply and Water Resources Division, National Risk Management Research Laboratory. EPA 600/R-00/016. Cincinnati, Ohio.
- [15.] Zalasiewicz, J., Williams, M., Steffen, W. and Crutzen, P., 2010. The new world of the Anthropocene. Environmental Science & Technology. 44 (7): 2228–2231.
- [16.] Ewing, S.A. and Singer, M.J., 2012. Soil quality. In: Huang, P.M., Li, Y., Sumner, M.E., Handbook of soil sciences. Resource management and environmental impacts. CRC press. Boca Raton. 26-1 – 26-28.
- [17.] Steenhuis, T.S., Staubitz, W., Andreini, M.S., Surface, J., Richard, T. L., Paulsen, R., Pickering, N.B., Hagerman, J.R. and Geohring, L.D., 1990. Preferential movement of pesticides and tracers in agricultural soils. Journal of Irrigation and Drainage Engineering. 116 (1): 50–66.
- [18.] Rubin, H., Narkis, N. and Carberry, J., 1998. Soil and Aquifer Pollution. Springer-Verlag. Berlin, Heidelberg.
- [19.] Mayer, A.S. and Hassanizadeh, S.M., 2005. Soil and groundwater contamination: Nonaqueous phase liquids. Water resources monograph. 17. AGU. Washington.
- [20.] Tim, U.S. and Mostaghim, S., 1991. Model for predicting virus movement through soil. Ground Water. 29 (2): 251–259.
- [21.] Sherameti, I. and Varma, A. (eds.), 2010. Soil heavy metals. Soil Biology. 19. Springer. Heidelberg.
- [22.] Meuser, H., 2013. Soil remediation and rehabilitation. Treatment of contaminated and disturbed land. Environmental pollution. 23. Springer. Dordrecht.

- [23.] Teixeira, W.G., Ceddia, M.B., Ottoni, M.V. and Donnagema, G.K. (eds.), 2014. Application of soil physics in environmental analyses. Measuring, modelling and data integration. Progress in soil science. Springer. Cham.
- [24.] The European Soil Database distribution version 2.0. 2004. European Commission and the European Soil Bureau Network, CD-ROM, EUR 19945 EN
- [25.] Panagos, P., Van Liedekerke, M., Filippi, N., Montanarella, L., 2006. MEUSIS: towards a new multi-scale European soil information system. ECONGEO, 5th European Congress on Regional Geoscientific Cartography and Information Systems. Barcelona, Spain. 175–177.
- [26.] <http://land.copernicus.eu/pan-european/corine-land-cover/clc-2006/view>
- [27.] Hartwich, R., Baritz, R., Fuchs, M., Krug, D. and Thiele, S., 2005. Erläuterungen zur Bodenregionenkarte der Europäischen Union und ihrer Nachbarstaaten 1:5,000,000 (version 2.0). Bundesanstalt für Geowissenschaften und Rohstoffe (BGR), Hannover.
- [28.] Moores, E.M. and Fairbridge, R.W. (Eds.), 1998. Encyclopedia of European and Asian Regional Geology. Chapman & Hall. London.
- [29.] IUSS Working Group WRB, 2014. 'World Reference Base for Soil Resources 2014'. World Soil Resources Report No. 106, FAO, Rome.
- [30.] Clapp, C.E., Hayes, M.H.B., Senesi, N., Bloom, P.R. and Jardine, P.M. (Eds.), 2001. Humic Substances and Chemical Contaminants. Soil Science Society of America, Madison, WI, USA.
- [31.] Tóth, G., Gardi, C., Bódis, K., Ivits, É., Aksoy, E., Jones, A., Jeffrey, S., Petursdottir, T., Montanarella, L., 2013. Continental-scale assessment of provisioning soil functions in Europe. Ecological Processes. 2:32. doi:10.1186/2192-1709-2-32
- [32.] Kutilek, M. and Nielsen, D.R., 1994. Soil Hydrology. Catena Verlag. Cremlingen-Destedt.
- [33.] Kumar, R. and van Sloun, J.K., 1989. Purification by adsorptive separation. Chemical Engineering Progress. 85 (1): 34–40.
- [34.] Ives, K.J., 1990. Testing of filter media. Aqua. 39 (3): 144–151.
- [35.] Helling, C.S., Chesters, G. and Corey, R.B., 1964. Contribution of organic matter and clay to soil cation-exchange capacity as affected by the pH of the saturating solution. Soil Science Society of America Journal. 28 (4): 517–520.
- [36.] Buckman, H.O. and Brady, N.C., 1972. The nature and properties of soils. London. The MacMillan Company.
- [37.] Sethi, R.K. and Chopra, S.L., 1977. Adsorption and the behavior of pesticides in soils. Pesticides. 11. 15–25.
- [38.] Greenland, D.J. and Hayes, M.B.H., 1981. The chemistry of soil processes. New York. John Wiley & Sons.
- [39.] Dixon, J.B. and Weed, S.B. (eds.), 1977. Minerals in soil environments. Soil Science Society of America. Madison. WI.
- [40.] Peech, M., 1939. Chemical studies on soils from Florida citrus groves. Agricultural Experiment Stations Bulletin, University of Florida. 340. 1–50.
- [41.] Loynachan, T. E., 1981. Lime requirement methods for cold-region soils. Soil Science Society of America Journal. 45. 75–80.
- [42.] Magdoff, F. R., Bartlett, R. J., 1985. Soil pH buffering revisited. Soil Science Society of America Proceedin. 49. 145–148.
- [43.] Anderson, M.A. and Rubin, A.J., 1981. Adsorption of inorganics at solid-liquid interfaces. Ann Arbor Science Publishers, Inc., Ann Arbor, MI.
- [44.] Xu, J. and Sparks, D.L. (eds.), 2013. Molecular environmental science. Progress in soil science. Springer. Dordrecht.
- [45.] Haggerty, G.M. and Bowman, R.S., 1994. Sorption of chromate and other inorganic anions by organo-zeolite. Environmental Science & Technology. 28 (3): 452–458.
- [46.] Naghavi, B. and Malone, R.F., 1986. Algae removal by fine sand/silt filtration. Water Research. 20 (3): 377–383.
- [47.] Gerba, C.P., Haas, C.N., 1988. Assessment of risks associated with enteric viruses in contaminated drinking water. In: Chemical and biological characterization of sludges, sediments, drudge spoils and drilling muds, ASTM STP 976. Philadelphia. American Society for Testing and Materials. 489–494.

- [48.] Gross, M.A. and Mitchell, D., 1990. Virus removal by sand filtration of septic tank effluent. *Journal of Environmental Engineering*. 116 (4): 711–720.
- [49.] Marks, R., 1989. Anleitung zur Bewertung des Leistungsvermögens des Landschaftshaushaltes. *Forschungen zur deutschen Landeskunde*, 1. Auflage, Trier.
- [50.] Schachtschabel, P., Blume, H.-P., Brümmer, G., Hartge, K.-H., Schwertmann, U., Fischer, W. R. et al., 1992. *Lehrbuch der Bodenkunde*. 13. Auflage; Ferdinand Enke Verlag, Stuttgart.
- [51.] Clark, S. and Pitt., R., 1999. Stormwater runoff treatment: evaluation of filtration media. EPA-600/R-00/010; U.S. Environmental Protection Agency, Water Supply and Water Resources Division, National Risk Management Research Laboratory, Cincinnati, OH.
- [52.] Jury, W.A., Spencer, W.F. and Farmer, W.J., 1983. Model for assessing behavior of pesticides and other trace organics using benchmark properties. I. Description of model. *Journal of Environmental Quality* 12. 558–564.
- [53.] Crites, R.W., 1985. Micropollutant removal in rapid infiltration. In: Takashi, A. (ed): *Artificial recharge of groundwater*. Butterworth Publishers. Boston. 579-608.
- [54.] Ruthven, D.M. and Kaul, B.K., 1993a. Adsorption of aromatic hydrocarbons in NaX zeolite. 1. Equilibrium. *Industrial Engineering and Chemistry Research*. 32. 2047–2052.
- [55.] Ruthven, D.M. and Kaul, B.K., 1993b. Adsorption of aromatic hydrocarbons in NaX zeolite. 2. Kinetics. *Industrial Engineering and Chemistry Research*. 32. 2053–2057.
- [56.] Mayer, A.S. and Miller, C.T., 1992. The influence of porous medium characteristics and measurement scale on pore-scale distributions of residual nonaqueous-phase liquids. *Journal of Contaminant Hydrology*. 11 (3-4): 189–213.
- [57.] Makó, A., 2005. Measuring the two-phase capillary pressure-saturation curves of soil samples saturated with nonpolar liquids. *Communication in Soil Science and Plant Analysis*. 36. 439–453.
- [58.] Hernádi, H. and Makó, A., 2014. Preliminary investigation to estimate soil NAPL retention using parametric pedotransfer functions. *International Agrophysics*. 28. 435–445.
- [59.] Cohen, A.D., Rollins, M.S., Zunic, W.M. and Durig, J.R., 1991. Effects of chemical and physical differences in peats on their ability to extract hydrocarbons from water. *Water Research* 25 (9): 1047–1060.
- [60.] Rael, J., Shelton, S., Dayaye, R., 1995. Permeable barriers to remove benzene: candidate media evaluation. *Journal of Environmental Engineering* 121 (5). 411-415.
- [61.] Roekaerts, M., 2002. *The biogeographical regions map of Europe*. European Environment Agency

List of abbreviations and definitions

Abbreviations Full name of parameters

BGR	Biogeographical region
ESDB	European soil database
NAPL	Nonaqueous phase liquid
PA	Percentage area
PTRDB	Pedotransfer Rules database
SMU	Soil mapping unit
STU	Soil taxonomy unit
WCI	Weightened capacity index

Input parameters

BS_SUB	Base saturation of the subsoil
BS_TOP	Base saturation of the topsoil
CLIMANR	Code for the grouped climate categories
CORGRP_CAT	Code for the grouped CORINE 2006 categories
DR	Depth to rock
IL	Code for the presence of an impermeable layer within the soil profile of the STU
MIN_SUB	Subsoil mineralogy
MIN_TOP	Topsoil mineralogy
OC_TOP	Topsoil organic carbon content
PARMADO	Code for dominant parent material of the STU
PARMASE	Code for secondary parent material of the STU
PD_SUB	Subsoil packing density
PD_TOP	Topsoil packing density
PEAT	Peat
SLOPE-DOM	Dominant slope class of the STU
SLOPE-SEC	Secondary Slope class of the STU
TEXTDEPCHG	Depth class to a textural change of the dominant and/or secondary surface texture of the STU
TEXTSRFDOM	Dominant surface textural class of the STU
TEXTSRFSEC	Secondary surface textural class of the STU
TEXTSUBDOM	Dominant sub-surface textural class of the STU
TEXTSUBSEC	Secondary sub-surface textural class of the STU
VS	Volume of stones
WRB_ADJ	First soil adjective code of the STU from the World Reference Base (WRB) for Soil Resources
WRB_GRP	Soil reference group code of the STU from the World Reference Base (WRB) for Soil Resources
WRBFU	Full soil code of the STU from the World Reference Base (WRB) for Soil Resources

Estimated parameters

AER	Aeration code
ANS_f	Anion sorption factor
BASE_SAT	Base saturation of the soil profile
CATS_f	Cation sorption factor
CEC_HUM	Cation exchange capacity related to humus content
CEC_MIN	Cation exchange capacity related to the soil minerals
CEC_SUM	Cation exchange capacity related to soil minerals and humus content
CPS	Clay percentage of the subsoil
CPT	Clay percentage of the topsoil
DG	Depth to the groundwater
DG_f	Groundwater depth factor

DISR	Depth to the impermeable rock
FILT_CAPAN	Anion filtering capacity (elements in anionic form and organic chemicals with negative charges)
FILT_CAPCA	Cation filtering capacity (elements in cationic form and organic chemicals with positive charges)
FILT_CAPNP	Non-polar organic chemicals filtering capacity
FILT_CAPSO	Solids and pathogenic microorganisms filtering capacity
FILT_NAPL	Nonaqueous Phase Liquids (NAPL) filtering capacity
HLT	Thickness of humus layer
HQ	Humus quality code
IL_f	Impermeable layer factor
IR	Infiltration rate
IR_f	Infiltration rate factor
LEACH_f	Leaching factor
MCPS	Modified clay percentage of the subsoil
MCPT	Modified clay percentage of the topsoil
MDSP	Maximum depth of soil profile with filter and storage properties
MIN_f	Mineral factor
MTDCH	Modified depth to a textural change code
SLOPEM	Modified slope category
SPCC	Clay content of the soil profile
SPD_f	Subsoil packing density factor
SPHC	Humus content of soil profile
SSA_HUM	Specific surface area related to humus content
SSA_MIN	Specific surface area related to soil minerals
SSA_SUM	Specific surface area related to soil minerals and humus content
STOR_CAPAN	Anion storing capacity (elements in anionic form and organic chemicals with negative charges)
STOR_CAPCA	Cation storing capacity (elements in cationic form and organic chemicals with positive charges)
STOR_CAPNP	Non-polar organic chemicals storing capacity
STOR_CAPSO	Solids and pathogenic microorganisms storing capacity
STOR_NAPL	Nonaqueous Phase Liquids (NAPL) storing capacity
TPD_f	Topsoil packing density factor
WB	Water balance code
WBM	Modified water balance

Input parameters used in the estimation procedures

TEXTDEPCHG: Depth class to a textural change of the dominant and/or secondary surface texture of the STU	6 Textural change between 20 and 60 cm depth
-----	7 Textural change between 60 and 120 cm depth
0 No information	-----
1 Textural change between 20 and 40 cm depth	TEXTSRFDOM: Dominant surface textural class of the STU
2 Textural change between 40 and 60 cm depth	-----
3 Textural change between 60 and 80 cm depth	0 No information
4 Textural change between 80 and 120 cm depth	9 No mineral texture (Peat soils)
5 No textural change between 20 and 120 cm depth	1 Coarse (18% < clay and > 65% sand)
	2 Medium (18% < clay < 35% and >= 15% sand, or 18% < clay and 15% < sand < 65%)
	3 Medium fine (< 35% clay and < 15% sand)

- 4 Fine (35% < clay < 60%)
- 5 Very fine (clay > 60 %)

 TEXTSRFSEC: Secondary surface
 textural class of the STU

- 0 No information
- 9 No mineral texture (Peat soils)
- 1 Coarse (18% < clay and > 65% sand)
- 2 Medium (18% < clay < 35% and >= 15% sand, or 18% < clay and 15% < sand < 65%)
- 3 Medium fine (< 35% clay and < 15% sand)
- 4 Fine (35% < clay < 60%)
- 5 Very fine (clay > 60 %)

 TEXTSUBDOM: Dominant sub-surface
 textural class of the STU

- 0 No information
- 9 No mineral texture (Peat soils)
- 1 Coarse (18% < clay and > 65% sand)
- 2 Medium (18% < clay < 35% and >= 15% sand, or 18% < clay and 15% < sand < 65%)
- 3 Medium fine (< 35% clay and < 15% sand)
- 4 Fine (35% < clay < 60%)
- 5 Very fine (clay > 60 %)

 TEXTSUBSEC :Secondary sub-surface
 textural class of the STU

- 0 No information
- 9 No mineral texture (Peat soils)
- 1 Coarse (18% < clay and > 65% sand)
- 2 Medium (18% < clay < 35% and >= 15% sand, or 18% < clay and 15% < sand < 65%)
- 3 Medium fine (< 35% clay and < 15% sand)
- 4 Fine (35% < clay < 60%)
- 5 Very fine (clay > 60 %)

 PARMADO: Code for dominant parent
 material of the STU

- 0 No information
- 1000 consolidated-clastic-sedimentary rocks
- 1100 psephite or rudite
- 1110 conglomerate

- 1111 pudding stone
- 1120 breccia
- 1200 psammite or arenite
- 1210 sandstone
- 1211 calcareous sandstone
- 1212 ferruginous sandstone
- 1213 clayey sandstone
- 1214 quartzitic sandstone/orthoquartzite
- 1215 micaceous sandstone
- 1220 arkose
- 1230 graywacke
- 1231 feldspathic graywacke
- 1300 pelite, lutite or argillite
- 1310 claystone/mudstone
- 1311 kaolinite
- 1312 bentonite
- 1320 siltstone
- 1400 facies bound rock
- 1410 flysch
- 1411 sandy flysch
- 1412 clayey and silty flysch
- 1413 conglomeratic flysch
- 1420 molasse
- 2000 sedimentary rocks (chemically precipitated, evaporated, or organogenic or biogenic in origin)
- 2100 calcareous rocks
- 2110 limestone
- 2111 hard limestone
- 2112 soft limestone
- 2113 marly limestone
- 2114 chalky limestone
- 2115 detrital limestone
- 2116 carbonaceous limestone
- 2117 lacustrine or freshwater limestone
- 2118 travertine/calcareous sinter
- 2119 cavernous limestone
- 2120 dolomite
- 2121 cavernous dolomite
- 2122 calcareous dolomite
- 2130 marlstone
- 2140 marl
- 2141 chalk marl
- 2142 gypsiferous marl
- 2150 chalk
- 2200 evaporites
- 2210 gypsum
- 2220 anhydrite
- 2230 halite
- 2300 siliceous rocks
- 2310 chert, hornstone, flint
- 2320 diatomite/radiolarite
- 3000 igneous rocks
- 3100 acid to intermediate plutonic rocks
- 3110 granite
- 3120 granodiorite

3130 diorite	4313 talc schist
3131 quartz diorite	4320 amphibolite
3132 gabbro diorite	4330 eclogite
3140 syenite	4400 ultrabasic regional metamorphic rocks
3200 basic plutonic rocks	4410 serpentinite
3210 gabbro	4411 greenstone
3300 ultrabasic plutonic rocks	4500 calcareous regional metamorphic rocks
3310 peridotite	4510 marble
3320 pyroxenite	4520 calcschist, skam
3400 acid to intermediate volcanic rocks	4600 rocks formed by contact metamorphism
3410 rhyolite	4610 contact slate
3411 obsidian	4611 nodular slate
3412 quartz porphyrite	4620 hornfels
3420 dacite	4630 calsilicate rocks
3430 andesite	4700 tectogenetic metamorphism rocks or cataclasmic metamorphism
3431 porphyrite (interm,)	4710 tectonic breccia
3440 phonolite	4720 cataclasite
3441 tephritic phonolite	4730 mylonite
3450 trachyte	5000 unconsolidated deposits (alluvium, weathering residuum and slope deposits)
3500 basic to ultrabasic volcanic rocks	5100 marine and estuarine sands
3510 basalt	5110 pre-quatarnary sand
3520 diabase	5111 tertiary sand
3530 pikrite	5120 quaternary sand
3600 dike rocks	5121 holocene coastal sand with shells
3610 aplite	5122 delta sand
3620 pegmatite	5200 marine and estuarine clays and silts
3630 lamprophyre	5210 pre-quatarnary clay and silt
3700 pyroclastic rocks (tephra)	5211 tertiary clay
3710 tuff/tuffstone	5212 tertiary silt
3711 agglomeratic tuff	5220 quaternary clay and silt
3712 block tuff	5221 holocene clay
3713 lapilli tuff	5222 holocene silt
3720 tuffite	5300 fluvial sands and gravels
3721 sandy tuffite	5310 river terrace sand or gravel
3722 silty tuffite	5311 river terrace sand
3723 clayey tuffite	5312 river terrace gravel
3730 volcanic scoria/volcanic breccia	5320 floodplain sand or gravel
3740 volcanic ash	5321 floodplain sand
3750 ignimbrite	5322 floodplain gravel
3760 pumice	5400 fluvial clays, silts and loams
4000 metamorphic rocks	5410 river clay and silt
4100 weakly metamorphic rocks	5411 terrace clay and silt
4110 (meta-)shale/argillite	5412 floodplain clay and silt
4120 slate	5420 river loam
4121 graphitic slate	5421 terrace loam
4200 acid regional metamorphic rocks	5430 overbank deposit
4210 (meta-)quartzite	5431 floodplain clay and silt
4211 quartzite schist	5432 floodplain loam
4220 phyllite	5500 lake deposits
4230 micaschist	5510 lake sand and delta sand
4240 gneiss	5520 lake marl, bog lime
4250 granulite (sensu stricto)	
4260 migmatite	
4300 basic regional metamorphic rocks	
4310 greenschist	
4311 prasinite	
4312 chlorite	

5530 lake silt	9100 redeposited natural materials
5600 residual and redeposited loams from silicate rocks	9110 sand and gravel fill
5610 residual loam	9120 loamy fill
5611 stony loam	9200 dump deposits
5612 clayey loam	9210 rubble/rubbish
5620 redeposited loam	9220 industrial ashes and slag
5621 running-ground	9230 industrial sludge
5700 residual and redeposited clays from calcareous rocks	9240 industrial waste
5710 residual clay	9300 anthropogenic organic materials
5711 clay with flints	-----
5712 ferruginous residual clay	-----
5713 calcareous clay	PARMASE: Code for secondary parent material of the STU
5714 non-calcareous clay	-----
5715 marly clay	0 No information
5720 redeposited clay	1000 consolidated-clastic-sedimentary rocks
5721 stony clay	1100 psephite or rudite
5800 slope deposits	1110 conglomerate
5810 slope-wash alluvium	1111 pudding stone
5820 colluvial deposit	1120 breccia
5830 talus scree	1200 psammite or arenite
5831 stratified slope deposits	1210 sandstone
6000 unconsolidated glacial deposits/glacial drift	1211 calcareous sandstone
6100 morainic deposits	1212 ferruginous sandstone
6110 glacial till	1213 clayey sandstone
6111 boulder clay	1214 quartzitic sandstone/orthoquartzite
6120 glacial debris	1215 micaceous sandstone
6200 glaciofluvial deposits	1220 arkose
6210 outwash sand, glacial sand	1230 graywacke
6220 outwash gravels glacial gravels	1231 feldspathic graywacke
6300 glaciolacustrine deposits	1300 pelite, lutite or argilite
6310 varves	1310 claystone/mudstone
7000 eolian deposits	1311 kaolinite
7100 loess	1312 bentonite
7110 loamy loess	1320 siltstone
7120 sandy loess	1400 facies bound rock
7200 eolian sands	1410 flysch
7210 dune sand	1411 sandy flysch
7220 cover sand	1412 clayey and silty flysch
8000 organic materials	1413 conglomeratic flysch
8100 peat (mires)	1420 molasse
8110 rainwater fed moor peat (raised bog)	2000 sedimentary rocks (chemically precipitated, evaporated, or organogenic or biogenic in origin)
8111 folic peat	2100 calcareous rocks
8112 fibric peat	2110 limestone
8113 terric peat	2111 hard limestone
8120 groundwater fed bog peat	2112 soft limestone
8200 slime and ooze deposits	2113 marly limestone
8210 gyttja, sapropel	2114 chalky limestone
8300 carbonaceous rocks (caustobiolite)	2115 detrital limestone
8310 lignite (brown coal)	2116 carbonaceous limestone
8320 hard coal	2117 lacustrine or freshwater limestone
8330 anthracite	2118 travertine/calcareous sinter
9000 anthropogenic deposits	2119 cavernous limestone

2120 dolomite	3750 ignimbrite
2121 cavernous dolomite	3760 pumice
2122 calcareous dolomite	4000 metamorphic rocks
2130 marlstone	4100 weakly metamorphic rocks
2140 marl	4110 (meta-)shale/argilite
2141 chalk marl	4120 slate
2142 gypsiferous marl	4121 graphitic slate
2150 chalk	4200 acid regional metamorphic rocks
2200 evaporites	4210 (meta-)quartzite
2210 gypsum	4211 quartzite schist
2220 anhydrite	4220 phyllite
2230 halite	4230 micaschist
2300 siliceous rocks	4240 gneiss
2310 chert, hornstone, flint	4250 granulite (sensu stricto)
2320 diatomite/radiolarite	4260 migmatite
3000 igneous rocks	4300 basic regional metamorphic rocks
3100 acid to intermediate plutonic rocks	4310 greenschist
3110 granite	4311 prasinite
3120 granodiorite	4312 chlorite
3130 diorite	4313 talc schist
3131 quartz diorite	4320 amphibolite
3132 gabbro diorite	4330 eclogite
3140 syenite	4400 ultrabasic regional metamorphic rocks
3200 basic plutonic rocks	4410 serpentinite
3210 gabbro	4411 greenstone
3300 ultrabasic plutonic rocks	4500 calcareous regional metamorphic rocks
3310 peridotite	4510 marble
3320 pyroxenite	4520 calcschist, skam
3400 acid to intermediate volcanic rocks	4600 rocks formed by contact metamorphism
3410 rhyolite	4610 contact slate
3411 obsidian	4611 nodular slate
3412 quartz porphyrite	4620 hornfels
3420 dacite	4630 calsilicate rocks
3430 andesite	4700 tectogenetic metamorphism rocks or cataclasmic metamorphism
3431 porphyrite (interm,)	4710 tectonic breccia
3440 phonolite	4720 cataclasite
3441 tephritic phonolite	4730 mylonite
3450 trachyte	5000 unconsolidated deposits (alluvium, weathering residuum and slope deposits)
3500 basic to ultrabasic volcanic rocks	5100 marine and estuarine sands
3510 basalt	5110 pre-quadernary sand
3520 diabase	5111 tertiary sand
3530 pikrite	5120 quadernary sand
3600 dike rocks	5121 holocene coastal sand with shells
3610 aplite	5122 delta sand
3620 pegmatite	5200 marine and estuarine clays and silts
3630 lamprophyre	5210 pre-quadernary clay and silt
3700 pyroclastic rocks (tephra)	5211 tertiary clay
3710 tuff/tuffstone	5212 tertiary silt
3711 agglomeratic tuff	5220 quadernary clay and silt
3712 block tuff	5221 holocene clay
3713 lapilli tuff	
3720 tuffite	
3721 sandy tuffite	
3722 silty tuffite	
3723 clayey tuffite	
3730 volcanic scoria/volcanic breccia	
3740 volcanic ash	

5222 holocene silt	7120 sandy loess
5300 fluvial sands and gravels	7200 eolian sands
5310 river terrace sand or gravel	7210 dune sand
5311 river terrace sand	7220 cover sand
5312 river terrace gravel	8000 organic materials
5320 floodplain sand or gravel	8100 peat (mires)
5321 floodplain sand	8110 rainwater fed moor peat (raised bog)
5322 floodplain gravel	8111 folic peat
5400 fluvial clays, silts and loams	8112 fibric peat
5410 river clay and silt	8113 terric peat
5411 terrace clay and silt	8120 groundwater fed bog peat
5412 floodplain clay and silt	8200 slime and ooze deposits
5420 river loam	8210 gyttja, sapropel
5421 terrace loam	8300 carbonaceous rocks (caustobiolite)
5430 overbank deposit	8310 lignite (brown coal)
5431 floodplain clay and silt	8320 hard coal 8330 anthracite
5432 floodplain loam	9000 anthropogenic deposits
5500 lake deposits	9100 redeposited natural materials
5510 lake sand and delta sand	9110 sand and gravel fill
5520 lake marl, bog lime	9120 loamy fill
5530 lake silt	9200 dump deposits
5600 residual and redeposited loams from silicate rocks	9210 rubble/rubbish
5610 residual loam	9220 industrial ashes and slag
5611 stony loam	9230 industrial sludge
5612 clayey loam	9240 industrial waste
5620 redeposited loam	9300 anthropogenic organic materials
5621 running-ground	-----
5700 residual and redeposited clays from calcareous rocks	-----
5710 residual clay	WRB_GRP: Soil reference group code of the STU from the World Reference Base (WRB) for Soil Resources
5711 clay with flints	-----
5712 ferruginous residual clay	AB Albeluvisol
5713 calcareous clay	AC Acrisol
5714 non-calcareous clay	AL Alisol
5715 marly clay	AN Andosol
5720 redeposited clay	AR Arenosol
5721 stony clay	AT Anthrosol
5800 slope deposits	CH Chernozem
5810 slope-wash alluvium	CL Calcisol
5820 colluvial deposit	CM Cambisol
5830 talus scree	CR Cryosol
5831 stratified slope deposits	DU Durisol
6000 unconsolidated glacial deposits/glacial drift	FL Fluvisol
6100 morainic deposits	FR Ferralsol
6110 glacial till	GL Gleysol
6111 boulder clay	GY Gypsisol
6120 glacial debris	HS Histosol
6200 glaciofluvial deposits	KS Kastanozem
6210 outwash sand, glacial sand	LP Leptosol
6220 outwash gravels glacial gravels	LV Luvisol
6300 glaciolacustrine deposits	LX Lixisol
6310 varves	NT Nitisol
7000 eolian deposits	PH Phaeozem
7100 loess	PL Planosol
7110 loamy loess	

PT Plinthosol
PZ Podzol
RG Regosol
SC Solonchak
SN Solonetz
UM Umbrisol
VR Vertisol
1 Town
2 Soil disturbed by man
3 Water body
4 Marsh
5 Glacier
6 Rock outcrops

WRB_ADJ: First soil adjective code of
the STU from the World Reference Base
(WRB) for Soil Resources

II Lamellic
Iv Luvic
Ix Lixic
ab Albic
ac Acric
ad Aridic
ae Aceric
ah Anthropic
ai Aric
al Alic
am Anthric
an Andic
ao Acroxic
ap Abruptic
aq Anthraquic
ar Arenic
au Alumic
ax Alcalic
az Arzic
ca Calcaric
cb Carbic
cc Calcic
ch Chernic
cl Chloridic
cn Carbonatic
cr Chromic
ct Cutanic
cy Cryic
dn Densic
du Duric
dy Dystric
es Eutrisilic
et Entic
eu Eutric
fg Fragic
fi Fibric
fl Ferralic
fo Folic

fr Ferric
fu Fulvic
fv Fluvic
ga Garbic
gc Glacic
ge Gelic
gi Gibbsic
gl Gleyic
gm Grumic
gp Gypsiric
gr Geric
gs Glossic
gt Gelistagnic
gy Gypsic
gz Greyic
ha Haplic
hg Hydragric
hi Histic
hk Hyperskeletal
ht Hortic
hu Humic
hy Hydric
ir Irragric
le Leptic
li Lithic
me Melanic
mg Magnestic
mo Mollic
ms Mesotrophic
mz Mazic
na Natric
ni Nitic
oa Oxyaquic
oh Ochric
om Ombric
or Orthic
pa Plaggic
pc Petroccic
pd Petroduric
pe Pellic
pf Profondic
pg Petrogypsic
ph Pachic
pi Placic
pl Plinthic
pn Planic
po Posic
pp Petroplinthic
pr Protic
ps Petrosalic
pt Petric
rd Reductic
rg Regic
rh Rheic
ro Rhodic
rp Ruptic
rs Rustic

ru Rubic
 rz Rendzic
 sa Sapric
 sd Spodic
 si Silic
 sk Skeletic
 sl Siltic
 so Sodic
 sp Spolic
 st Stagnic
 su Sulphatic
 sz Salic
 tf Tephric
 ti Thionic
 tr Terric
 tu Turbic
 tx Toxic
 ty Takyric
 ub Urbic
 um Umbric
 vi Vitric
 vm Vermic
 vr Vertic
 vt Vetic
 xa Xanthic
 ye Yermic
 1 Town
 2 Soil disturbed by man
 3 Water body
 4 Marsh
 5 Glacier
 6 Rock outcrops

 WRBFU: Full soil code of the STU from
 the World Reference Base (WRB) for Soil
 Resources

AB Albeluvisol
 ABal Alic Albeluvisol
 ABap Abruptic Albeluvisol
 ABar Arenic Albeluvisol
 ABau Aluminic Albeluvisol
 ABeun Endoeutric Albeluvisol
 ABfg Fragic Albeluvisol
 ABfr Ferric Albeluvisol
 ABge Gelic Albeluvisol
 ABgl Gleyic Albeluvisol
 ABha Haplic Albeluvisol
 ABhi Histic Albeluvisol
 ABsl Siltic Albeluvisol
 ABst Stagnic Albeluvisol
 ABum Umbric Albeluvisol
 AC Acrisol
 ACab Albic Acrisol
 ACan Andic Acrisol
 ACap Abruptic Acrisol

ACar Arenic Acrisol
 ACau Aluminic Acrisol
 ACcr Chromic Acrisol
 ACdyh Hyperdystric Acrisol
 ACfr Ferric Acrisol
 ACgl Gleyic Acrisol
 ACgr Geric Acrisol
 ACha Haplic Acrisol
 AChu Humic Acrisol
 ACle Leptic Acrisol
 ACll Lamellic Acrisol
 ACohh Hyperochric Acrisol
 ACpf Profondic Acrisol
 ACpl Plinthic Acrisol
 ACro Rhodic Acrisol
 ACsk Skeletic Acrisol
 ACst Stagnic Acrisol
 ACum Umbric Acrisol
 ACvi Vitric Acrisol
 ACvt Vetic Acrisol
 AL Alisol
 ALab Albic Alisol
 ALan Andic Alisol
 ALap Abruptic Alisol
 ALar Arenic Alisol
 ALcr Chromic Alisol
 ALdyh Hyperdystric Alisol
 ALfr Ferric Alisol
 ALgl Gleyic Alisol
 ALha Haplic Alisol
 ALhu Humic Alisol
 ALll Lamellic Alisol
 ALni Nitic Alisol
 ALpf Profondic Alisol
 ALpl Plinthic Alisol
 ALro Rhodic Alisol
 ALsk Skeletic Alisol
 ALst Stagnic Alisol
 ALum Umbric Alisol
 ALvr Vertic Alisol
 AN Andosol
 ANao Acroxic Andosol
 ANar Arenic Andosol
 ANca Calcaric Andosol
 ANdu Duric Andosol
 ANdy Dystric Andosol
 ANes Eutrisilic Andosol
 ANeu Eutric Andosol
 ANfu Fulvic Andosol
 ANgl Gleyic Andosol
 ANha Haplic Andosol
 ANhi Histic Andosol
 ANhy Hydric Andosol
 ANle Leptic Andosol
 ANlv Luvic Andosol
 ANme Melanic Andosol
 ANmo Mollic Andosol

ANph Pachic Andosol
 ANpi Placic Andosol
 ANsi Silic Andosol
 ANsk Skeletic Andosol
 ANso Sodic Andosol
 ANth Thaptic Andosol
 ANum Umbric Andosol
 ANvi Vitric Andosol
 ANvt Vetic Andosol
 AR Arenosol
 ARab Albic Arenosol
 ARad Aridic Arenosol
 ARca Calcaric Arenosol
 ARduw Hypoduric Arenosol
 ARdy Dystric Arenosol
 AREu Eutric Arenosol
 ARfg Fragic Arenosol
 ARfl Ferralic Arenosol
 ARge Gelic Arenosol
 ARgl Gleyic Arenosol
 ARgp Gypsiric Arenosol
 ARha Haplic Arenosol
 ARll Lamellic Arenosol
 ARlvw Hypoluvisc Arenosol
 ARpl Plinthic Arenosol
 ARpr Protic Arenosol
 ARru Rubic Arenosol
 ARszw Hyposalic Arenosol
 ARTf Tephric Arenosol
 ARye Yermic Arenosol
 AT Anthrosol
 ATar Arenic Anthrosol
 ATfl Ferralic Anthrosol
 ATgl Gleyic Anthrosol
 ATHg Hydragric Anthrosol
 ATht Hortic Anthrosol
 ATir Irragric Anthrosol
 ATlv Luvic Anthrosol
 ATpa Plaggic Anthrosol
 ATrg Regic Anthrosol
 ATsd Spodic Anthrosol
 ATst Stagnic Anthrosol
 ATtr Terric Anthrosol
 CH Chernozem
 CHcc Calcic Chernozem
 CHch Chernic Chernozem
 CHgl Gleyic Chernozem
 CHgs Glossic Chernozem
 CHha Haplic Chernozem
 CHlv Luvic Chernozem
 CHsl Siltic Chernozem
 CHvm Vermic Chernozem
 CHvr Vertic Chernozem
 CL Calcisol
 CLad Aridic Calcisol
 CLcch Hypercalcic Calcisol
 CLccw Hypocalcic Calcisol

CLgl Gleyic Calcisol
 CLha Haplic Calcisol
 CLle Leptic Calcisol
 CLlv Luvic Calcisol
 CLohh Hyperochric Calcisol
 CLpt Petric Calcisol
 CLsk Skeletic Calcisol
 CLso Sodic Calcisol
 CLszn Endosalic Calcisol
 CLty Takyric Calcisol
 CLvr Vertic Calcisol
 CLye Yermic Calcisol
 CM Cambisol
 CMad Aridic Cambisol
 CMan Andic Cambisol
 CMca Calcaric Cambisol
 CMcr Chromic Cambisol
 CMdy Dystric Cambisol
 CMeu Eutric Cambisol
 CMfl Ferralic Cambisol
 CMfv Fluvic Cambisol
 CMge Gelic Cambisol
 CMgl Gleyic Cambisol
 CMgp Gypsiric Cambisol
 CMgt Gelistagnic Cambisol
 CMha Haplic Cambisol
 CMle Leptic Cambisol
 CMmo Mollic Cambisol
 CMohh Hyperochric Cambisol
 CMpl Plinthic Cambisol
 CMro Rhodic Cambisol
 CMsk Skeletic Cambisol
 CMso Sodic Cambisol
 CMst Stagnic Cambisol
 CMszn Endosalic Cambisol
 CMty Takyric Cambisol
 CMvi Vitric Cambisol
 CMvr Vertic Cambisol
 CMye Yermic Cambisol
 CR Cryosol
 CRad Aridic Cryosol
 CRan Andic Cryosol
 CRcc Calcic Cryosol
 CRgc Glacic Cryosol
 CRgl Gleyic Cryosol
 CRgy Gypsic Cryosol
 CRha Haplic Cryosol
 CRhi Histic Cryosol
 CRle Leptic Cryosol
 CRLi Lithic Cryosol
 CRmo Mollic Cryosol
 CRna Natric Cryosol
 CRoa Oxyaquic Cryosol
 CRst Stagnic Cryosol
 CRsz Salic Cryosol
 CRTi Thionic Cryosol
 CRTu Turbic Cryosol

CRum Umbric Cryosol
 CRye Yermic Cryosol
 DU Durisol
 DUad Aridic Durisol
 DUar Arenic Durisol
 DUcc Calcic Durisol
 DUcr Chromic Durisol
 DUgy Gypsic Durisol
 DUha Haplic Durisol
 DUle Leptic Durisol
 DUlv Luvic Durisol
 DUohh Hyperochric Durisol
 DUpt Petric Durisol
 DUty Takyric Durisol
 DUvr Vertic Durisol
 DUye Yermic Durisol
 FL Fluvisol
 FLad Aridic Fluvisol
 FLar Arenic Fluvisol
 FLca Calcaric Fluvisol
 FLdy Dystric Fluvisol
 FLeu Eutric Fluvisol
 FLge Gelic Fluvisol
 FLgl Gleyic Fluvisol
 FLgp Gypsiric Fluvisol
 FLha Haplic Fluvisol
 FLhi Histic Fluvisol
 FLhu Humic Fluvisol
 FLmo Mollic Fluvisol
 FLsk Skeletic Fluvisol
 FLso Sodic Fluvisol
 FLst Stagnic Fluvisol
 FLsz Salic Fluvisol
 FLtf Tephric Fluvisol
 FLti Thionic Fluvisol
 FLty Takyric Fluvisol
 FLum Umbric Fluvisol
 FLye Yermic Fluvisol
 FR Ferralsol
 FRac Acric Ferralsol
 FRan Andic Ferralsol
 FRar Arenic Ferralsol
 FRau Alumic Ferralsol
 FRdyh Hyperdystric Ferralsol
 FREuh Hypereutric Ferralsol
 FRfr Ferric Ferralsol
 FRgi Gibbsic Ferralsol
 FRgl Gleyic Ferralsol
 FRgr Geric Ferralsol
 FRha Haplic Ferralsol
 FRhi Histic Ferralsol
 FRhu Humic Ferralsol
 FRlx Lixic Ferralsol
 FRmo Mollic Ferralsol
 FRpl Plinthic Ferralsol
 FRpo Posic Ferralsol
 FRro Rhodic Ferralsol

FRstn Endostagnic Ferralsol
 FRum Umbric Ferralsol
 FRvt Vetic Ferralsol
 FRxa Xanthic Ferralsol
 GL Gleysol
 GLan Andic Gleysol
 GLap Abruptic Gleysol
 GLaq Anthraquic Gleysol
 GLar Arenic Gleysol
 GLau Alumic Gleysol
 GLax Alcalic Gleysol
 GLca Calcaric Gleysol
 GLcc Calcic Gleysol
 GLdy Dystric Gleysol
 GLeu Eutric Gleysol
 GLge Gelic Gleysol
 GLgy Gypsic Gleysol
 GLha Haplic Gleysol
 GLhi Histic Gleysol
 GLhu Humic Gleysol
 GLmo Mollic Gleysol
 GLpl Plinthic Gleysol
 GLso Sodic Gleysol
 GLszo Endosalic Gleysol
 GLtf Tephric Gleysol
 GLti Thionic Gleysol
 GLtx Toxic Gleysol
 GLty Takyric Gleysol
 GLum Umbric Gleysol
 GLvi Vitric Gleysol
 GY Gypsisol
 GYad Aridic Gypsisol
 GYaz Arzic Gypsisol
 GYcc Calcic Gypsisol
 GYdu Duric Gypsisol
 GYgyh Hypergypsic Gypsisol
 GYgyw Hypogypsic Gypsisol
 GYha Haplic Gypsisol
 GYle Leptic Gypsisol
 GYlv Luvic Gypsisol
 GYohh Hyperochric Gypsisol
 GYpt Petric Gypsisol
 GYsk Skeletic Gypsisol
 GYso Sodic Gypsisol
 GYszo Endosalic Gypsisol
 GYty Takyric Gypsisol
 GYvr Vertic Gypsisol
 GYye Yermic Gypsisol
 HS Histosol
 HSax Alcalic Histosol
 HScy Cryic Histosol
 HSdy Dystric Histosol
 HSeu Eutric Histosol
 HSfi Fibric Histosol
 HSfo Folic Histosol
 HSGc Glacic Histosol
 HSge Gelic Histosol

HSom Ombric Histosol
 HSrh Rheic Histosol
 HSsa Sapric Histosol
 HSsz Salic Histosol
 HSti Thionic Histosol
 HStx Toxic Histosol
 KS Kastanozem
 KSam Anthric Kastanozem
 KScC Calcic Kastanozem
 KScr Chromic Kastanozem
 KSGy Gypsic Kastanozem
 KSha Haplic Kastanozem
 KSlv Luvic Kastanozem
 KSSl Siltic Kastanozem
 KSsow Hyposodic Kastanozem
 KSvr Vertic Kastanozem
 LP Leptosol
 LPad Aridic Leptosol
 LPca Calcaric Leptosol
 LPdy Dystric Leptosol
 LPeu Eutric Leptosol
 LPge Gelic Leptosol
 LPgl Gleyic Leptosol
 LPgp Gypsiric Leptosol
 LPha Haplic Leptosol
 LPhk Hyperskeletal Leptosol
 LPhu Humic Leptosol
 LPli Lithic Leptosol
 LPmo Mollic Leptosol
 LPrz Rendzic Leptosol
 LPum Umbric Leptosol
 LPvr Vertic Leptosol
 LPye Yermic Leptosol
 LV Luvisol
 LVab Albic Luvisol
 LVan Andic Luvisol
 LVar Arenic Luvisol
 LVcc Calcic Luvisol
 LVcr Chromic Luvisol
 LVct Cutanic Luvisol
 LVdy Dystric Luvisol
 LVfr Ferric Luvisol
 LVgl Gleyic Luvisol
 LVha Haplic Luvisol
 LVle Leptic Luvisol
 LVll Lamellic Luvisol
 LVohh Hyperochric Luvisol
 LVpf Profondic Luvisol
 LVro Rhodic Luvisol
 LVsow Hyposodic Luvisol
 LVst Stagnic Luvisol
 LVvi Vitric Luvisol
 LVvr Vertic Luvisol
 LX Lixisol
 LXab Albic Lixisol
 LXan Andic Lixisol
 LXap Abruptic Lixisol

LXar Arenic Lixisol
 LXcc Calcic Lixisol
 LXcr Chromic Lixisol
 LXfr Ferric Lixisol
 LXgl Gleyic Lixisol
 LXgr Geric Lixisol
 LXha Haplic Lixisol
 LXhu Humic Lixisol
 LXle Leptic Lixisol
 LXll Lamellic Lixisol
 LXohh Hyperochric Lixisol
 LXpf Profondic Lixisol
 LXpl Plinthic Lixisol
 LXro Rhodic Lixisol
 LXst Stagnic Lixisol
 LXvi Vitric Lixisol
 LXvt Vetic Lixisol
 NT Nitisol
 NTal Alic Nitisol
 NTan Andic Nitisol
 NTau Alumic Nitisol
 NTdy Dystric Nitisol
 NTeu Eutric Nitisol
 NTfl Ferralic Nitisol
 NTha Haplic Nitisol
 NThu Humic Nitisol
 NTmo Mollic Nitisol
 NTro Rhodic Nitisol
 NTum Umbric Nitisol
 NTVt Vetic Nitisol
 PH Phaeozem
 PHab Albic Phaeozem
 PHan Andic Phaeozem
 PHap Abruptic Phaeozem
 PHca Calcaric Phaeozem
 PHcr Chromic Phaeozem
 PHgl Gleyic Phaeozem
 PHgs Glossic Phaeozem
 PHgz Greyic Phaeozem
 PHha Haplic Phaeozem
 PHle Leptic Phaeozem
 PHlv Luvic Phaeozem
 PHph Pachic Phaeozem
 PHsk Skeletic Phaeozem
 PHsl Siltic Phaeozem
 PHso Sodic Phaeozem
 PHst Stagnic Phaeozem
 PHtf Tephric Phaeozem
 PHvi Vitric Phaeozem
 PHvm Vermic Phaeozem
 PHvr Vertic Phaeozem
 PL Planosol
 PLab Albic Planosol
 PLal Alic Planosol
 PLar Arenic Planosol
 PLau Alumic Planosol
 PLax Alcalic Planosol

PLca Calcaric Planosol
 PLcc Calcic Planosol
 PLcr Chromic Planosol
 PLdy Dystric Planosol
 PLeu Eutric Planosol
 PLfr Ferric Planosol
 PLge Gelic Planosol
 PLgl Gleyic Planosol
 PLgr Geric Planosol
 PLgy Gypsic Planosol
 PLha Haplic Planosol
 PLhi Histic Planosol
 PLlv Luvic Planosol
 PLmo Mollic Planosol
 PLpf? Petroferric Planosol
 PLpl Plinthic Planosol
 PLro Rhodic Planosol
 PLso Sodic Planosol
 PLszo Endosalic Planosol
 PLti Thionic Planosol
 PLum Umbric Planosol
 PLvr Vertic Planosol
 PT Plinthosol
 PTab Albic Plinthosol
 PTac Acric Plinthosol
 PTal Alic Plinthosol
 PTap Abruptic Plinthosol
 PTau Aluminic Plinthosol
 PTdun Endoduric Plinthosol
 PTeun Endoeutric Plinthosol
 PTfr Ferric Plinthosol
 PTgr Geric Plinthosol
 PTgs Glossic Plinthosol
 PTha Haplic Plinthosol
 PThu Humic Plinthosol
 PTph Pachic Plinthosol
 PTpt Petric Plinthosol
 PTst Stagnic Plinthosol
 PTum Umbric Plinthosol
 PTvt Vertic Plinthosol
 PZ Podzol
 PZam Anthric Podzol
 PZcb Carbic Podzol
 PZdn Densic Podzol
 PZet Entic Podzol
 PZfg Fragic Podzol
 PZge Gelic Podzol
 PZgl Gleyic Podzol
 PZha Haplic Podzol
 PZhi Histic Podzol
 PZll Lamellic Podzol
 PZpi Placic Podzol
 PZrs Rustic Podzol
 PZsk Skeletic Podzol
 PZst Stagnic Podzol
 PZum Umbric Podzol
 RG Regosol

RGad Aridic Regosol
 RGah Anthropic Regosol
 RGai Aric Regosol
 RGanb Thaptoandic Regosol
 RGar Arenic Regosol
 RGca Calcaric Regosol
 RGdy Dystric Regosol
 RGeu Eutric Regosol
 RGga Garbic Regosol
 RGge Gelic Regosol
 RGgl Gleyic Regosol
 RGgp Gypsiric Regosol
 RGgt Gelistagnic Regosol
 RGha Haplic Regosol
 RGhu Humic Regosol
 Rgle Leptic Regosol
 RGohh Hyperochric Regosol
 RGrd Reductic Regosol
 RGsk Skeletic Regosol
 RGsow Hyposodic Regosol
 RGsp Spolic Regosol
 RGst Stagnic Regosol
 RGszw Hyposalic Regosol
 RGtf Tephric Regosol
 RGty Takyric Regosol
 RGub Urbic Regosol
 RGvib Thaptovitric Regosol
 RGvm Vermic Regosol
 Rgye Yermic Regosol
 SC Solonchak
 SCad Aridic Solonchak
 SCae Aceric Solonchak
 SCcc Calcic Solonchak
 SCcl Chloridic Solonchak
 SCcn Carbonatic Solonchak
 SCdu Duric Solonchak
 SCge Gelic Solonchak
 SCgl Gleyic Solonchak
 SCgy Gypsic Solonchak
 SCha Haplic Solonchak
 SChi Histic Solonchak
 SCmo Mollic Solonchak
 SCoh Ochric Solonchak
 SCps Petrosalic Solonchak
 SCso Sodic Solonchak
 SCst Stagnic Solonchak
 SCsu Sulphatic Solonchak
 SCszh Hypersalic Solonchak
 SCty Takyric Solonchak
 SCvr Vertic Solonchak
 SCye Yermic Solonchak
 SN Solonetz
 SNab Albic Solonetz
 SNad Aridic Solonetz
 SNcc Calcic Solonetz
 SNdu Duric Solonetz
 SNgl Gleyic Solonetz

SNgy Gypsic Solonetz
 SNha Haplic Solonetz
 SNhu Humic Solonetz
 SNmg Magnesian Solonetz
 SNmo Mollic Solonetz
 SNst Stagnic Solonetz
 SNSz Salic Solonetz
 SNty Takyric Solonetz
 SNvr Vertic Solonetz
 SNye Yermic Solonetz
 UM Umbrisol
 UMaB Albic Umbrisol
 UMam Anthric Umbrisol
 UMar Arenic Umbrisol
 UMfl Ferralic Umbrisol
 UMge Gelic Umbrisol
 UMgl Gleyic Umbrisol
 UMha Haplic Umbrisol
 UMhu Humic Umbrisol
 UMle Leptic Umbrisol
 UMSk Skeletic Umbrisol
 UMst Stagnic Umbrisol
 VR Vertisol
 VRaL Alic Vertisol
 VRcc Calcic Vertisol
 VRcr Chromic Vertisol
 VRdu Duric Vertisol
 VReu Eutric Vertisol
 VRgm Grumic Vertisol
 VRgp Gypsic Vertisol
 VRgy Gypsic Vertisol
 VRha Haplic Vertisol
 VRms Mesotrophic Vertisol
 VRmz Mazic Vertisol
 VRna Natric Vertisol
 VRpe Pellic Vertisol
 VRsow Hyposodic Vertisol
 VRsz Salic Vertisol
 VRTi Thionic Vertisol
 11111 Town
 22222 Soil disturbed by man
 33333 Water body
 44444 Marsh
 55555 Glacier
 66666 Rock outcrops
 No information

 DR = Depth to rock.
 S = Shallow (< 40 cm)
 M = Moderate (40 - 80 cm)
 D = Deep (80 - 120 cm)
 V = Very deep (> 120 cm)

 VS = Volume of stones.
 00 = 0 % stones

10 = 10 % stones
 15 = 15 % stones
 20 = 20 % stones

 MIN_TOP = Topsoil mineralogy.
 KQ = 1/1 Minerals + Quartz
 KX = 1/1 Min. + Oxy. & Hydroxy.
 MK = 2/1 & 1/1 Minerals
 M = 2/1 & 2/1/1 non swel. Min.
 MS = Swel. & non swel. 2/1 Min.
 S = Swelling 2/1 Minerals
 TV = Vitric Minerals
 TO = Andic Minerals
 NA = Not applicable

 MIN_SUB = Subsoil mineralogy.
 KQ = 1/1 Minerals + Quartz
 KX = 1/1 Min. + Oxy. & Hydroxy.
 MK = 2/1 & 1/1 Minerals
 M = 2/1 & 2/1/1 non swel. Min.
 MS = Swel. & non swel. 2/1 Min.
 S = Swelling 2/1 Minerals
 TV = Vitric Minerals
 TO = Andic Minerals
 NA = Not applicable

 PD_TOP = Topsoil packing density.
 L = Low
 M = Medium
 H = High

 PD_SUB = Subsoil packing density.
 L = Low
 M = Medium
 H = High

 SLOPE-DOM: Dominant slope class of the STU

 0 No information
 1 Level (dominant slope ranging from 0 to 8 %)
 2 Sloping (dominant slope ranging from 8 to 15 %)
 3 Moderately steep (dominant slope ranging from 15 to 25 %)
 4 Steep (dominant slope over 25 %)

 SLOPE-SEC: Secondary Slope class of the STU

- 0 No information
- 1 Level (dominant slope ranging from 0 to 8 %)
- 2 Sloping (dominant slope ranging from 8 to 15 %)
- 3 Moderately steep (dominant slope ranging from 15 to 25 %)
- 4 Steep (dominant slope over 25 %)

 IL: Code for the presence of an impermeable layer within the soil profile of the STU

- 0 No information
- 1 No impermeable layer within 150 cm
- 2 Impermeable layer between 80 and 150 cm
- 3 Impermeable layer between 40 and 80 cm
- 4 Impermeable layer within 40 cm

 BS_SUB = Base saturation of the subsoil.

- H = High (> 50 %)
- L = Low (< 50 %)

 BS_TOP = Base saturation of the topsoil.

- H = High (> 75 %)
- M = Medium (50 - 75 %)
- L = Low (< 50 %)

 PEAT = Peat.

- N = No
- Y = Yes

 OC_TOP = Topsoil organic carbon content.

- H = High (> 6 %)
- M = Medium (2 - 6 %)
- L = Low (1 - 2 %)
- V = Very low (< 1 %)

 CLIMANR: Code for the grouped climate categories

- 1 Northern (climate number: 11, 12, 21, 22, 23, 31, 34, 35)
- 2 Atlantic (climate number: 32, 41, 42, 43, 431)
- 3 Sub-atlantic (climate number: 44, 51, 52, 58, 441)

- 4 Northern sub-continental (climate number: 33, 45, 48, 451, 481)
- 5 Mediterranean semi-arid (climate number: 46, 47, 461)
- 6 Southern sub-continental (climate number: 53, 54, 55, 541, 551, 571)
- 7 Mediterranean (climate number: 56, 57)
- 8 Temperate mountainous (climate number: 49)

 CORGRP_CAT: Code for the grouped CORINE 2006 categories

- 210 Agricultural areas (CORINE_06: 211, 212, 213, 241, 242, 243, 244)
- 220 Vineyard and fruit tree areas (CORINE_06: 221, 222, 223)
- 230 Meadow, pasture and natural grassland areas (CORINE_06: 231, 321, 322)
- 310 Forest (wood) areas (CORINE_06: 311, 312, 313)
- 320 Woody and scrub areas (CORINE_06: 321, 322)
- 410 Peat bog areas (CORINE_06: 412)

Estimated parameters and rules

1., DISR = Depth to the impermeable rock

S: < 40 cm

M: 40-80 cm

D: 80-120 cm

V: 120 cm <

DISR: ← PARMADO

← PARMASE

← DR

2., DG = Depth to the groundwater

S: 0-80 cm

M: 80-120 cm

D: 120-200 cm

V: 200 cm <

DG: ← WRBFU

3., MDSP = Maximum depth of soil profile with filter and storage properties

S: 0-20 cm

M: 0-60 cm

D: 0-100 cm

V: 0-150 cm

MDSP: ← DISR

← DG

4., MTDCH = Modified depth to a textural change code

0: no textural change

1: 30 cm

2: 40 cm

3: 50 cm

4: 60 cm

5: 70 cm

6: 80 cm

MTDCH: ← TEXTDEPCHG

← MDSP

5., CPT = Clay percentage of the topsoil

0: no information

1: ~ 10 %

2: ~ 20 %

3: ~ 30 %

4: ~ 50 %

5: ~ 80 %

9: ~ peat

CPT: ← TEXTSRFDOM

← TEXTSRFSEC

6., CPS = Clay percentage of the subsoil

0: no information

1: ~ 10 %

2: ~ 20 %

3: ~ 30 %

4: ~ 50 %

5: ~ 80 %

9: ~ peat

CPS: ← TEXTSUBDOM

← TEXTSUBSEC

7., MCPT = Modified clay percentage of the topsoil

0: peat

1: VL (very low)

2: L (low)

3: M (medium)

4: H (high)

5: VH (very high)

MCPT: ← CPT

← VS

← PD_TOP

8., MCPS = Modified clay percentage of the subsoil

0: peat

1: VL (very low)

2: L (low)

3: M (medium)

4: H (high)

5: VH (very high)

MCPT: ← CPS

← VS

← PD_TOP

9., SPCC = Clay content of the soil profile

(1 - 10); 1: low, 10: high

SPCC: ← MCPT

← MCPS

← MDSP

← MTDCH

10., HLT = Thickness of humus layer

1: thin

2: medium

3: thick

HLT: ← WRB_GRP

← MDSP

11., HQ = Humus quality code

F: fulvic acid

H: humic acid

T: peat

HQ: ← WRB_GRP

12., SPHC = Humus content of soil profile

(0 - 5); 1: low, 5: high

SPHC: ← PEAT

← OC_TOP

← CPT

← CPS

← HLT

← VS

← PD_TOP

13., WB = Water balance code

- 1: no leaching
- 2: low leaching
- 3: moderate leaching
- 4: high leaching
- 5: very high leaching

WB: ← CLIMANR

14., SLOPEM = Modified slope category

- 0: no information
- 1: 0-8 %
- 2: 8-15 %
- 3: 15-25 %
- 4: 25 % <

SLOPEM: ← SLOPE_DOM
← SLOPE_SEC

15., WBM = Modified water balance

- 0: no information
- 1: 0-8 %
- 2: 8-15 %
- 3: 15-25 %
- 4: 25 % <

WBM: ← SLOPEM
← CORGRP_CAT
← WB

16., TPD_f = Topsoil packing density factor

- 1: PD_TOP = L
- 0,8: PD_TOP = M
- 0,6: PD_TOP = H

TPD_f: ← PD_TOP

17., SPD_f = Subsoil packing density factor

- 1: PD_SUB = L
- 0,9: PD_SUB = M
- 0,8: PD_SUB = H

SPD_f: ← PD_SUB

18., IL_f = Impermeable layer factor

- 1: no impermeable layer
- 0,8: impermeable layer between 80-150 cm
- 0,6: impermeable layer between 40-80 cm
- 0,4: impermeable layer within 40 cm

IL_f: ← IL

19., IR = Infiltration rate
(1 - 10); 1: low, 10: high

IR: ← SPCC
← TPD_f
← SPD_f
← IL_f

20., LEACH_f = Leaching factor

1: WBM = 0 or 1

0,9: WBM = 2

0,8: WBM = 3

0,7: WBM = 4

0,6: WBM = 5

LEACH: ← WBM

21., IR_f = Infiltration rate factor

1: IR = 1

0,9: IR = 2

0,85: IR = 3

0,8: IR = 4

0,75: IR = 5

0,7: IR = 6

0,65: IR = 7

0,6: IR = 8

0,55: IR = 9

0,5: IR = 10

IR_f: ← IR

22., CEC_HUM = Cation exchange capacity related to humus content

(1 - 10); 1: low; 10: high

CEC_HUM: ← SPHC

← HQ

23., SSA_HUM = Specific surface area related to humus content

(1 - 10); 1: low; 10: high

SSA_HUM: ← SPHC

← HQ

24., CEC_MIN = Cation exchange capacity related to the soil minerals

(1 - 10); 1: low; 10: high

CEC_MIN: ← SPCC

← MIN_TOP

← MIN_SUB

25., SSA_MIN = Specific surface area related to soil minerals

(1 - 10); 1: low; 10: high

SSA_MIN: ← SPCC

← MIN_TOP

← MIN_SUB

26., CEC_SUM = Cation exchange capacity related to soil minerals and humus content

(1 - 10); 1: low; 10: high

CEC_SUM: ← CEC_MIN

← CEC_HUM

25., SSA_SUM = Specific surface area related to soil minerals and humus content

(1 - 10); 1: low; 10: high

SSA_SUM: ← SSA_MIN

← SSA_HUM

26., BASE_SAT = Base saturation of the soil profile

(1 - 10); 1: low; 10: high

BASE_SAT: ← BS_TOP

← BS_SUB

27., AER = Aeration code

1: DG = S

2: DG = M

3: DG = D

4: DG = V

AER: ← DG

28., CATS_f = Cation sorption factor

(0,1 - 1); 0,1: low; 1: high

CATS_f: ← BASE_SAT

← AER

29., ANS_f = Anion sorption factor

(0,1 - 1); 0,1: low; 1: high

ANS_f: ← BASE_SAT

← AER

30., DG_f = Groundwater depth factor

1: DG = S, M or D

2: DG = V

DG_f: ← DG

31., MIN_f = Mineral factor

1: all minerals, except KX (1/1 Min. + Oxy. & Hydroxy.)

1,5: KX (1/1 Min. + Oxy. & Hydroxy.)

32., STOR_CAPCA = Cation storing capacity (elements in cationic form and organic chemicals with positive charges)

(1-10; 1: poor; 10: good)

STOR_CAPCA: ← CEC_MIN

← CATS_f

← CEC_HUM

← LEACH_f

33., FILT_CAPCA = Cation filtering capacity (elements in cationic form and organic chemicals with positive charges)

(1-10; 1: poor; 10: good)

FILT_CAPCA: ← CEC_MIN

← CATS_f

← CEC_HUM

← IR_f

← DG_f

34., STOR_CAPAN = Anion storing capacity (elements in anionic form and organic chemicals with negative charges)

(1-10; 1: poor; 10: good)

STOR_CAPAN: ← SSA_MIN

← MIN_f

← SSA_HUM

← ANS_f

← LEACH_f

35., FILT_CAPAN = Anion filtering capacity (elements in anionic form and organic chemicals with negative charges) (1-10; 1: poor; 10: good)

FILT_CAPAN: ← SSA_MIN

← SSA_HUM

← ANS_f
← IR_f
← DG_f
← MIN_f

36., STOR_CAPSO = Solids and pathogenic microorganisms storing capacity
(1-10; 1: poor; 10: good)

STOR_CAPSO: ← SSA_SUM
← LEACH_f

37., FILT_CAPSO = Solids and pathogenic microorganisms filtering capacity
(1-10; 1: poor; 10: good)

FILT_CAPSO: ← SSA_SUM
← IR_f
← DG_f

38., STOR_CAPNP = Non-polar organic chemicals storing capacity
(1-10; 1: poor; 10: good)

STOR_CAPNP: ← SSA_SUM
← LEACH_f

39., FILT_CAPNP = Non-polar organic chemicals filtering capacity
(1-10; 1: poor; 10: good)

FILT_CAPNP: ← SSA_SUM
← IR_f
← DG_f

40., STOR_NAPL = Nonaqueous Phase Liquids (NAPL) storing capacity
(1-10; 1: poor; 10: good)

STOR_NAPL: ← SPCC
← SSA_SUM
← LEACH_f

41., FILT_NAPL = Nonaqueous Phase Liquids (NAPL) filtering capacity
(1-10; 1: poor; 10: good)

FILT_NAPL: ← SSA_SUM
← IR_f
← DG_f

List of figures

1.	The proposed filtration and storing capacity model	9
2.	Diagram for the evaluation of the soil storing and filtering functions	17
3.	Map of cation storing capacity	19
4.	Map of anion storing capacity	20
5.	Map of solids and pathogenic microorganisms storing capacity	21
6.	Map of non-polar organic chemicals storing capacity	22
7.	Map of nonaqueous Phase Liquids storing capacity	23
8.	Map of cation filtering capacity	24
9.	Map of anion filtering capacity	25
10.	Map of solids and pathogenic microorganisms filtering capacity	26
11.	Map of non-polar organic chemicals filtering capacity	27
12.	Map of nonaqueous Phase Liquids filtering capacity	28
13.	The Biogeographical Regions map of Europe	29
14.	Comparison of WCI_{norm} values of European Biogeographical Regions to evaluate the spatial pattern of storing capacity of different substances	30
15.	Comparison of WCI_{norm} values of European Biogeographical Regions to evaluate the spatial pattern of filtering capacity of different substances	31

***Europe Direct is a service to help you find answers
to your questions about the European Union.***

Freephone number (*):

00 800 6 7 8 9 10 11

(*) The information given is free, as are most calls (though some operators, phone boxes or hotels may charge you).

More information on the European Union is available on the internet (<http://europa.eu>).

HOW TO OBTAIN EU PUBLICATIONS

Free publications:

- one copy:
via EU Bookshop (<http://bookshop.europa.eu>);
- more than one copy or posters/maps:
from the European Union's representations (http://ec.europa.eu/represent_en.htm);
from the delegations in non-EU countries (http://eeas.europa.eu/delegations/index_en.htm);
by contacting the Europe Direct service (http://europa.eu/europedirect/index_en.htm) or
calling 00 800 6 7 8 9 10 11 (freephone number from anywhere in the EU) (*).

(*) The information given is free, as are most calls (though some operators, phone boxes or hotels may charge you).

Priced publications:

- via EU Bookshop (<http://bookshop.europa.eu>).

JRC Mission

As the science and knowledge service of the European Commission, the Joint Research Centre's mission is to support EU policies with independent evidence throughout the whole policy cycle.



EU Science Hub
ec.europa.eu/jrc



@EU_ScienceHub



EU Science Hub - Joint Research Centre



Joint Research Centre



EU Science Hub

doi:10.2788/49218

ISBN 978-92-79-64939-4

